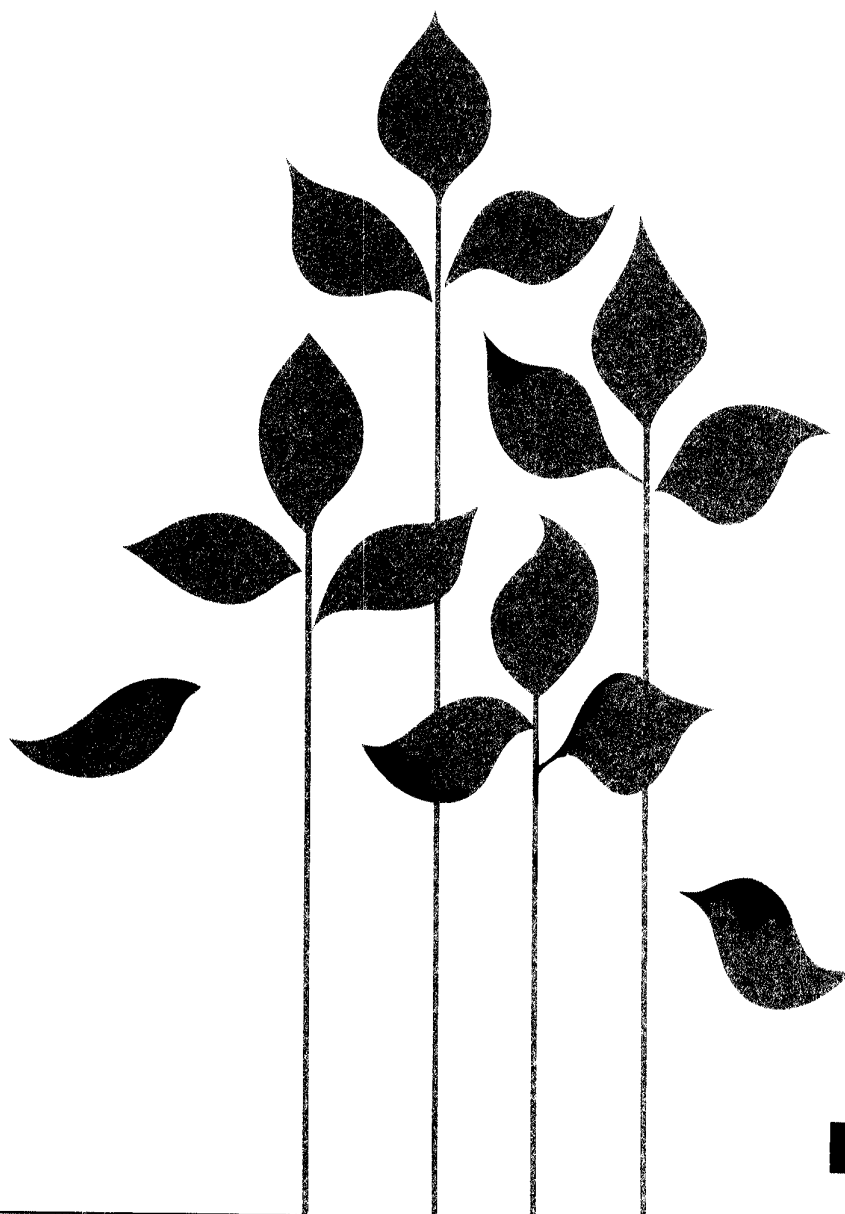


Canadian Plant Disease Survey

Volume 55 • Number 4 • 1975



Agriculture
Canada

Canadian Plant Disease Survey

Volume 55, Number 4, 1975

CPDSAS 55(4) 117-158 (1975) ISSN 0008-476X

Contents

- 117 Cooperative seed treatment trials - 1975
J.T. Mills
- 121 Prevalence of alfalfa crown and root diseases in the Peace River Region of Alberta and British Columbia
J.S. McKenzie and J.G.N. Davidson
- 126 Potato seed-piece decay: a bibliography, 1930-1975
J.P. Miska and G.A. Nelson
- 147 Resistance of turfgrasses to low-temperature-basidiomycete snow mold and recovery from damage
J. Drew Smith
- 155 Ergot tolerance in spring rye
Frank Sosulski and C.C. Bernier
- 158 Author index to volume 55

The *Canadian Plant Disease Survey* is a periodical of information and record on the occurrence and severity of plant diseases in Canada and on the assessment of losses from disease. Other original information such as the development of methods of investigation and control, including the evaluation of new materials, will also be accepted. Review papers and compilations of practical value to plant pathologists will be included from time to time.

Canadian Plant Disease Survey est un périodique d'information sur la fréquence des maladies des plantes au Canada, leur gravité, et les pertes qu'elles occasionnent. La rédaction accepte d'autres communications originales notamment sur la mise au point de nouvelles méthodes d'enquête et de lutte ainsi que sur l'évaluation des nouveaux produits. De temps à autre, il inclut des revues et des synthèses de rapports d'intérêt immédiat pour les phytopathologistes.

Research Branch, Agriculture Canada

Editor: W.L. Seaman, Research Station, Agriculture Canada, Ottawa, Ontario K1A 0C6

Editorial Board: R.A. Shoemaker, J.T. Slykhuis, C.D. McKeen, Chairman

Cooperative seed treatment trials - 1975¹

J.T. Mills

Twenty-four seed treatment chemicals were tested at two locations for their efficacy in controlling bunt of wheat [*Tilletia caries* and *T. foetida*], covered smut of oats [*Ustilago kollerii*], and covered smut of barley [*U. hordei*] and for their effects on the emergence of flax. Smut infection of untreated seed was low with the exception of 25% bunt at Brandon. Vitaflo 280 and 18 other treatments gave significantly reduced levels of bunt and of oat and barley smut at both stations. Flax emergence was significantly increased after treatment with CFG 3000. Bay-meb 6447 at the 5.00 and 10.00 g/kg and 26019 RP at the 1.50 g/kg rate reduced flax emergence.

Can. Plant Dis. Surv. 55:117-120. 1975

On a évalué à deux endroits différents l'efficacité de 24 produits de traitement des semences contre la carie du blé (*Tilletia caries* et *T. foetida*) et les charbons couverts de l'avoine (*Ustilago kollerii*) et de l'orge (*U. hordei*), ainsi que l'effet de ces produits sur la levée du lin. Les semences non traitées ont peu souffert du charbon, mais à Brandon 25% ont été attaquées par la carie. Le Vitaflo 280 et 18 autres traitements ont significativement réduit la fréquence de la carie et des charbons de l'avoine et de l'orge aux deux stations. Le traitement au CFG 3000 a fortement augmenté le taux de levée du lin, mais l'application de Bay-meb 6447 aux taux recommandés de 5.00 et 10.00 g/kg ou de 26019 RP à 1.50 g/kg l'a réduit.

In 1975, 24 seed treatment chemicals were tested for their efficacy in controlling common bunt of wheat [*Tilletia foetida* (Wallr.) Liro and *T. caries* (DC.) Tul.], covered smut of oats [*Ustilago kollerii* Wille], and covered smut of barley [*U. hordei* (Pers.) Lagerh.] and for their effects on the emergence of flax under Manitoba conditions.

Materials and methods

The chemical composition, where available, and the product name and source of the materials used are listed in Table 1. Vitaflo 280 was included as a comparison standard.

Seeds of 'Norteno M67' wheat (*Triticum aestivum* L.), 'Random' oats (*Avena sativa* L.), and 'Herta' barley (*Hordeum distichon* L.) were used in the smut tests. 'Raja' flax (*Linum usitatissimum* L.) was used for emergence tests.

Prior to chemical treatment the cereals were inoculated with the appropriate dry smut spores at the rate of 1 g per 200 g of wheat, oats, or barley seed. The chemical dosages used were those suggested by the manufacturer (Table 2). Each sample was hand-shaken in a glass jar to cover the seed uniformly with the chemical. After 3 days or more, 200 seeds were removed from each jar and placed in a paper envelope. Envelopes that contained seed of the same treatment were stored in polyethylene bags at 15°C for up to 4 weeks before seeding.

Tests were carried out at Brandon and duplicated at Morden, Manitoba. There were four replicates at each location. Each replicate consisted of 200 seeds planted in a row 4 m long; all rows were planted 25 cm apart; plots were arranged in a randomized block design. Emergence of flax was recorded 3-4 weeks after seeding.

At Brandon, wheat and oats were sown on May 12 and May 26 respectively and barley and flax on May 27; at Morden wheat and oats were sown on May 15 and May 22, and barley and flax on May 28.

The number of smutted heads in each row was recorded after the crop had headed and are expressed as means of the number of heads in the untreated rows. The results are given as means of four replicates, at each planting site. The "LSD-05" was determined from the means of the treatments at each station.

Results and discussion

Smut infection of untreated seed varied from 3% to 25% for wheat, from 4% to 8% for oats, and from 1% to 3% for barley.

The comparison standard Vitaflo 280 and 18 other treatments gave significantly reduced levels of bunt, and of oat and barley smut at both stations.

Phytotoxicity was observed on wheat at both stations after treatment with Bay-meb 6447 at the 5.00 and 10.00 g/kg rates.

Emergence of untreated flax varied from 48% to 59% (Table 2). Flax emergence was significantly increased

¹ Contribution No. 684, Research Station, Agriculture Canada, Winnipeg, Manitoba R3T 2M9.

with CFG 3000 at Morden and Brandon. Bay-meb 6447 at the 5.00 and 10.00 g/kg rates and 26019 RP at the 1.50 g/kg rate significantly reduced emergence at both stations.

Acknowledgments

The writer thanks members of the staff of the Morden and Brandon Research Stations for their cooperation and assistance.

Table 1. Seed treatment materials used in the cooperative tests

Treatment no.	Source*	Product name	Active ingredient
1		Untreated check	
2	Chemagro	Bay-meb 6447	1-(4-chlorophenoxy)-3,3-dimethyl-1H-1,2,4-triazol-1-yl)-2-butanone (25%) pyracarbolid 10% + maneb 16.7% maneb 25.0%
3	Chipman	TF 3262	identity not available
4	Chipman	TF 3309	identity not available
5	Ciba-Geigy	CFG 2980	identity not available
6	Ciba-Geigy	CFG 2990	identity not available
7	Ciba-Geigy	CFG 3000	identity not available
8	Ciba-Geigy	CFG 3010	identity not available
9	DuPont	DPX 12	identity not available
10	DuPont	DPX 14	identity not available
11	Interprovincial	Busan 25D	2-(thiocyanomethylthio) benzothiazole (25%)
12	Interprovincial	Busan 30L	2-(thiocyanomethylthio) benzothiazole (30%)
13	Interprovincial	BL 1794-30	identity not available
14	May & Baker	26019 RP	(1-(isopropylcarbonyl)-3-(3,5-dichlorophenyl) hydantoin)
15	Merck	Me 125	identity not available
16	Nor-Am	SN 43493	identity not available
17	Nor-Am	SN 49183	identity not available
18	Rohm & Haas	RHC 366	identity not available
19	Uniroyal	Vitaflo 280	Vitavax (carbathiin) 14.9% + thiram 13.2%
20	Uniroyal	UBI 2067	identity not available
21	Uniroyal	UBI 2078	identity not available
22	Uniroyal	UBI 2079	identity not available
23	Uniroyal	UBI 2080	identity not available
24	Uniroyal	UBI 2083	identity not available
25	Uniroyal	UBI 2085	identity not available
26		Untreated check	

*Chemagro Ltd., Mississauga, Ontario; Chipman Chemicals Ltd., Hamilton, Ontario; Ciba-Geigy Canada Ltd., Cambridge (Galt) Ontario; E.I. DuPont de Nemours & Co., Inc., Wilmington, Delaware; Interprovincial Cooperatives Ltd., Winnipeg, Manitoba; May & Baker (Canada) Ltd., Montréal, Québec; Merck Sharp and Dohme, Rahway, New Jersey; Nor-Am Agricultural Products Inc., Woodstock, Illinois; Rohm & Haas Co. of Canada Ltd., West Hill, Ontario; Uniroyal Chemical Division, Elmira, Ontario.

Table 2. Effects of seed-treatment chemicals on smuts in wheat, oats, and barley and emergence of flax

Treatment no.	Product name	Formulation*	Dosage (g or ml/kg)	Mean no. smutted heads†						Emergence‡‡	
				Wheat		Oats		Barley		Flax	
				B**	M**	B**	M**	B**	M**	B**	M**
1	Untreated check			38.75	10.25	20.50	15.25	14.25	1.00	101.00	111.25
2	Bay-meb 6447	WP	5.00	0	0	0	0	0	0	72.75	82.25
			10.00	0	0	0	0	0	0	51.25	69.50
3	TF 3262	SL	1.04	0.25	0.75						
			4.12			0	0				
			2.92					0	0		
4	TF 3309	SL	1.73	0	0						
			4.50			0.75	0				
			3.22					0	0		

Treatment no.	Product name	Formulation *	Dosage (g or ml/ kg)	Mean no. smutted heads†						Emergence††	
				Wheat		Oats		Barley		Flax	
				B **	M **	B **	M **	B **	M **	B **	M **
5	CFG 2980	WP	2.08 3.67 2.60 4.46	1.25	0	0	0.75	0	0	117.75	146.75
6	CFG 2990	WP	2.08 3.67 2.60 4.46	1.00	0	1.75	0.75	0.75	1.75	112.25	122.25
7	CFG 3000	WP	2.08 3.67 2.60 4.46	1.75	0	1.00	1.50	0	0	130.00	143.75
8	CFG 3010	WP	2.08 3.67 2.60 4.46	1.50	0	0	0.50	0	0	110.75	113.00
9	DPX 12	WP	2.08 3.70 2.60 2.25	0	0	0	0	0	0	111.00	112.75
10	DPX 14	WP	2.08 3.70 2.60 2.25	0	0.75	0	0	0	0	116.25	124.75
11	Busan 25D	D	2.10 3.67 2.60 4.45	2.00	1.50	0	0.50	0	0	112.50	125.00
12	Busan 30L	SN	0.80 1.37 0.97 0.83 1.10	0.25	0.50	0	0.50	0.25	0	101.75 102.25	112.50 109.00
13	BL 1794-30	SN	1.56 2.34 2.75 4.13 1.95 2.92 1.67 2.79	1.50 2.25	0.25 1.00	1.00 0.25	0.75 0	1.50 0	0 0	70.75 82.50 78.50 80.25	105.25 111.00 100.00 98.00
14	26019 RP	WP	1.50 2.00	0 0	0 0					80.25	98.00
15	Me 125	SN	0.66 1.32	11.00 17.75	1.75 2.50	9.50 16.00	13.50 10.75	8.00 10.25	2.75 6.50	80.00 80.25	104.00 99.25
16	SN 43493	WP	2.10 3.90 2.30	0	0	3.00	2.25	6.50	1.50	104.00	116.00
17	SN 49183	WP	2.10 3.90 2.30	0	1.50	0.75	1.75	0	0	119.50	137.50
18	RHC 366	SL	0.52 1.04 2.08 0.92 1.84 3.68 0.66 1.31 2.62 1.12 2.24	0.25 0 0	0.50 0 0	4.00 0.50 0	7.75 4.75 0.25	0 0 0	0.25 0 0	102.00 119.75	123.25 123.75

Table 2 (cont'd)

Treatment no.	Product name	Formulation*	Dosage (g or ml/kg)	Mean no. smutted heads†						Emergence††		
				Wheat		Oats		Barley		Flax		
				B**	M**	B**	M**	B**	M**	B**	M**	
19	Vitaflo 280	SL	4.48 1.82 3.22 2.28 3.90	0	1.00	0.25	0	0	0	120.50	128.25	
20	UBI 2067	WP	2.75 1.95			0	0	0	0	106.50	122.50	
21	UBI 2078	SL	1.56 2.76 1.95 3.34	0	0	0	0	0	0	114.75	104.00	
22	UBI 2079	SL	1.39 2.45 1.73 2.79	2.00	2.75	0	0.50	0	0	94.25	109.75	
23	UBI 2080	SL	0.72 1.27 0.90 1.54	1.50	1.00	0	0	0	0	105.50	115.50	
24	UBI 2083	SL	1.82 3.22 2.28 3.90	0.50	1.00	0	0	0	0	104.75	109.00	
25	UBI 2085	SL	3.22 2.28			0	0.25	0	0			
26	Untreated check			46.75	12.25	16.25	18.00	10.25	2.75	97.00	118.75	
LSD (0.05)				7.76	2.83	4.42	3.63	3.91	1.65	22.81	15.26	
Mean no. of heads				187	447	237	357	468	557			

*Formulation code: D = dust, SN = solution, SL = slurry, WP = wettable powder

**B = Brandon

M = Morden

†% smut = $\frac{\text{mean no. smutted heads}}{\text{mean no. of heads}} \times 100$

††Flax emergence based on mean of 4 reps each having 200 seeds planted.

Prevalence of alfalfa crown and root diseases in the Peace River Region of Alberta and British Columbia¹

J.S. McKenzie and J.G.N. Davidson

Following the 1973-74 winter, numerous alfalfa fields showed severe injury in the Peace River Region of Alberta and British Columbia. A survey of root and crown diseases was conducted in 40 2nd-year or older alfalfa (*Medicago sativa*) fields. The root/crown rot complex was divided into four principal types of symptoms: root rot, internal crown rot, external crown rot, and winter crown rot. Moderate or severe root rot occurred in 68% of the fields observed in the northern part of the region in Alberta, whereas in the remaining area only 24% of the fields had just a trace amount of root rot. Internal crown rot was prevalent throughout the survey area. External crown rot and winter crown rot were of minor significance. Twelve percent of the fields observed had no disease symptoms, 55% had only one type of symptom, and 33% had more than one.

Can. Plant Dis. Surv. 55:121-125. 1975

Un grand nombre de champs de luzerne de la région de la rivière de la Paix, en Alberta et en Colombie-Britannique, ont gravement souffert de l'hiver 1973-1974. On a conduit un relevé des types de pourridiés dans 40 luzernières (*Medicago sativa*) de deux ans ou plus. On a ainsi pu définir quatre types de pourridiés selon les symptômes notés: le pourridié des racines, le pourridié interne du collet, le pourridié externe du collet et le pourridié hivernal du collet. Dans le nord de la région, en Alberta, 68% des champs examinés étaient gravement ou moyennement atteints du pourridié des racines alors que 24% des champs restants seulement ne manifestaient qu'un taux d'infestation négligeable. Le type le plus fréquent de pourridié dans toute la région est le pourridié interne du collet. Le pourridié externe du collet et le pourridié hivernal sont relativement rares. Sur les champs examinés, 12% n'en présentaient pas de symptômes, 55% n'en présentaient qu'un cas et 33%, plus d'un.

Extensive damage to perennial grasses and legumes occurred during the 1973-74 winter in the Peace River Region of Alberta and British Columbia. This winter was characterized by unusually deep and prolonged snow cover. Severe injury has also been reported following the winters of 1955-56 (8), 1926-27 (2) and 1922-23 (1).

With the recent opening of five dehydration plants in this region, alfalfa (*Medicago sativa* L.) production has gained increasing importance. However, yields have been less than 1 ton per acre in some areas and an unsatisfactory level of winter survival has created additional difficulties for many growers. Hence a project was initiated to assess the extent and causes of poor winter survival and low yields in this region.

Diseases and injuries affecting the overwintering of alfalfa have been recently reviewed by Graham et al. (11), Jung and Larson (17), Kehr et al. (18), Leath et al. (19), Bolton (4), and Heinrichs (15). It has become increasingly apparent that crown rot and root deterioration is a product of an interaction of multiple factors of both the physical and biological environments. Knowledge of the pathogens and environmental factors involved is far from complete and symptoms produced by different causes overlap (19). Moreover, this problem

has not been previously investigated in this region and very seldom north of the 55th parallel (25). We therefore decided to assess and describe the types of symptoms, to determine the causal factors involved, and finally to arrive at solutions to minimize occurrence. This paper documents the first phase of this project, to identify the major types of damage occurring on alfalfa in the Peace River Region, as found in surveys of 40 2nd-year or older fields in 1974.

An attempt was made to determine major regional differences in frequency of symptom types. A strictly quantitative survey was not attempted. Twenty-eight alfalfa fields were surveyed throughout the region between June 16 and June 23, 1974. Seven fields were surveyed in the Beaverlodge area on May 23, and 5 fields in the Dawson Creek and Fort St. John area on August 15. Sampling was started 8-15 m (25-50 feet) from the edge of each field, and several plants were dug at each of 5 to 10 locations throughout the field. Except in the case of small patches of winter crown rot, above-ground conditions were not factors in choosing location. Roots and crowns were sliced vertically to show the type and severity of injury. Each field was rated as nil, trace, moderate, or severe for each disease symptom. The index was a qualitative evaluation of the total field sample. Fields tended to fall into one of four clear-cut groups with very little intergradation. Nil indicated that symptoms were not present. Trace indicated that early symptoms were present. Moderate indicated that some symptoms were well developed but either occurred

¹ Contribution No. NRG 75-17, Agriculture Canada, Research Station, Beaverlodge, Alberta T0H 0C0

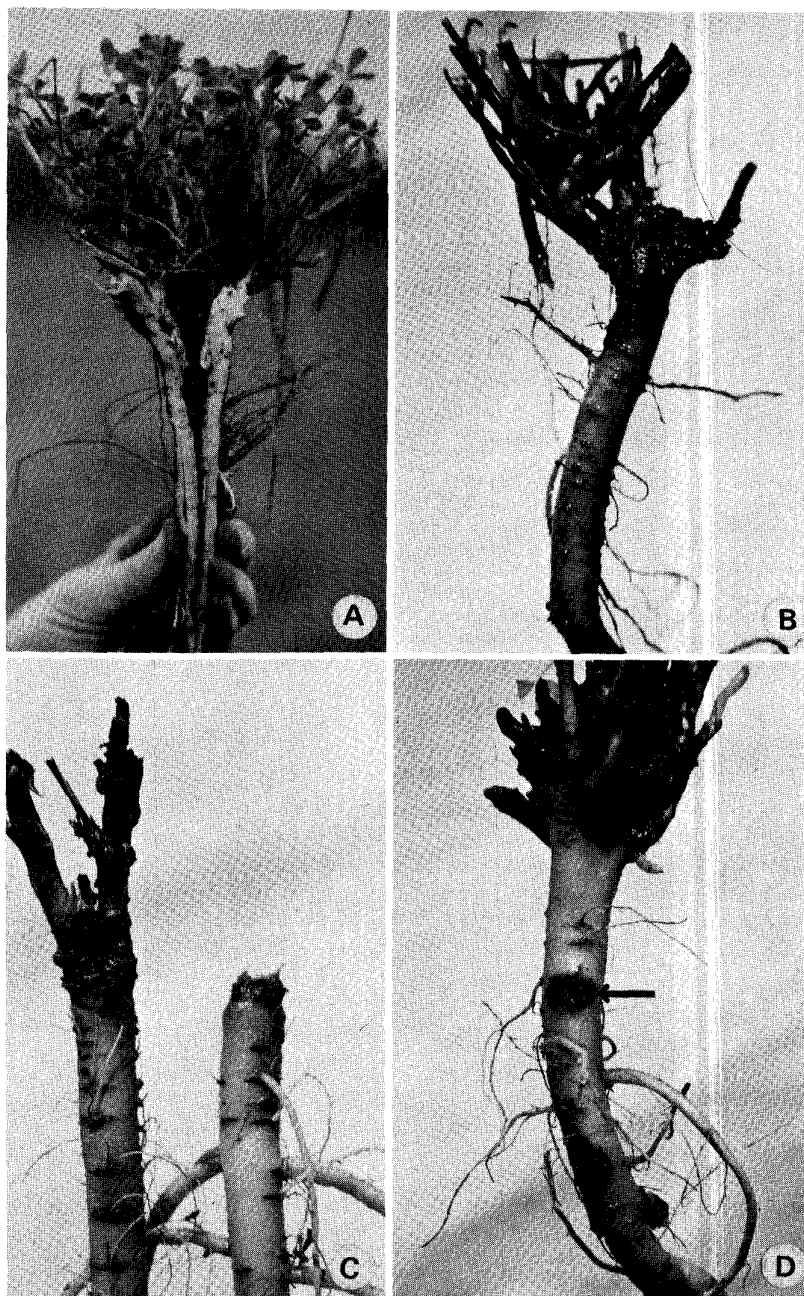


Figure 1. The four main types of crown and root disorders found in the Peace River Region in 1974.

- A) Internal crown rot symptoms, indicating advanced stage where central stele of root is damaged.
- B) External crown rot, showing necrosis of the root-crown cortex.
- C) Winter crown rot indicating disintegration of the crown tissue.
- D) Root rot (arrow) showing site of initiation where lateral roots emerge.

sporadically or appeared to cause no serious deterioration of the stand. Severe indicated that symptoms were

well developed throughout the field and caused obvious deterioration of the stand.

Results

Description of symptoms

Four main types of injury occurring singly or in various combinations were distinguished.

Internal crown rot occurs frequently in older stands. Dark brown necrotic areas are initiated in the crown at the site of cut stems or in axils. The necrosis proceeds into the crown to form a wedge-shaped decayed area. This eventually progresses into the central stele of the taproot (Fig. 1A). The vigor of plants is reduced because fewer shoots originate from the reduced crown surface. Normally crown buds originate in the fall then develop rapidly in early spring, but in severely affected crowns new shoots arise only from the lower part of the crown in late spring.

External crown rot injury is characterized by necrosis in the transition zone of the crown and root cortex. It appears to begin as an external lesion which expands and results in partial girdling and partial killing of the crown and crown buds (Fig. 1B). After 2 or more seasons, girdling is complete, resulting in death of most of the crown. A marked increase in the extent of external crown rot has recently been observed in patches in one intensively monitored alfalfa field in the fall, prior to freeze-up.

Winter crown rot is a characteristic of both old and young stands. It occurs primarily in patches within a field. Crown tissues become soft and brownish yellow in appearance, then disintegrate until the whole crown is rotted off (Fig. 1C). The rot is not selective of tissues within the crown, and taproots usually are not damaged. It appears to be more common in low, wet areas and completely kills the crown, apparently in a single winter. It is associated with snow mold.

Root rot injury is characterized by blackish-brown lesions appearing on root surfaces, usually expanding from where a lateral root emerges from a taproot (Fig. 1D) or a main root. These lesions progress until they completely rot through the root.

Survey

Figure 2 outlines the Peace River Region of Alberta and British Columbia. The region is divided into two areas on the basis of where root rot appeared to be of major significance. Area 1 includes the region around Manning, Alberta, north to the Fort Vermilion, High Level districts. Area 2 includes the remaining portion of the Peace River Region in Alberta and British Columbia.

The results of the 1974 survey are given in Table 1. Root rot occurred in 68% of the fields surveyed in Area 1 and in most of those fields damage was either moderate or severe. In Area 2 only 24% of the fields had just a trace amount of root rot. Internal crown rot was slightly more prevalent in Area 2 than in Area 1 (Fig. 2). Fifty-seven percent of the 19 fields examined in Area 1 and 67% of the 21 fields in Area 2 had internal crown rot. External crown rot occurred more frequently in Area 1, while winter crown rot was more frequent in Area 2.

Both external crown rot and winter crown rot were of minor occurrence in 1974. Twelve percent of the fields observed had no disease symptoms, 55% had only one type of symptom, and 33% had more than one.

A comparison of the fall samples with the spring samples is valid for all symptoms except winter crown rot. Since winter crown rot was of only minor occurrence throughout the survey, data from the fall sampling were included in the analysis.

Discussion

The four types of symptoms distinguished here do not indicate causal agents or the season in which initiation or development occurs. They resemble symptoms previously observed, and are attributed to various causes at various seasons.

Symptoms illustrated and described elsewhere that are similar to our internal crown rot type have been termed heart rot (27, 28), crown rot, or crown and root rot (4, 6, 19, 24). They have been variously attributed to environmental conditions (27), *Rhizoctonia solani* Kühn (4, 24), *Fusarium* spp. (6, 19), *Stagonospora meliloti* (Lasch) Petr. (9, 16); similar symptoms have also been produced by artificial freezing (28). The later stages of crown bud rot, which has been attributed to a complex including *R. solani*, *Fusarium roseum* Link sensu Snyder and Hansen [*F. avenaceum* (Fr.) Sacc., *F. acuminatum* (Ell. and Ev.) Wr.], and *Ascochyta imperfecta* Peck (*Phoma medicaginis* Malbr. & Roum.), are also similar (12, 13, 14).

External crown rot types of symptom have been termed collar rot (27), phloem injury (28), and winter crown rot in its earlier or partial stages (7). They were attributed to environmental conditions (27) and the low-temperature basidiomycete (7) and also were produced by artificial freezing (28). The greatest increase in external crown rot that we have observed, however, occurred in the fall prior to freezing or snow cover.

Winter crown rot (4, 5, 7), also termed snow mold (20), and the low-temperature basidiomycete disease (11), has so far been attributed exclusively to the low-temperature basidiomycete. Isolates of this organism are quite variable in morphology and physiology (20, 26) and may prove to be a complex of basidiomycetes. This type of symptom has also been produced by artificial freezing (28).

Similar root rot symptoms have been variously termed brown root rot attributed to *Plenodomus meliloti* Dearness and Sanford (22, 25); root or root and crown rot due to *R. solani* (11, 24), *Fusarium* spp. (6, 19), or *Phytophthora megasperma* Drechs. (*P. cryptogea* Pethy. and Laff.) (10, 21), although dissimilar symptoms were also described and shown for *P. megasperma* (11); and root canker due to *R. solani* (3, 4, 9, 23).

We have, therefore, been careful to separate symptoms from causes, and in this report the attempt has been made to distinguish the main types of symptoms that

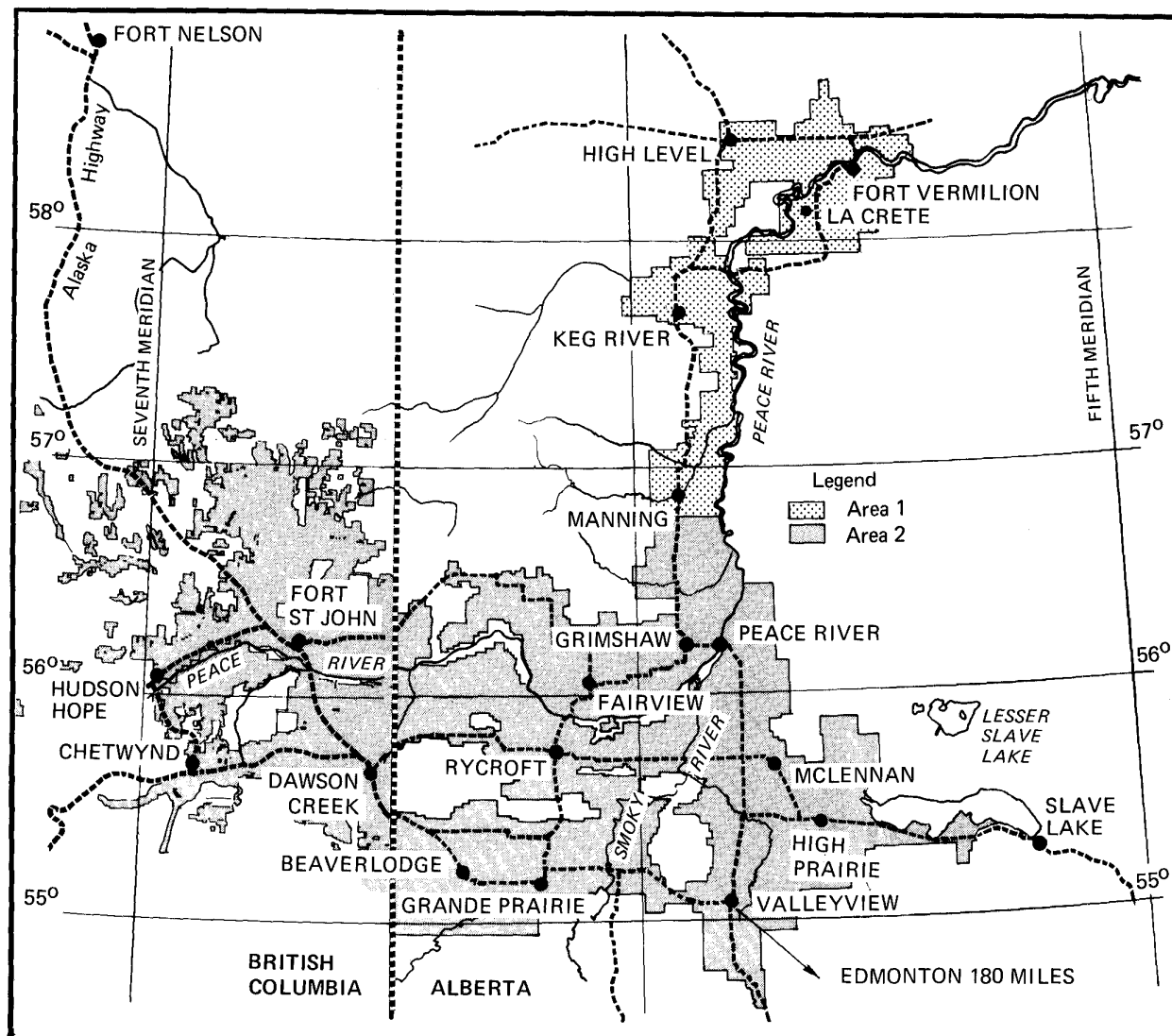


Figure 2. The agricultural area of the Peace River Region in Alberta and British Columbia. Area 1 extends from 10 miles south of Manning, Alberta, north to the High Level and Fort Vermilion area. Area 2 consists of the remaining parts of the Peace River Region.

occur in the Peace River Region before investigating causes and interactions. Descriptions of the symptoms of crown and root disorders of alfalfa and their relative distribution in the region were developed from the results of the survey for one season. The cultivars Beaver, Grimm, Rambler, and Roamer predominate in the region, but no attempt was made to identify cultivars or to specify the particular soil type in which a disorder was found. The prevalence of root rot in Area 1 and its minor significance in Area 2 may be important in understanding the disease and environmental relationships in these areas. Studies are continuing to identify the causes of these symptoms, the organisms involved, and to assess management practices that may reduce severity.

Acknowledgments

The technical assistance of Gordon McLean and Colin Brodie is gratefully acknowledged.

Literature cited

1. Albright, W. D. 1925. Report of the Superintendent for the year 1923. Dominion Exp. Sub-Sta., Beaverlodge, Alberta. Can. Dep. Agr., Dominion Exp. Farm, Ottawa, 101 pp.
2. Albright, W. D. 1928. Report of the Superintendent for the year 1927. Dominion Exp. Sub-Sta., Beaverlodge, Alberta. Can. Dep. Agr., Dominion Exp. Farm, Ottawa, 64 pp.
3. Baker, K. F. 1970. Types of Rhizoctonia diseases and their occurrence. Pages 125-148 in J. R. Parmeter, Jr., ed., *Rhizoctonia solani: biology and pathology*. Univ. Calif. Press, Berkeley, Calif., 225 pp.
4. Bolton, J. L. 1962. *Alfalfa: botany, cultivation, and utilization*. Leonard Hill, London. 473 pp.

Table 1. Percentage of alfalfa fields observed to have disease symptoms† in two areas of the Peace River Region of Alberta and British Columbia in 1974

Area*	No. fields observed	Internal crown rot			External crown rot			Winter crown rot			Root rot		
		T	M	S	T	M	S	T	M	S	T	M	S
1	19	32	15	10	11	5	0	0	5	0	0	53	15
2	21	33	24	10	5	0	0	5	5	0	24	0	0

* See Figure 2.

† T = trace; M = moderate; S = severe.

5. Broadfoot, W. C., and M. W. Cormack. 1941. A low-temperature basidiomycete causing early spring killing of grasses and legumes in Alberta. *Phytopathology* 31:1058-1059.
6. Cormack, M. W. 1937. *Fusarium* spp. as root parasites of alfalfa and sweet clover in Alberta. *Can. J. Res., C.*, 15:493-509.
7. Cormack, M. W. 1948. Winter crown rot or snow mold of alfalfa, clovers, and grasses in Alberta. *Can. J. Res., C.*, 26:71-85.
8. Elliott, C. R. 1957. Winter killing in alfalfa. *Rep. West. Forage Crops Conf.* June 21, 22. Agassiz, B.C. p.26.
9. Erwin, D. C. 1954. Relation of *Stagonospora*, *Rhizoctonia*, and associated fungi to crown rot in alfalfa. *Phytopathology* 44:137-144.
10. Erwin, D. C. 1954. Root rot of alfalfa caused by *Phytophthora cryptogea*. *Phytopathology* 44:700-704.
11. Graham, J. H., K. W. Kreitlow, and L. R. Faulkner. 1972. Diseases. Pages 497-526 in C. H. Hanson ed., *Alfalfa science and technology*. Amer. Soc. Agron. Agron. Ser. 15.
12. Hawn, E. J. 1958. Studies on the epidemiology of crown bud rot of alfalfa in southern Alberta. *Can. J. Bot.* 36:239-250.
13. Hawn, E. J. 1959. Histological study of crown bud rot in alfalfa. *Can. J. Bot.* 37:1247-1249.
14. Hawn, E. J., and M. W. Cormack. 1952. Crown bud rot in alfalfa. *Phytopathology* 42:510-511.
15. Heinrichs, D. H. 1969. Alfalfa in Canada. *Can. Dep. Agr. Publ.* 1377. 28 pp.
16. Jones, F. R., and J. L. Weimer. 1938. *Stagonospora* leaf spot and root rot of forage legumes. *J. Agr. Res.* 57:791-812.
17. Jung, G. A., and K. L. Larson. 1972. Cold, drought and heat tolerance. Pages 185-209 in C. H. Hanson, ed., *Alfalfa science and technology*. Amer. Soc. Agron. Agron. Ser. 15.
18. Kehr, W. R., F. I. Frosheiser, R. D. Wilcoxson, and D. K. Barnes. 1972. Breeding for disease resistance. Pages 335-354 in C. H. Hanson, ed., *Alfalfa science and technology*. Amer. Soc. Agron., Agron. Ser. 15.
19. Leath, K. T., F. L. Lukezic, H. W. Crittenden, E. S. Elliott, P. M. Halisky, F. L. Howard, and S. A. Ostazeski. 1971. The *Fusarium* root rot complex of selected forage legumes in the northeast. *Penn. State Univ. Bull.* 777. 66 pp.
20. Lebeau, J. B., and M. W. Cormack. 1961. Development and nature of snowmold damage in Western Canada. Pages 544-549 in *Recent Advances in Bot.*, Sec. 5, Univ. Toronto Press.
21. Marks, G. C., and J. E. Mitchell. 1971. Penetration and infection of alfalfa roots by *Phytophthora megasperma* and the pathological anatomy of infected roots. *Can. J. Bot.* 49:63-67.
22. Sanford, G. B. 1933. A root rot of sweet clover and related crops caused by *Plenodomus meliloti* Dearness and Sanford. *Can. J. Res.* 8:337-348.
23. Smith, O. F. 1943. *Rhizoctonia* root canker of alfalfa (*Medicago sativa*). *Phytopathology* 33:1081-1085.
24. Stanford, E. H., E. L. Jones, V. P. Osterli, B. R. Houston, R. F. Smith, and A. D. Reed. 1954. Alfalfa production in California. *Calif. Agr. Exp. Sta. Circ.* 442. 44 pp.
25. Tsukamoto, J. Y. 1965. Phenotypic characteristics of alfalfa tolerant to brown root rot. *Can. J. Plant Sci.* 45:197-198.
26. Ward, E. W. B., J. B. Lebeau, and M. W. Cormack. 1961. Groupings of isolates of a low-temperature basidiomycete on the basis of cultural behavior and pathogenicity. *Can. J. Bot.* 39:297-306.
27. Weimer, J. L. 1927. Observations on some alfalfa root troubles. *U. S. Dep. Agr. Circ.* 425. 10 pp.
28. Weimer, J. L. 1930. Alfalfa root injuries resulting from freezing. *J. Agr. Res.* 40:121-143.

Potato seed-piece decay: a bibliography, 1930–1975

Compiled by J. P. Miska and G. A. Nelson¹

Can. Plant Dis. Surv. 55:126-146. 1975

Contents

Introduction 127

Bibliographies 127

Erwinia 127

Biology
CO₂ accumulation associated with
Control
Losses caused by
Pathogenicity of
Soil as source of infection

Erwinia atroseptica 127–131

Apparatus for soil moisture
Biochemistry and temperature relations
Black leg in relation to
Chemical control
Control
Detection of
Effect of, on plant tissues
Effect of, on yield
Effect of crop rotation
Effect of overhead irrigation
Effect of sprinkler irrigation
Factors affecting
Incidence of
Inhibition of
Method for testing resistance to
Method of diagnosing
Mode of entry and spread
Mode of infection
Effect of succinic acid
Overwintering
Proteinase activity
Selective isolation of
Serological diagnosis and symptoms of
Spread of
Survival of, in soil
Technique for differentiation of, from saprophytes
Technique for evaluating resistance of
Testing chemicals against
Tuber infection
Ultrastructure
Varietal reaction to

Erwinia carotovora 131–133

Assessing infection
Control
Decomposition of pectic substances
Defence reaction
Effect of, on yield
Effect of EDTA
Effect of gamma radiation on
Effect of ionizing radiation on
Effect of light on
Effect of red light on

Effect of streptomycin on
Factors affecting
Fungi-toxicity of aerosols
Interaction with *Gibberella pulicaris*
Medium for detecting
Enzyme activity
Tissue extracts
Phenolic compounds in relation to
Ring-rot symptoms produced by
Serology of
Technique for isolating
Ultrastructure
Varietal reaction to

Erwinia phytophthora 133

Erwinia solani 133

Fusarium 133–136

Chemical control
Control
Effect of crop rotation
Effect of fungicides on
Effect of gamma radiation on
Effect of gibberellic acid and gibberellin derivatives on
Factors affecting
Fungicides ineffective against
Legislation against
Pathogenicity of
Resistance
Varietal reaction to

Fusarium coeruleum 136–138

Breeding against
Chemical control
Control
Effect of, on yield
Effect of disinfecting and bruising on
Factors affecting
Nutritional study of
Pathogenicity of
Phenol metabolism of
Physiology of
Susceptibility to
Testing resistance
Transmission of, by soil
Varietal reaction to

Fusarium culmorum 138

Fusarium oxysporum 139

Fusarium sambucinum f.6 (see *F. sulphureum*)

Fusarium solani 139

Fusarium sporotrichioides 139

Fusarium sulphureum 139

Storage diseases 139

Addendum 139–142

Author index 142–146

¹ Research Station, Agriculture Canada, Lethbridge, Alberta T1J 4B1

This world bibliography is intended to provide information to research workers on potato seed-piece decay. It lists scientific and technical papers published in periodicals relating to plant pathology and mycology. The compilation was developed from a card index maintained by Dr. G. A. Nelson and has been enlarged to include titles cited since 1930 in journals and by potato disease abstracting services.

Citations are arranged first by subject heading, then alphabetically by author. All titles are given in English. The entries are numbered consecutively and each gives author, date, title, source, and language of publication. Citations appearing under more than one subject heading are cross referenced and an author index appears at the end of the bibliography. The subject headings are worded according to *Review of Applied Mycology*, now *Review of Plant Pathology*. Entries in the Addendum are chiefly of the 1970's and are listed alphabetically by author; they are cited by numbers (prefix A) at the end of the appropriate subject heading.

The potato is one of our most important vegetable crops. A bibliography on decay of potato seed pieces will be a valuable source for scientists who wish to study seed-piece decay further.

Not all the titles in this bibliography are available in this library. For further information please contact Librarian, Research Station, Agriculture Canada, Lethbridge, Alberta, Canada T1J 4B1

Bibliographies

- 1 Miska, J. P. 1973. Agriculture 1906 - 1972; a bibliography of research. Lethbridge Res. Sta., Can. Dep. Agr. 192 p.
- 1a Montaldo, A. 1969. Latin American bibliography on potatoes. Univ. Cent. Venez. 177 p.
- 2 Review of Applied Mycology. 1-48 (1922-69). From vol. 49 (1970) superseded by Review of Plant Pathology.
- 3 Review of Plant Pathology. 49- (1970-) vols. 1-48 (1922 - 1969) preceded by Review of Applied Mycology.
- 4 U.S. Department of Agriculture. Agricultural Research Service. 1970. Bibliography of potato diseases through 1945: with common and scientific names. Misc. Pub. 1152. 243 p.

Erwinia

In the soft rot phase of seed-piece decay, the predominant incitant is *Erwinia atroseptica*. However *E. carotovora* has been implicated in this type of decay. The older binomial *E. phytophthora* is synonymous with *E. atroseptica*.

- 5 Scottish Horticultural Research Institute. 1968. Annual report, 5th. 63 p.
 - 6 Stone, W. J. H. 1966. A highly virulent *Erwinia* isolate from Arizona vegetables. Plant Dis. Rep. 50:414-418.
- See also 280, 349, 530, A12, A18, A21, A24, A27, A36, A40, A48, A50, A60, A68 A78, A79, A97, A102, A103.

Erwinia, Biology

- 7 European Association for Potato Research. Triennial Conference, 4th, Brest, 8-13 Sept., 1969., 1970. Proceedings. Wageningen. 298 p.
- 8 Pérombelon, M. C. M. 1970. The biology of contamination of the potato tuber by soft rotting *Erwinia* spp. Pages 196-197 in Proc. Fourth Trienn. Conf. Eur. Assoc. Potato Res. Brest, 8-13 Sept., 1969. Wageningen. 298 p.

Erwinia, CO₂ accumulation associated with

- 9 Nielsen, L. W. 1968. Accumulation of respiratory CO₂ around potato tubers in relation to bacterial soft rot. Amer. Potato J. 45:174-181.

Erwinia, control

- 10 Pérombelon, M. C. M. 1970. Preliminary implications for potato growing of recent research on bacterial soft rot / black leg complex of potatoes. Page 131 in Proc. Fourth Trienn. Conf. Eur. Assoc. Potato Res. Brest, 8-13 Sept., 1969. Wageningen. 298 p.
- 11 Singh, R. S., and M. M. Joshi, 1969. Antifungal and antibacterial activity of methyl arsenic sulphide and 4-benzozquinone N-benzol hydrazome oxime formulations. Pesticides 3(9): 19-21.

Erwinia, Losses caused by

- 12 Hughes, I. K. 1961. Potato diseases in South-east Queensland. Queensl. Agr. J. 87: 607-618.

Erwinia, Pathogenicity of

- 13 Gehring, F. 1962. Studies on the course of infection of carnation bacteriosis caused by *Pectobacterium parthenii* var. *dianthicola* (Starr)Helmers and on the enzymatic properties of this bacterium in comparison with *Pseudomonas caryophylli* and some typical wet rot agents [in German, English summary]. Phytopathol. Z. 43:383-407.

Erwinia, Soil as source of infection

- 14 Anon. 1970. Plant pathology. Rep. Agr. Res. Coun. U.K. 1969 - 1970 pp. 17-20.

Erwinia atroseptica (See also *E. phytophthora*)

- 15 Abdel-Rékhim, M.A. 1967. Black leg of potatoes in the U.A.R. [in Russian]. Biol. Nauk. 10(4): 114-116.
- 16 Altman, J. 1958. Studies on the control of plant diseases with antibiotics, with particular reference to streptomycin. Diss. Abstr. 19(2): 201.
- 17 Anon. 1971. Black leg of potatoes. Advis. Leaflet. Min. Agr. Fish., London, 107. 4 p.
- 17a Australia. Biological Branch, Dep. of Agric. 1936. Potato diseases of Victoria. J. Agric. Victoria 34:464-481.
- 18 Australia. Victorian Plant Research Institute. 1970. Reports, No. 3 July 1962 - Dec. 1965; No. 4, Jan. 1966 - Dec. 1967. 99 p.
- 19 Bates, G. R. 1960. Report of the Ministry of Agriculture of Rhodesia, 1958 - 1959. pp. 43-51.
- 20 Bétencourt, A., and J. P. Prunier. 1965. Concerning lenticel dry rot of potato tubers caused by *Erwinia carotovora* [in French, English summary]. Eur. Potato J. 8(4):230-242.
- 21 Blair, I.D. 1937. Deterioration in the potato. Bull. Canterbury Agr. Coll., Lincoln, 94. 2 p.
- 22 Blotskaya, Zh. V. 1972. Comparative evaluation of methods for diagnosing potato virus [in Russian]. Zashch. Rast., Moscow, 17(2):18.
- 23 Bobes, I., O. Pall, E. Perseca, D. Musat, and N. Florea. 1971. Studies on the frequency of parasitic micro-organisms on some stored agricultural foodstuffs [in Romanian, English summary]. Microbiologia, Bucarest, 2:141-147.
- 24 Boerema, H., H. A. Van Kesteren, M. M. J. Dorenbosch, and E. De Weert. 1971. Report on diseases new to the Netherlands. Pages 47-50 in Jaarb., 1967 - 1968 [in Dutch]. Versl. Meded. Plziektenk. Dienst. Wageningen, 145. 142 p.
- 25 Bonde, R. 1939. Comparative studies of the bacteria associated with potato blackleg and seed-piece decay. Phytopathology 29:831-851.
- 26 Bonde, R. 1939. The role of insects in the dissemination of potato blackleg and seed-piece decay. J. Agr. Res. 59:889-917.
- 27 Bourke, P.M. 1966. Contribution to the early history of the black leg disease of the potato. J. Dep. Agr. Repub. Ir. 63:103-109.

Erwinia atroseptica (contd.)

- 28 Clayton, E. E. 1929. Potato seed treatment experiments on Long Island with special reference to the organic mercury instant dips. New York Agr. Exp. Sta. 564. 32 p.
- 29 Davidsson, I. 1951. Investigations on crop diseases [in Icelandic, English summary]. Reykjavik, Ríkisprentsmíðjan Gutenberg. 21 p.
- 30 Delgado-Sanchez, S., L. Fucikovsky, and M. Cadena-Hinojosa. 1969. The occurrence of some previously unreported diseases of potatoes in Mexico. Plant Dis. Rep. 53:189-190.
- 31 Dembskaya, L. 1959. Resistance in the potato to diseases [in Russian]. S.-Kh. Sibiri 11:63-64.
- 32 Deveza, M. C. 1969. Diseases and pests of potato [in Spanish]. Publ. Serv. Agr. Mocamb. Ser. C. Separ. 53, 29 p.
- 33 Dobretsov, A. N. 1963. A short survey of bacterial diseases of agricultural crops in the Kansk forest steppes [in Russian]. Tr. Krasnoyarsk. Nauch. Issled. Inst. Sel'. Khoz. 2:165-169.
- 34 Epps, W. M. 1957. Control of potato seed piece decay in South Carolina 1952 - 1956. Plant Dis. Rep. 41:148-150.
- 35 Ficke, W. 1972. Model experiments on the longevity of *Pectobacterium carotovorum* var. *atrosepticum* (van Hall) Dowson [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 26(3):47-50.
- 36 Fredricks, A. L., and H. M. Metcalf. 1970. Potato blackleg disease. Amer. Potato J. 47:337-343.
- 37 Graham, D. C. 1958. Occurrence of soft rot bacteria in Scottish soils. Nature 181 (4601): 61.
- 38 Hellmers, E., and W. J. Dowson. 1953. Further investigations of potato black leg. Acta Agr. Scand. 3(1):103-112.
- 39 Henniger, H., B. Pett, W. Bartel, and M. Scholz. 1972. The effect of carbon dioxide levels in the air on black leg infection *Pectobacterium carotovorum* var. *atrosepticum* on potato tubers [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 26(6):112-116.
- 40 Hidalgo, O. A., L. W. Nielsen, and E. R. French. 1972. Black leg *Erwinia atroseptica* of potato in Peru [in Spanish]. Fitopatologia 4(1/2):15-18.
- 41 Huguelet, J., and D. C. Nelsen. 1971. The influence of cultural practices on the incidence of seedpiece decay, blackleg and *Rhizoctonia* disease. Amer. Potato J. 48:306. (Abstr.)
- 42 Jetne, M. 1955. Stem rot in the potato field [in Norwegian]. Norsk Landbr. 21(3):51-52.
- 43 Klapp, E., G. Morgenweck, and F. Spenneman. 1936. On the influence of the locality on yield and seed value of the potato. Investigations in 21 five - year progeny series [in German]. Landwirt. Jahrb. 83(2):153-207.
- 44 Klemm, M., G. Masurat, and S. Stephan. 1957. The occurrence of the most important diseases and pests of cultivated plants in the year 1953 in the zone of the German Democratic Republic [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 11(5):81-104.
- 45 Lepik, E. 1937. An internal rot of the potato caused by *Sclerotinia sclerotiorum* [in German]. Phytopathol. Z. 10:234.
- 46 Line, R. F., and C. J. Eide. 1960. Control of potato seed-piece decay. Plant Dis. Rep. 44:698-701.
- 47 Maas-Geesteranus, H. P. 1972. Contribution to the knowledge of potato blackleg [in German]. Beitrag Kennt. Schwarzb. Kartoff. pp. 151-156.
- 48 Maine. Agricultural Experiment Station. 1950. Report of progress, Ex 66th, June 30, 1950. Maine Agr. Exp. Sta. Bull. 483:41-61.
- 49 Malcolmson, J. F. 1959. A study of *Erwinia* isolates obtained from soft rots and blackleg of potatoes. Trans. Brit. Mycol. Soc. 42:261-269.
- 50 Masurat, G., R. Peschel, and S. Stephan. 1967. The occurrence of the most important diseases and pests of agricultural and horticultural plants in 1966 in the German Democratic Republic [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 21:137-168.
- 51 Masurat, G., and S. Stephan. 1959. The occurrence of the most important diseases and pests of crop and garden plants in the year 1957 in the zone of the German Democratic Republic [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 13(4):61-74.
- 52 Masurat, G., and S. Stephan. 1960. The occurrence of the most important diseases and pests of crop and garden plants in 1958 and 1959 in the German Democratic Republic [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 14(8):141-178.
- 53 Milheiro, A. V. 1967. Influence of the time of planting on the incidence of potato black leg [in Portuguese, English summary]. Rev. Agr. Mocambique, 9(87):40-41.
- 53a The Netherlands. Instituut voor Plantenziektenkundig Onderzoek. Annual report of the Institute for Phytopathological Research, Wageningen, in 1961 [in Dutch, English summary]. 188 p.
- 54 New South Wales. Plant disease conference held at Hawkesbury Agricultural College, June - July, 1955. Vol. 1. 308 p. 1955.
- 55 Nigeria. Ministry of Agriculture. Research and Specialist Services. 1962. Annual report, 1961 - 1962. 53 p.
- 56 Noble, M. and M. Marshall. 1952. A note on black leg of potato. Plant Pathol. 1:134.
- 57 N. Ireland. Ministry of Agriculture. Plant Pathology Division. 1964 Report, 1962. 12:229-246.
- 58 N. Ireland. Ministry of Agriculture. Plant Pathology Division. 1965. Report, 1964. 14: 91: 109.
- 59 Pérombelon, M. C. M. 1972. The extent and survival of contamination of potato stocks in Scotland by *Erwinia carotovora* var. *carotovora* and *E. carotovora* var. *atroseptica*. Ann. Appl. Biol. 71:111-117.
- 60 Pett, B. 1970. On resistance against drying of *Erwinia atroseptica* (pathogen of potato black leg and soft rot)[in German, English summary]. Zentrbl. Bakteriell. Parasitenk. Kde. Abt. 2, 125(3):322-325.
- 61 Pett, B., and H. Henniger. 1972. The presence of pathogenic agents of bacterial blackleg (*Pectobacterium carotovorum* var. *atrosepticum* (van Hall) Dowson) in potato tubers with dry rot [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 26(6):117-118.
- 62 Phillips, D. H. 1959. Report of the Mycological Department, 1958. Rep. States Jersey. U.K. pp. 45-62.
- 64 Ragozina, I. I., Yu. I. Shneider, and D. V. Lipsits. 1969. Pectolytic enzymes in *Erwinia atroseptica* culture and infected potato tissue [in Russian]. Dokl. Akad. Nauk SSSR 188(4):937-939.
- 65 Rhodesia. Ministry of Agriculture. Botany and Plant Pathology. Nyasaland. 1956. Report of the Minister. 1955 - 1956. pp. 66-70.
- 66 Robbs, C. F. 1960. Studies on the diseases "black leg" and "soft rot" of potato [in Portuguese]. Rev. Agr. Piracic. 35:91-95.
- 67 Robinson, D. B., G. W. Ayers, and J. E. Campbell. 1960. Chemical control of blackleg, dry rot, and verticillium wilt of potato. Amer. Potato J. 37:203-212.
- 68 Ruehle, G. D. 1940. Bacterial soft rot of potatoes in southern Florida. Fla. Agr. Exp. Sta. Tech. Bull. 348. 36 p.
- 69 Ruschmann, G. 1937. Farmyard and fermented manure effects [in German]. Landwirt. Jahrb. 84(2): 263-278.
- 70 Sabet, K. A. 1955. Non-sporing bacteria responsible for soft-rots and related diseases of vegetables. Proc. Egypt. Acad. Sci. 10, 5 p.
- 71 Salzmann, R. 1963. Report on the work of the Federal Agricultural Experiment Station Zürich - Oerlikon for the years 1960, 1961, and 1962 [in German]. Landwirt. Jahrb. Schweiz. 74, ed. Fr. 64:193-290.
- 72 Smith, W. L. 1950. Pathogenic differences manifested by *Erwinia atroseptica* and *Erwinia carotovora*. Phytopathology 40:1011-1017.

Erwinia atroseptica (contd.)

- 73 Staples, R. R. 1958. Report of the Department of Research and Specialist Services (Southern Rhodesia) for the year ended 30th September, 1957. Pages 7-86 in Rep. Min. Agr. Rhod. Nyasaland, 1956 - 1957.
- 74 Torres, H., E. R. French, and L. W. Nielsen. 1970. Potato diseases in Peru. Plant Dis. Rep. 54:315-318.
- 75 Tranina, N. F. 1960. The effect of soil cultivation on the root microflora of some plants [in Russian]. Tr. Inst. Mikrobiol. 7:107-114.
- 76 Wade, G. C. 1954. Potato diseases in Tasmania. Tasmanian J. Agr. 25:240-252.
See also 153, 154, 172, 204, 229, 286, 287, A1, A2, A3, A7, A8, A17, A26, A29, A30, A31, A32, A33, A34, A35, A38, A39, A41, A42, A46, A47, A49, A54, A55, A56, A60, A61, A62, A64, A65, A66, A69, A73, A74, A77, A78, A79, A81, A82, A85, A89, A93, A94, A97, A100, A101, A104, A106, A107, A108, A109.

Erwinia atroseptica, Apparatus for soil moistures

- 78 Read, D. C., G. W. Ayers, J. E. Campbell, and D. B. Robinson. 1961. Note on apparatus for maintaining constant soil temperatures and soil moistures, used in studies on soil inhabiting insects and diseases. Can. J. Plant Sci. 41:876-879.

Erwinia atroseptica, Biochemistry and temperature relations

- 79 Graham, D. C., and W. J. Dowson. 1960. The coliform bacteria associated with potato black-leg and other soft rots. I. Their pathogenicity in relation to temperature. II. Biochemical characteristics of low - and high - temperature strains. Ann. Appl. Biol. 48:51-57.

Erwinia atroseptica, Black leg in relation to

- 80 Scottish Horticultural Research Institute. 1971. Annual report, 7th. 69 p.

Erwinia atroseptica, Chemical control

- 81 Bonde, R. 1953. Preliminary studies on the control of bacterial decay of the potato with antibiotics. Amer. Potato J. 30:143-147.
- 82 Bonde, R. 1955. Antibiotic treatment of seed potatoes in relation to seed-piece decay, blackleg, plant growth, and yield rate. Plant Dis. Rep. 39:120-123.
- 83 Bonde, R. 1955. Further studies on the control of bacterial decay of potato seed pieces with antibiotics. Amer. Potato J. 32:387. (Abstr.)
- 84 Bonde, R., and P. de Souza. 1954. Studies on the control of potato bacterial seed-piece decay and blackleg with antibiotics. Amer. Potato J. 31:311-316.
- 85 Duncan, H. E., and M. E. Gallegly. 1962. Field trials for chemical control of potato seedpiece decay and blackleg. Phytopathology 52:164. (Abstr.)

Erwinia atroseptica, Control

- 86 Binilauskaitė, I. 1962. Spread of 'black leg' of potato and some measures for its control in the conditions of the Lithuanian S.S.R. [in Lithuanian, Russian summary]. Liet. TSR Mok. Akad. Darbai, Ser. C, 3(29):3-12.
- 87 Canada. Department of Agriculture. 1965. Control of potato diseases in Newfoundland. Can. Dep. Agr. Pub. 1248. 12 p.
- 88 Canada. Department of Agriculture. Experimental Farm, Charlottetown, P.E.I. 1963. Research report, 1958 - 1961. 24 p.
- 89 Edinburgh. School of Agriculture. 1971. Experimental work. 146 p.
- 90 Edinburgh. School of Agriculture. 1972. Annual report, 1971. 90 p.
- 91 Fink, H. C. 1958. Streptomycin-fungicide mixtures as potato seed piece treatments. Plant Dis. Rep. 42:965-971.

- 92 Florida. Agricultural Experiment Station. 1954. Annual report for the year ending June 30, 1953. 354 p.
- 93 Graham, D. C., and J. L. Hardie. 1971. Prospects for control of potato blackleg disease by the use of stem cuttings. Pages 219-224 in Proc. Sixth Brit. Insect. Fungic. Conf. 1971, Brighton, 15 - 18 Nov. 1971. 2 v.
- 94 Graham, D. C., and P. C. Harper. 1967. Potato blackleg and tuber soft rot. Scot. Agr. 46:68-74.
- 95 European Association for Potato Research. 2nd International Conference, Pisa, Sept. 2-7, 1963. Proceedings. Wageningen, 1964. 247 p.
- 96 Kapustin, M. N. 1967. Bacterial wet rots of potato [in Russian]. Zashch. Rast., Moscow, 12(7):30-31.
- 97 Kovacicova, E., and B. Urosevic. 1967. Some bacterial parasites observed on the market-garden Solanaceae of Tunisia [in French, English summary]. Ann. Inst. Nat. Rech. Agron. Tunisie, 40(7), 20 p.
- 98 Lazar, I., and E. Bucur. 1964. A contribution to the study of blackleg and soft rot of potato tubers in Romania [in Romanian]. Studii Cerc. Biol., Bot. Ser. 16:453-465.
- 99 Robinson, D. B., and R. R. Hurst. 1956. Control of potato black-leg with antibiotics. Amer. Potato J. 33:56-59.
- 100 Schultz, E. S., R. Bonde, and W. P. Raleigh. 1934. Isolated tuber-unit seed plots for the control of potato virus diseases and blackleg in northern Maine. Maine Agr. Exp. Sta. Bull. 370. 32 p.
- 101 Shuvalova, S. Z. 1963. Effectiveness of the treatment of potatoes for the control of common scab [in Russian]. Sb. Nauch. Rab. Kurgansk. 1962. 7:63-68.
- 102 Shuvalova, S. Z. 1966. Protection of potato from virus and bacterial diseases [in Russian]. Sb. Nauch. Rab. Kurgansk. Obl. Gos. Sel'. Khoz. Opyt. Stn. 1:133-139.
- 103 Tucker, J. 1937. The value of seed potato certification to the potato industry. Amer. Potato J. 14:39-45.
See also 148

Erwinia atroseptica, Detection of

- 104 Shneider, Yu. I., and O. Khilkova. 1967. Succinic acid and black leg of potato [in Russian]. Zashch. Rast. Moscow, 12(1):56.

Erwinia atroseptica, Effect of, on yield

- 105 Klarner, S. 1963. The problem of the development of production of chemicals for plant protection in Poland in the light of agricultural requirements and economic aspects [in Polish]. Postepy Nauk Roln. 10(3):27-50.

Erwinia atroseptica, Effect of, on plant tissues

- 106 Volcani, Z., A. J. Riker, and A. C. Hildebrandt. Destruction of various tissues in culture by certain bacteria. Phytopathology 43:92-94.

Erwinia atroseptica, Effect of crop rotation

- 107 Babaev, S. A. 1973. Influence of predecessors on potato diseases [in Russian]. Vestn. Sel'skokhoz. Nauki Kazak. 16:52-54.

Erwinia atroseptica, Effect of overhead irrigation

- 108 Bochow, H., and A. Heide. 1969. Phytosanitary effects of additional sprinkling of field crops [in German]. Arch. Pflanzenschutz. 5(3):167-178.

Erwinia atroseptica, Effect of sprinkler irrigation

- 109 Heide, A. 1968. Phytosanitary effects of field sprinkling [in German, English summary]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst. 22(8):164-167.

Erwinia atroseptica, Factors affecting

- 110 Edinburgh. School of Agriculture. 1970. Experimental work. 1969. 101 p.
- 111 Graham, D. C., and P. C. Harper. 1966. Effect of inorganic fertilizers on the incidence of potato blackleg disease. Eur. Potato J. 9:141-145.

Erwinia atroseptica, Factors affecting (contd.)

- 112 Harper, P. C., and A. E. W. Boyd. 1963. Growth cracking and bacterial soft rot in potato tubers. *Plant Pathol.* 12:139-142.
- 113 Herold, M., E. Pett, and D. C. Graham. 1969. The influence of pre-infection temperature and period of wound healing on the infection of damaged potato tubers with *Erwinia atroseptica* [in German, English summary]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst* 23(1):6-9.
- 114 Hingorani, M. K., and S. K. Addy. 1954. Factors influencing bacterial soft rot of potatoes. *Indian Phytopathol.* 6:110-115.
- 115 Lazar, I., and E. Bucur. 1964. Recent research in Romania on blackleg and bacterial soft rot of potato [in Romanian, German, French summary]. *Eur. Potato J.* 7:102-111.
- 116 Logan, C. 1964. Bacterial hard rot of potato [in German, French summary]. *Eur. Potato J.* 7:45-56.
- 117 Lund, B. M., and J. C. Nicholls. 1970. Factors influencing the soft-rotting of potato tubers by bacteria. *Potato Res.* 13:210-214.
- 118 Malyugin, P. A. 1964. Diseases of potato during irrigation in the sandy semi-desert of Western Kazakhstan [in Ukrainian]. *Sb. Tr. Zashch. Rast.* 2:20-24.
- 119 Nielsen, L. W. 1964. Pathogenesis of three *Erwinia* species to potato tuber tissue in a CO₂-N atmosphere. *Phytopathology* 54: 902. (Abstr.)
- 120 Scottish Horticultural Research Institute. 1972. Annual report, 18th, for the year 1971.
- 121 Smith, W. L., and H. F. Smart. 1955. Relation of soft rot development to protective barriers in Irish potato slices. *Phytopathology* 45:649-654.
- 122 Van den Boom, T. 1967. Studies on the conditions needed for the occurrence of potato black leg [in German]. *Phytopathol. Z.* 58:239-276.
- 123 Voronkevich, I. V., and L. A. Butsevich. 1964. Importance of soil infection and conditions of potato growth for the development of "black leg" [in Russian]. *Dokl. Vses. Akad. Sel'skokhoz. Nauk.* 1964. 9:30-33.

See also 86, 94, 96, 99

Erwinia atroseptica, Incidence of

- 124 Masurat, G., and S. Stephan. 1964. The occurrence of most important diseases and pests of agricultural and horticultural plants in 1963 in the territory of the German Democratic Republic [in German]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst* 18(6):141-166.

See also 94

Erwinia atroseptica, Inhibition of

- 125 Vicente, R. 1954. Inhibition of the rot of the potato tuber during its germination period [in Spanish]. *An. Edafol. Fisiol. Veg.* 13(9-10):705-723.

Erwinia atroseptica, Method for testing resistance to

- 126 Kiel, W. 1967. Development of a laboratory method of testing the resistance of potatoes to *Erwinia atroseptica*, the cause of tuber soft rot [in German, English summary]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst* 21:237-240.

Erwinia atroseptica, Method of diagnosing

- 127 Graham, D. C. 1963. Serological diagnosis of potato blackleg and tuber soft rot. *Plant Pathol.* 12:142-144.

Erwinia atroseptica, Mode of entry and spread

- 128 Fox, R. T. V., J. G. Manners, and A. Myers. 1971. Ultrastructure of entry and spread of *Erwinia carotovora* var. *atroseptica* into potato tubers. *Potato Res.* 14:61-73.

Erwinia atroseptica, Mode of infection

- 129 Maas - Geesteranus, H. P. 1971. Mode of infection...[in Dutch, English summary]. Pages 51-54 in *Jaarverslag 1970*, Inst. Plantenziektenkundig Onderzoek, Wageningen.

Erwinia atroseptica, Effect of succinic acid

- 130 Gerasimova, T. P. 1968. Influence of succinic acid on the occurrence of black leg of potato [in Russian]. *Tr. Nauch. Issled. Inst. Kartof. Khva.* 5:191-195.

See also 104

Erwinia atroseptica, Overwintering

- 131 Logan, C. 1969. The survival of the potato black leg pathogen overwinter. *Rec. Agr. Res. Min. Agr. N. Ir.* 17:115-121.

Erwinia atroseptica, Proteinase activity

- 132 Ragozina, I. I., I. Yu. Shneider, and D. V. Lipsits. 1969. Activity of proteinases in potato tissues infected by 'black leg' *Erwinia atroseptica* [in Russian]. *Dokl. Akad. Nauk SSSR*, 184(1):242-245.

Erwinia atroseptica, Selective isolation of

- 133 Logan, C. 1963. A selective medium for the isolation of soft rot coliforms from soil. *Nature* 199(4893):623.

Erwinia atroseptica, Serological diagnosis and symptoms of

- 134 Shneider, Yu. I., and K. F. Murzakova. 1964. Bacteriosis of potato [in Russian]. *Zashch. Rast. Moscow*, 9(9):32-33.

Erwinia atroseptica, Spread of

- 135 Graham, D. C. 1962. Black leg disease of potatoes. *Scott. Agr.* 41:211-215.
- 136 Lund, B. M., and G. M. Wyatt. 1972. The effect of oxygen and carbon dioxide concentration on bacterial soft rot of potatoes. I. King Edward potatoes inoculated with *Erwinia carotovora* var. *atroseptica*. *Potato Res.* 15:174-179.
- 137 Nickel, J. L. 1954. Results of potato seed-piece treatment tests in Kern County, California. *Amer. Potato J.* 31:245-251.

Erwinia atroseptica, Survival of, in soil

- 138 Scottish Horticultural Research Institute. 1968. Annual report, 14th. 71 p.

Erwinia atroseptica, Technique for differentiation of, from saprophytes

- 139 Novakova, J. 1957. A new method of isolation of blackleg - pathogens from diseased plants. *Phytopathol. Z.* 29:72-74.

Erwinia atroseptica, Technique for evaluating resistance to

- 140 Henniger, H. 1965. Investigations on tuber and storage rot of potato. I. On the technique of testing for resistance with the causal organism of blackleg, *Erwinia atroseptica* [in German]. *Züchter* 35(4):174-180.
- 141 Shneider, Yu. I. 1965. Assessment of resistance in potato varieties [in Russian]. *Zashch. Rast. Vred. Bolez.* 10(12):22-23.

Erwinia atroseptica, Testing chemicals against

- 142 Brazda, G. 1971. The potato tuber piece test - a method for testing the efficiency of bactericides and bacteriostats against *Erwinia atroseptica* [in German]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst* 25(3):59-60.

Erwinia atroseptica, Tuber infection

- 143 Cambridge. Plant Breeding Institute. 1971. Annual report, 1970. 126 p.

Erwinia atroseptica, Ultrastructure

- 144 Fox, R. T. V. 1971. The ultrastructure of potato tubers infected by the soft rot organism *Erwinia carotovora* var. *atroseptica*. Pages 95-120 in *Proc. Third Int. Conf. Plant Pathog. Bact. Wageningen*, 14-21 Apr., 1971. 365 p.
- 145 Plombelon, M. C. M. 1971. A quantitative method for assessing virulence of *Erwinia carotovora* var. *atroseptica* and susceptibility to rotting of potato tuber tissue. pages 299-303 in *Proc. Third Int. Conf. Plant Pathog. Bact. Wageningen*, 14-21 Apr., 1971. 365 p.

Erwinia atroseptica, Varietal reaction to

- 146 Davies, H. T., D. A. Young, J. Munro, L. C. Young, and the Atlantic Regional Potato Breeding Committee. 1963. Hunter, a new potato variety with excellent cooking quality and field immune to viruses X and A. Amer. Potato J. 40:275-278.
- 147 Gregor, J. W. 1964. Director's report. Potatoes. Rec. Scott. Plant Breed. Sta. 1964. pp. 5-12.
- 148 Knutson, K., R. F. Line, and C. J. Eide. 1959. Varietal response to seed piece decay. Plant Dis. Rep. 43:546-548.

See also 115, 199

Erwinia carotovora

- 149 Alvarado, E. L. F., and N. J. Guzman. 1968. Potato storage rots [in Spanish, English summary]. Rev. Inst. Colomb. Agropec. 3:47-61.
- 150 Amani. 1967. Soft rot of ornamentals and vegetables. Iran. J. Plant Pathol. 4(2):1-10.
- 151 Ark, P. A. 1946. Some laboratory and field data on ring-rot of potatoes in California. Amer. Potato J. 23:170-180.
- 152 Bennett, F. T. 1946. Soft rot of potatoes in 1945 crops. J. Min. Agr. 53:56-58.
- 153 Bonde, R. 1930. Some conditions determining potato seed-piece decay and blackleg induced by maggots. Phytopathology 20:128. (Abstr.)
- 154 Bonde, R. 1950. Factors affecting potato blackleg and seed-piece decay. Maine Agr. Exp. Sta. Bull. 482. 31 p.
- 155 Bortels, H. 1951. Further studies on the relation between the establishment of infection and the course of the weather in potato wet rot. Preliminary note [in German]. Phytopathol. Z. 18:360-362.
- 156 Biological Institute for Agriculture and Forestry at Braunschweig, Germany. 1952. Annual report for 1951 [in German]. 100 p.
- 157 Canada. Department of Agriculture. 1955. Report of the Minister of Agriculture for Canada for the year ended March 31, 1955.
- 158 Ciampi, L. R. 1972. Bacterial diseases of potato in Chile [in Spanish]. Fitopatologia 7(1/2): 5-14.
- 159 Ciampi, P. 1972. Taxonomic study of causal agents of soft rots on potato tubers [in Spanish]. Agr. Tec. 32(4):176-181.
- 160 Dainello, F. J. 1970. Suberization studies in the potato (*Solanum tuberosum* L.). Diss. Abstr. Int. 30(7):2978B.
- 161 Davidson, R. S. 1948. Factors affecting the development of bacterial soft rot of potato tuber initials. Phytopathology 38:673-687.
- 162 Dennis, R. W. G., and E. G. Gray. 1954. A first list of the fungi of Zetland (Shetland). Trans. Bot. Soc. Edinburgh 36(3):214-233.
- 163 Dewey, D. H., and W. R. Barger. 1948. The occurrence of bacterial soft rot on potatoes resulting from washing in deep vats. Proc. Amer. Soc. Hort. Sci. 52:325-330.
- 164 Dowson, W. J., and D. Rudd Jones. 1951. Bacterial wet rot of potato tubers following *Phytophthora infestans*. Ann. Appl. Biol. 38:231-236.
- 165 Dykstra, T. P. 1941. Results of experiments in control of bacterial ring rot of potatoes in 1940. Amer. Potato J. 18:27-55.
- 166 Eddins, A. H., G. D. Ruehle, and G. R. Townsend. 1949. Potato diseases in Florida. Florida Agr. Exp. Sta. Bull. 427. 96 p.
- 167 Fernandez Valiela, M. V., and A. V. Calderoni. 1965. The search for potato growing areas in the Argentine Republic [in Portuguese]. Atas Inst. Micol. 2:60-76.
- 168 Fernando, M., and G. Stevenson. 1952. Studies in the physiology of parasitism. XVI. Effect of the condition of potato tissue, as modified by temperature and water-content, upon attack by certain organisms and their pectinase enzymes. Ann. Bot. 16(61):103-114.
- 169 Graham, D. C., and Z. Volcani. 1961. Experiments on the control of black-leg disease of potato by disinfection of seed tubers with mercury compounds and streptomycin. Eur. Potato J. 4:129-137.
- 170 Gregg, M. 1952. Studies in the physiology of parasitism. XVII. Enzyme secretion by strains of *Bacterium carotovorum* and other pathogens in relation to parasitic vigour. Ann. Bot. 16(62):235-250.
- 171 Hansen, F. 1953. Investigations on the storage of potatoes [in Danish, English summary]. Tidsskr. Planteavl. 56:222-245.
- 172 Hingorani, M. K., and S. K. Addy. 1953. A comparative study of *Erwinia carotovora*, *Erwinia aroideae*, and *Erwinia atroseptica*. Indian Phytopathol. 5:40-43.
- 173 Kendrick, J. B., R. T. Wedding, and A. O. Paulus. 1959. Temperature - relative humidity index for predicting the occurrence of bacterial soft rot of Irish potatoes. Phytopathology 49:701-705.
- 174 Kharchenko, S. M. 1961. The antibiotic properties of species in the *Monovorticillata* sect. of *Penicillium* isolated from the rhizosphere of agricultural plants in the Ukraine [in Ukrainian]. Mikrobiol. Zh. (Kyyiv) 23:46-50.
- 175 Leach, J. G. 1940. Potato blackleg. Pages 168-178 in Insect transmission of plant diseases. New York, McGraw-Hill.
- 176 Leach, J. G. 1930. Survival of the potato-blackleg pathogene in the soil and some factors influencing infection. Phytopathology 20:127. (Abstr.)
- 176a Leach, J. G. 1930. Potato black-leg: The survival of the pathogene in the soil and some factors influencing infection. Phytopathology 20:215-228.
- 177 Lund, B. M. 1971. Bacterial spoilage of vegetables and certain fruits. J. Appl. Bacteriol. 34:9-20.
- 178 Lyon, G. D. 1972. Occurrence of rishitin and phytuberin in potato tubers inoculated with *Erwinia carotovora* var. *atroseptica*. Physiol. Plant. Pathol. 2:411-416.
- 179 Maas - Geesteranus, H. P. 1972. Soft rot and black leg on potatoes. [in Dutch]. Bedrijfsontwikkeling 3:941-945.
- 180 Mateev, A. 1959. Cleaning and approval at the potato seed stations [in Russian]. Gradinarstvo 1(6):7-11.
- 181 Mujica, R. F. 1941. List of potato diseases and pests of which the existence has been established in the country [in Spanish]. Bol. Sanid. Veg., Santiago 1(1):70-72.
- 182 New South Wales. Department of Agriculture. Biological Branch. Division of Science Services. 1954. Plant disease survey for the twelve months ending 30th June, 1953. 23d annual report. 38 p.
- 183 Peschel, R. 1969. The occurrence of the most important diseases and pests of agricultural and horticultural plants in 1968 in the German Democratic Republic [in German]. Nachrichtenbl. Dtsch. Pflanzenschutzdienst 23(8):141-170.
- 184 Potato Association of America. Colorado State University, Fort Collins, July 25-27, 1966. Annual meeting, 50th. Amer. Potato J. 43:340-348.
- 185 Plant diseases. Agr. Gaz. N.S.W. 60:646-650. 1949.
- 186 Plant diseases. Notes contributed by the Biological Branch. Agr. Gaz. N.S.W. 60:595-600. 1949.
- 187 Plant diseases - Blackleg of potatoes in New South Wales. The question of seed transmission. Agr. Gaz. N.S.W. 63:534-536. 1952.
- 188 Quebec Society for the Protection of Plants. 1952. Reports, 32d and 33d. 1951, 1952. 232 p.
- 189 Reid, W. J., R. C. Wright, and W. M. Peacock. 1940. Prevention of damage by the seed-corn maggot to potato seed pieces. U.S. Dep. Agr. Tech. Bull. 719. 37 p.
- 190 Robbs, C. F. 1960. Phytopathogenic bacteria in Brazil [in Portuguese, English summary]. Inst. Econ. Rural, Rio de J., Str. Divulg. Pesq. 2, 63 p.
- 191 Rose, D. H., and H. A. Schomer. 1944. Relation of heat and desiccation to bacterial soft rot of potatoes. Amer. Potato J. 21:149-161.
- 192 Rudd Jones, D., and W. J. Dowson. 1950. On the bacteria responsible for soft rot in stored potatoes, and the reaction of the tuber to invasion by *Bacterium carotovorum* (Jones) Lehmann & Neumann. Ann. Appl. Biol. 37:563-569.
- 193 Sampson, P. J., and A. C. Hayward. 1971. Some characteristics of pectolytic bacteria associated with potato in Tasmania. Aust. J. Biol. Sci. 24:917-923.

Erwinia carotovora (contd.)

- 194 Sherf, A. F. 1944. Infection experiments with potato ring rot and the effect of soil temperature on the disease. *Amer. Potato J.* 21:27-29.
 - 195 Smarda, J. 1963. Lysogeny and bacteriocinogeny. *Folia Microbiol.* 8(4):254-263.
 - 196 Smith, M. A. 1944. Bacterial soft rot of spinach. *Phytopathology* 34:747-752.
 - 197 Smith, M. A., and G. B. Ramsey. 1947. Bacterial lenticel infection of early potatoes. *Phytopathology* 37:225-242.
 - 198 Staruigina, L. P. 1949. Agents of soft rot in cabbage grown for seed [in Russian]. *Microbiology* 17:160-170.
 - 199 Taylor, C. F., and F. M. Blodgett. 1937. Control of a wilt disease of potato by formaldehyde dust. *Amer. Potato J.* 14:154-157.
 - 200 Wahlin, B. 1950. Parasitic infection of agricultural crops in East Gothland and the North Kalmar district 1950 [in Swedish]. *Vaxtskyddsnotiser, Vaxtskyddsanst. Stockholm*, 1950. (6):71-76.
 - 201 Watson, R. D. 1943. Charcoal rot of Irish potatoes. *Phytopathology* 33:1120. (Abstr.)
 - 202 White, N. H. 1946. Potato tuber rots. *Tasmanian J. Agr.* 17:235-241.
- See also 17, 23, 26, 34, 35, 39, 47, 58, 66, 72, 79, 98, 106, 114, 115, 145, 467, 526, A1, A2, A3, A7, A13, A22, A23, A28, A34, A60, A63, A78, A79, A85, A86, A87, A88, A89, A99, A104.

Erwinia carotovora, Assessing infection

- 203 Rothamsted. Experiment Station. 1971. Report for 1970. Pt. I. 385 p.

Erwinia carotovora, Control

- 204 Burkholder, W. H., and W. L. Smith. 1949. *Erwinia atroseptica* (Van Hall) Jennison and *Erwinia carotovora* (Jones) Holland. *Phytopathology* 39:887-897.
- 205 Canada. Department of Agriculture. 1958. Report of the Minister of Agriculture for Canada for the year ended March 31, 1958. 144 p.
- 206 Crossan, D. F. 1959. Control of potato seed-piece decay. *Plant Dis. Rep.* 43:543-545.
- 207 Rhode Island. Agricultural Experiment Station. 1947. Annual report, 59th. 56 p.
- 208 Shneider, Yu. I., T. P., Gerasimova, and A. A. Il'icheva. 1968. Chemical method of control of black leg of potato [in Ukrainian]. *Bakteriol. Bolez. Rast. Metod Bori Nimi. Kiev.* pp. 313-315.
- 209 Vielwerth, V. 1947. Is effective protection against blackening of potato stems and tuber rot possible? [in Czechoslovakian]. *Ochr. Rost.* 19-20(10-11):28-35.

Erwinia carotovora, Decomposition of pectic substances by

- 210 Gorlenko, M. V. 1961. Bacterial diseases of plants [in Russian]. 2d rev. ed. Moscow, Gos. Izdat.
- 211 Starr, M. P., and F. Moran. 1962. Eliminative split of pectic substances by photopathogenic soft-rot bacteria. *Science* 135(3507):920-921.

Erwinia carotovora, Defence reaction

- 212 Hawn, W. 1969. Studies on the defence reaction of potato tubers against black leg and soft rot caused by *Erwinia carotovora* [In German, English summary]. *Acta Phytopathol. Acad. Sci. Hung.* 4:63-76.

Erwinia carotovora, Effect of, on yield

- 213 Conroy, R. J. 1958. Powdery dry rot of potato. *Agr. Gaz. N.S.W.* 69:299-302.

Erwinia carotovora, Effect of EDTA

- 214 Zucker, M., and L. Hankin. 1970. Effectiveness of ethyldiaminetetraacetic acid (EDTA) in controlling soft rot of potatoes. *Plant Dis. Rep.* 54:863-865.

Erwinia carotovora, Effect of gamma radiation

- 215 Beraha, L., G. B. Ransey, M. A. Smith, and W. R. Wright. 1959. Effects of gamma radiation on some important potato tuber decays. *Amer. Potato J.* 36:333-338.

Erwinia carotovora, Effect of ionizing radiation

- 216 Hooker, W. J., and D. T. Duncan. 1959. Storage rot susceptibility of potato tubers exposed to ionizing irradiation. *Amer. Potato J.* 36:162-172.

Erwinia carotovora, Effect of light

- 217 Jaffe, M. J. 1965. Chlorophyll production, lignin deposition, synthesis of phenolic compounds and inhibition of bacterial soft rot by photoactivation in the potato tuber. *Diss. Abstr.* 25(9):4927.

Erwinia carotovora, Effect of red light

- 218 Jaffe, M. J., R. Dickey, and F. M. R. Isenberg. 1969. Inhibition of bacterial soft rot in the potato tuber by red light photoactivation. *Phyton Rev. Int. Bot. Exp.* 26:69-76.

Erwinia carotovora, Effect of streptomycin

- 219 Olgyay, M. 1956. Infection experiments with *Colletotrichum atramentarium* on potatoes [in Hungarian]. *Agrartud. Egy. Evkönyv.* 2:195-204.
- 220 Waggoner, P. E. 1956. Chemical treatment of potato seed in Connecticut 1955. *Plant Dis. Rep.* 40:411-413.

Erwinia carotovora, Factors affecting

- 221 L'vova, N. M. 1964. Effect of temperatures near 0°C. on the microflora of potatoes and their resistance to microorganisms during storage [in Russian]. *Sb. Tr. Leningrad. Inst. Sov. Torg.* 23:47-56.
- 222 Lynch, P. B. 1953. Potato variety trials. *N.Z.J. Agr.* 86:321.

- 223 Masurat, G., R. Peschel, and S. Stephan. 1966. The occurrence of the most important diseases and pests of agricultural and horticultural plants in 1965 in the German Democratic Republic [in German]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst* 20:121-142.

- 224 Plant diseases. Corm rot of bananas. Blackleg of potatoes. Septoria spot of citrus. *Agr. Gaz. N.S.W.* 64:546-549. 1953.

- 225 Plant diseases - blackleg of potatoes in New South Wales. The question of seed transmission. *Agr. Gaz. N.S.W.* 63:534-536. 1952.

See also 119, 207, 243

Erwinia carotovora, Fungi-toxicity of aerosols

- 226 Koula, V. 1971. Preparation and physical and chemical properties of aerosol solutions containing organic tin compounds. Fungitoxicity and warm aerosols containing organic tin compounds [In Czechoslovakian, English summary]. *Ochr. Rost.* 7:211-217; 219-224.

Erwinia carotovora, Interaction with *Gibberella pulicaris*

- 227 Noll, A. 1972. On the interaction of *Gibberella pulicaris* and *Erwinia carotovora* in storage rots of potato tubers [German, English summary]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst* Stuttgart. 24(1):1-3.

- 228 Stachewicz, H. 1970. Studies on potato dry rot with special reference to late blight and soft rot [in German, English summary]. *Arch. Pflanzenschutz.* 6:455-467.

Erwinia carotovora, Medium for detecting

- 229 Lapwood, D. H. 1957. Studies in the physiology of parasitism. XXIII. On the parasitic vigour of certain bacteria in relation to their capacity to secrete pectolytic enzymes. *Ann. Bot.* 21:167-184.

- 230 Plrombelon, M. C. M. 1972. Reliable and rapid method for detecting contamination of potato tubers by *Erwinia carotovora*. *Plant Dis. Rep.* 56:552-554.

Erwinia carotovora, Enzyme activity

- 231 Mount, M. S., D. F. Bateman, and H. G. Basham. 1970. Induction of electrolyte loss, tissue maceration, and cellular death of potato tissue by an endopolygalacturonate trans-eliminase. *Phytopathology* 60:924-931.

Erwinia carotovora, Tissue extracts

- 232 Spalding, D. H., and B. C. Smale. 1969. Comparative interactions of *Erwinia carotovora* and *Erwinia amylovora* with pear and potato tissues. *Plant Dis. Rep.* 53:255-256.

Erwinia carotovora, Phenolic compounds in relation to

- 233 Lovrekovich, L., H. Lovrekovich, and M. A. Stahmann. 1967. Inhibition of phenol oxidation by *Erwinia carotovora* in potato tuber tissue and its significance in disease resistance. *Phytopathology* 57:737-742.

Erwinia carotovora, Ring-rot symptoms produced by

- 234 Davidson, R. S. 1946. Ring-rot-like symptoms produced by soft-rot bacteria in potato tubers. *Phytopathology* 36:237-239.

Erwinia carotovora, Serology of

- 235 Okabe, N., and M. Goto. 1956. Studies on the strains of *Erwinia carotovora* (Jones) Holland. I. Antigenic structures of flagella and their relations to pathogenicity and maltose fermentation [in Japanese, English summary]. *Bull. Fac. Agr. Shizuoka Univ.* 6:16-32.

Erwinia carotovora, Technique for isolating

- 236 Kerr, A. 1953. A method of isolating soft-rotting bacteria from soils. *Nature* 127(4390):1155.

Erwinia carotovora, Ultrastructure

- 237 Fox, R. T. V., J. G. Manners, and A. Myers. 1972. Ultrastructure of tissue disintegration and host reactions in potato tubers infected by *Erwinia carotovora* var. *atroseptica*. *Potato Res.* 15:130-145.

Erwinia carotovora, Varietal reaction to

- 238 Dobias, K. 1970. The resistance of varieties of the world potato collection to *Erwinia carotovora* [in Czechoslovakian, English summary]. *Rostl. Vyr.* 16(7):687-692.
- 239 Hey, A. 1954. The phytopathogenic bases of seed potato production [in German]. *Dtsch. Landwirt.* 5:302-306.
- 240 Hollis, J. P., and R. W. Goss. 1950. Factors influencing invasion of potato by *Erwinia carotovora*. *Phytopathology* 40:860-868.
- 241 Mills, W. R. 1964. Pennchip, a new potato variety resistant to late blight and scab with superior chipping quality. *Amer. Potato J.* 41:54-58.
- 242 New South Wales. Department of Agriculture. Division of Science Services. 1954. Plant disease survey for the twelve months ending 30th June, 1953. 23rd annual report.
- 243 Nielsen, L. W. 1954. The susceptibility of seven potato varieties to bruising and soft rot. *Phytopathology* 44:30-35.
- 244 Olofsson, B. 1963. Potato black leg [in Swedish]. *Vaxtskyddsnotiser*, Stockholm 27(5-6):71-75.
- 245 Telneset, S. O. 1964. Studies on potato tuber rot. *Diss. Abstr.* 24(11):4341.

Erwinia phytophthora (See also *E. atroseptica*)

- 246 Bailey, H. L. 1942. Report on the division of plant pest control. *Rep. Vermont Agr. Exp. Sta.* 1941 - 1942. pp. 43-51.
- 247 Borg, A. 1949. Some plant diseases and pests of agricultural plants in West Gothland 1949 [in Swedish]. *Vaxtskyddsnotiser*, Stockholm, 1949(6):6-10.
- 248 Bortels, H. 1949. On the dependence of the virulence and other properties of phytopathogenic bacteria, and of the success of inoculation, on the course of the weather [in German]. *Phytopathol. Z.* 15:376-393.
- 249 Callbeck, L. C. 1949. Potato vine killing in Prince Edward Island. *Amer. Potato J.* 26:409-419.

- 250 Emilsson, B., and N. Gustafsson. 1949. Studies relating to the control of potato blight. IV. Further experiments with haulm-killing preparations [in Swedish, English summary]. *Kgl. Lanthruksakad. Tidskr.* 88(2):188-200.
- 251 Fernow, K. M., and O. C. Garcés. 1949. Production of certified potato seed [in Spanish]. *Rev. Fac. Agron. Medellín* 10(36):257-295.
- 252 Kirulis, A. 1942. The microscopic fungi as natural enemies of plant diseases in Latvia [in German]. *Arb. Landwirtsch. Akad. Mitn* 1:479-536.
- 253 Lefebvre, C. L. 1950. Observations on plant diseases in Alaska. *Plant Dis. Rep.* 34:3-4.
- 254 Lehmann, H. 1938. Further contribution to the problem of physiological specialization of *Phytophthora infestans* de Bary, the causal agent of potato blight [in German]. *Phytopathol. Z.* 11:121-154.
- 255 Macek, J. 1950. The influence of temperature on the health and yield of stored potatoes [in Czechoslovakian, English summary]. *Ochr. Rost.* 23(4):304-316.
- 256 Burke, O. D. The occurrence in the United States of the tuber ring rot and wilt of the potato. *Plant Dis. Rep.* 22:444-445. 1938.
- 257 Peralta, G. J. 1949. Report by the plant pathologist for the year 1946 - 1947. *Rep. Dep. Agr. Malta*, Oct. 1938 to Sept. 1946 and the agricultural year 1946 - 1947. 74-79.
- 258 Stapp, C. 1938. Potato blackleg [in German]. *Kranke Pflanze* 15(6):103-106.
- 259 Schaal, L. A., W. C. Edmundson, and R. Kunkel. 1949. Yampa, a new scab - resistant potato. *Amer. Potato J.* 2:335-342.
- 260 Stevenson, F. J. and G. E. DeLong. 1949. Canus: a new potato variety adapted to Alberta and other sections of the Dominion of Canada. *Amer. Potato J.* 26:326-330.
- 261 Störmer, I. 1943. Hygienic precautions for seed potatoes [in German]. *Mitt. Landwirt.*, Berlin, 58(25):475-478.
- See also 29, 171, 189, 200

Erwinia solani

- 262 Jensen, J. H., and R. W. Goss. 1941. Infection of first-year potato seedlings with *Fusarium solani* var. *eumartii*. *Amer. Potato J.* 18:239-242.

Fusarium

- 263 Ayers, G. W. 1972. *Fusarium* decay in potatoes. *Can. Agr.* 17:38-39.
- 264 Ayers, G. W. 1970. Potato seed treatment. *Pestic. Res. Rep.*, Res. Br., Agr. Can. pp. 260-261.
- 265 Bald, J. G. 1941. A report on agricultural features of the Australian potato industry. *Pamphl. Coun. Sci. Ind. Res. Aust.* 106, 72 p.
- 266 Bazan de Segura, C. 1956. Verticilliose of potatoes in Peru. *Plant Dis. Rep.* 40:1091.
- 266a Bazan de Segura, C. 1958. Further notes on verticillium wilt of potato in Peru: *FAO Plant Prot. Bull.* 6:125-126.
- 267 Bean, J. L. 1947. *Eumerus strigatus* reared from decayed potatoes. *J. Econ. Entomol.* 40:452-454.
- 268 Beaumont, A., and L. N. Staniland. 1937. Thirteenth annual report Dep. Plant Pathology. Seale-Hayne Agricultural College, Newton Abbot, Devon, for the year ending September 30th, 1936. 35 p.
- 269 Bhargava, K. S., and H. Kishore. 1953. Occurrence of *Verticillium* in potato tubers in India. *Nature* 171 (4357):800-801.
- 270 Blodgett, E. C., and A. Rich. 1949. Potato tuber diseases, defects, and insect injuries in the Pacific Northwest. *Wash. Agr. Exp. Sta. Pop. Bull.* 195, 116 p.
- 271 Brandenburger, W. Studies on skin necrosis of the potato tuber [in German, English summary]. *Phytopathol. Z.* 34:229-268.
- 272 Brewer, P. J. 1962. Potato stem-end rot caused by *Fusarium* spp. *S. Afr. J. Agr. Sci.* 5:475-479.
- 273 Busch, L. V., and R. G. Rowberry. 1972. Potato seed-piece treatment in Ontario. *Amer. Potato J.* 49:7-11.

Fusarium (contd.)

- 274 Canada. Department of Agriculture. 1966. *Fusarium* rot of potatoes. This Month with C.D.A. 12:9-10.
- 275 Chamberlain, E. E. 1935. Fungi present in the stem-end of potato tubers. N.Z.J. Sci. Tech. 14(4):242-246.
- 276 Correll, D. S. 1948. Collecting wild potatoes in Mexico. U.S. Dep. Agr. Circ. 797. 40 p.
- 277 Costa, A. S., and H. P. Krug. 1937. Potato diseases in Sao Paulo [in Spanish]. Bol. Inst. Agron. Campinas 14. 55 p.
- 278 Das, C. R., and A. Pal. 1968. Influence of *Rhizopus nigricans* Ehrenb. on the development of *Alternaria solani* (Ell. & Mart.) Jones & Grout. Phytopathol. Z. 63:40-46.
- 279 Dippenaar, B. J. 1934. *Fusarium* - rot in potatoes. Farming in South Africa 9(95):58.
- 280 Easton, G. D., M. E. Nagle, and D. L. Bailey. 1970. Potato seed piece treatment in Washington. Amer. Potato J. 47:469-474.
- 281 Edgerton, C. W. 1938. Report of seed certification conference. Amer. Potato J. 15:130-140.
- 282 Elpidina, O. K. 1935. On toxins of wilting. C.R. Acad. Sci. U.R.S.S., N.S. 3(8):360-364.
- 283 Feddersen, H. D. 1962. Target spot of potatoes. Trials show value of spraying. J. Agr. S. Aust. 65:300-308.
- 284 Felton, M. W. 1948. The development of stem end discoloration in Bliss Triumph potatoes held in warm storage. Amer. Potato J. 25:49-50. (Abstr.)
- 285 Feuerbach, P. 1948. Potato loss from decay [in German]. Pflanzenschutz 1(3):29-32.
- 285a Fischer, R., and H. Neumann. 1932. Report on the work of the Committee of Potato Experts during the year 1932 [in German]. Neuheiten Geb. Pflanzenschusch. 1932(5-6):101-108.
- 286 Florida. Agricultural Experiment Station. 1950. Annual report for the year ending June 30, 1949. 333 p.
- 287 Florida. Agricultural Experiment Station. 1959. Annual report for the year ending June 30, 1958. 411 p.
- 288 Folsom, D., W. C. Libby, G. W. Simpson, and O. L. Wyman. 1938. Net necrosis of potatoes. Maine Coll. Agr. Exp. Serv. Bull. 246. 12 p.
- 289 Friedman, B. A. 1950. Behavior of potato internal brown spot in stored tubers. Phytopathology 40:899-901.
- 290 Friedman, B. A., and D. Folsom. 1953. Potato tuber glassy-end and jelly-end rot in northeast in 1949 and 1952. Plant Dis. Rep. 37:455-459.
- 291 Galloway, L. D. 1936. Report of the Imperial Mycologist. Sci. Rep. Agr. Res. Inst. Pusa, 1934 - 1935. pp. 120-130.
- 292 Glöckner, G. 1940. Investigations on the 'scorch' disease of potatoes in the Rhine Province [in German]. Angew. Bot. 22(3):201-252.
- 293 Graham, D. C., and G. A. Hamilton. 1970. Control of potato gangrene and skin spot diseases by fumigation of tubers with sec-butylamine. Nature 227(5255):297-298.
- 294 Granovsky, A. A. 1944. The value of DDT for the control of potato insects. Amer. Potato J. 21:89-91.
- 295 Gratz, L. O. 1930. Disease and climate as pertaining to the Florida and Maine potato sections. Phytopathology 20:267-288.
- 296 Guimaraes, F. F. 1953. Potato growing in Brazil. Amer. Potato J. 30:124-129.
- 297 Hirst, J. M., G. A. Hide, O. J. Stedman, and R. L. Griffith. 1973. Yield compensation in gappy potato crops and methods to measure effects of fungi pathogenic on seed tubers. Ann. Appl. Biol. 73:143-150.
- 298 Idaho. Agricultural Experiment Station. 1945. Annual report, 49, 59, 51, 52 for the years 1941, 1942, 1944, and 1945. Idaho Agr. Exp. Sta. Bull. 244. 63 p.
- 299 Idaho. Agricultural Experiment Station. 1946. Annual report, 53rd for the year ending 30th June, 1946. Idaho Agr. Exp. Sta. Bull. 268. 60 p.
- 300 Iowa. Agricultural Experiment Station. Botany and Plant Pathology Section. Report, pt. 1. pp. 119:135.
- 301 Jamalain, E. A. 1955. *Fusarium* species causing plant diseases in Finland. Acta Agr. Fenn. 83:159-172.
- 302 Koblet, R. 1947. Investigations on the influence of nitrogen manuring on the incidence of disease and palatability of the potato [in German]. Ann. Agr. Suisse 48(6-7):665-699.
- 303 Kraus, J. E. 1945. Influence of certain factors on second growth on Russet Burbank potatoes. Amer. Potato J. 22:134-142.
- 304 Leach, J. G., and H. Darling. 1935. Symptoms of potato wilt in Minnesota this year. Plant Dis. Rep. 19:299-302.
- 305 Lehmann, H. 1937. The existing primary material for the breeding of *Phytophthora* - resistant potatoes [in German]. Züchter 9(2):29-35.
- 306 Lunden, A. P. 1938. Objects and methods in the work of breeding potatoes (*Solanum tuberosum*) for resistance to disease [in Norwegian]. Meld. Norg. Landbruksheisk. Hisk. 18(3):183-198.
- 307 Lutz, J. M. 1953. *Fusarium* tuber rots of late potatoes as related to injuries and certain chemical treatments. Amer. Potato J. 30:131-134.
- 308 Maine. Agricultural Experiment Station. 1934. Summary report of progress, 1933. Maine Agr. Exp. Sta. Bull. 369:558-581.
- 309 Mammen. 1937. Important diseases of potato tubers [in German]. Mitt. Landwirt. Berlin, 52(41):861-862.
- 310 Melhus, I. E., D. R. Shepherd, and M. A. Corkle. 1941. Diseases of potatoes in Iowa. Proc. Iowa Acad. Sci. 48:133-146.
- 311 Miller, P. R., and N. Nance. 1949. Preliminary estimates of acreages of crop lands in the United States infested with some organisms causing plant diseases. Plant Dis. Rep. Suppl. 185. 207-252.
- 312 Mol, J., and H. A. Ormel. 1946. Some observations on powdery scab *Spongiospora subterranea* Wallr [in Dutch]. Tijdschr. Plantenziekten. 52:18-22.
- 313 Morwood, R. B. 1933. Potato diseases. Queensl. Agr. J. 40:382-395.
- 314 Mulder, D. 1958. Plant diseases of economic importance in the Northern Region, United Arab Republic. FAO Plant Prot. Bull. 7(1):1-5.
- 315 Napper, M. E. 1933. Observations on potato blight (*Phytophthora infestans*) in relation to weather conditions. J. Pomol. Hort. Sci. 11:177-184.
- 316 Natrass, R. M. 1932. The wilt disease of potatoes. Cyprus Agr. J. 26(4):138-139.
- 316a New South Wales. Biological Branch, Dep. Agr. 1943. Plant Notes contributed by the Branch. Agr. Gaz. N.S.W. 54:463-466.
- 317 New York. Agricultural Experiment Station. 1945. Annual report, 63rd. 62 p.
- 318 Nielsen, L. W. 1949. *Fusarium* seed-piece decay of potatoes in Idaho and its relation to blackleg. Idaho Agr. Exp. Sta. Res. Bull. 15. 31 p.
- 319 Nielsen, L. W., and J. T. Johnson. 1972. Seed potato contamination with fusarial propagules and their removal by washing. Amer. Potato J. 49:391-396.
- 321 Oort, A. J. P. 1955. New views and new results in the field of crop protection. Phytat.-Phytopharm. (Num!ro Special), 105-114.
- 323 Padwick, G. W. 1941. Report of the Imperial Mycologist. Sci. Rep. Agr. Res. Inst., New Delhi, 1939-40. pp. 94-104.
- 324 Palm, E. T., and R. A. Young. 1957. The compatibility of certain organic fungicides and antibiotics in treatment mixtures as indicated by stability and phytotoxicity. Plant Dis. Rep. 41:151-155.
- 325 Phillips, D. V., C. Leben, and C. C. Allison. 1967. A mechanism for the reduction of *Fusarium* wilt by a *Cephalosporium* species. Phytopathology 57:916-919.
- 328 Rhodesia. Department of Agriculture. Nyasaland. 1960. Annual report for the years 1958 1959. Pt. 11. 158 p.
- 329 Seminario, B., E. R. French, and L. W. Nielsen. 1970. Potato tuber resistance to fusaria affecting potatoes in Peru [in Spanish]. Amer. Potato J. 47:118-123.

Fusarium (contd.)

- 330 Snyder, W. C., and H. N. Hansen. 1945. The species concept in *Fusarium* with reference to discolor and other sections. *Amer. J. Bot.* 32:657-666.
- 331 South Australia. Waite Agricultural Research Institute. 1954. Report, 1952 - 1953. 70 p.
- 332 Stanghellini, M. E., and J. D. Russell. 1971. Induction of bacterial seed piece decay by various soil-borne fungi. *Phytopathology* 61:1324. (Abstr.)
- 333 Vanderwalle, R., and G. Roland. 1945. A contribution to the study of potato 'mildew' [in French]. *Parasitica* 1:41-57.
- 334 Vinot, M., and Bernaux, P. 1948. Potato charcoal rot in the Mediterranean region. (*Macrophomina phaseoli* (Maublanc) Ashby) [in French]. *Ann. Epiphyt.*, N.S. 14:91-102.
- 335 Wade, E. K. 1967. Potato seed piece treatments conducted in Wisconsin. *Amer. Potato J.* 44:341. (Abstr.)
- 336 Wager, R. M. 1953. Stolbur wilt of potato [in Russian]. *Mikrobiologiya* 22:198-202.
- 337 Washington. Agricultural Experiment Station. 1944. Annual report, 54, for the fiscal year ended 30th June, 1944. *Bull.* 455, 168 p.
- 338 Zimmermann-Griess, S. 1956. Keeping and planting qualities of sprouted seed potatoes. *Ktavim (Rec. Agr. Res. Sta. Rehovot, Israel)* 4(2):21-23.
- See also 18, 21, 23, 46, 71, 189, 201, 251, 256, 364, 365, 398, 527, 529, A4, A5, A6, A9, A11, A15, A16, A19, A20, A25, A30, A37, A39, A44, A45, A52, A57, A58, A59, A65, A66, A67, A70, A71, A72, A75, A76, A80, A91, A93, A94, A96, A103, A105.
- Fusarium*, Chemical control**
- 339 Arjunarao, V., and V. Kuznetso. 1972. Incidence of antagonistic actinomycetes against fusaria. *Curr. Sci.* 41:468.
- 340 Bonde, R., and F. Hyland. 1960. Effects of antibiotic and fungicidal treatments on wound periderm formation, plant emergence, and yields produced by cut seed potatoes. *Amer. Potato J.* 37:279-288.
- 341 Cunningham, H. S., and O. A. Reinking. 1946. *Fusarium* seedpiece decay of potatoes on Long Island and its control. *New York Agr. Exp. Sta. Bull.* 721.
- 342 Duncan, H. E., and M. E. Gallegly. 1963. Field trials for chemical control of seedpiece decay and blackleg of potato. *Amer. Potato J.* 40:279-284.
- 343 Fisher, K. D. 1962. The effects of seed piece treatment on pre-cut Norland and Red La Soda certified seed potatoes. *Diss. Abstr.* 22(1):21.
- 344 Guthrie, J. W. 1960. The influence of seed-piece treatment on disease control and yield of Russet Burbank potatoes. *Idaho Agr. Exp. Sta. Bull.* 329.
- 345 Hoyman, W. G. 1964. Treatment of potato seed tubers. *Proc. 3rd Annu. Wash. State Potato Veg. Conf. Moses Lake.* pp. 5-6.
- 346 Landis, B. J., J. A. Onsager, L. Fox, and L. L. Foiles. 1971. Chemical control of seed-corn maggot, *Hylemya platura* (Mergin) and seed-piece decay in potato seed pieces. *Amer. Potato J.* 48:374-380.
- 347 Leach, S. S. 1970. Evaluation of postharvest-prestorage fungicidal treatments for the control of *Fusarium* tuber rot of potatoes. *Phytopathology* 60:1299. (Abstr.)
- 348 Leach, S. S. 1971. Postharvest treatments for the control of *Fusarium* dry rot development in potatoes. *Plant Dis. Rep.* 55:723-726.
- 349 Line, R. F., and C. J. Eide. 1961. Chemical control of potato seed piece decay. *Amer. Potato J.* 38:388-395.
- 350 Lutman, B. F. 1937. Disinfectants and cut-seed potatoes. *Vermont Agr. Exp. Sta. Bull.* 418.
- 351 Lutz, J. M. 1953. *Fusarium* tuber rots of late potatoes as related to injuries and certain chemical treatments. *Amer. Potato J.* 30:131-134.
- 352 McKeen, C. D., and K. Slingsby. 1971. Evaluation of chemical seed treatments to control seed piece rot of potatoes. *Pestic. Res. Rep., Res. Br., Agr. Can.* p. 293.
- 353 Moore, W. C. 1949. Plant diseases in the United Kingdom. UNESCO, E/Conf. 7/Sec/W. 52. 8 p.
- 354 Nadvodnyuk, Yu. N. 1960. On the infection of potato by dry rot [in Ukrainian]. *Nauchn. Zap. Belotserkov. Sel'skokhoz. Inst.* 10:205-212.
- 355 Nadvodnyuk, Yu. N. 1962. The results of using *Trichoderma* for the control of dry rot of potato [in Ukrainian, English summary]. *Mikrobiol. Zh. (Kyiv)* 24(4):38-43.
- 356 Nelson, G. A., and W. E. Torfason. 1970. Effect of chemical treatment of potato seed pieces on yield. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 258-259.
- 357 Nelson, G. A., W. E. Torfason, H. T. Allen, and S. Molnar. 1972. Control of decay of fresh-cut and pre-cut potato seed pieces by chemical treatment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 278-280.
- 358 Nelson, G. A., W. E. Torfason, H. T. Allen, and S. Molnar. 1971. Potato seed-piece decay control by chemical treatment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 294-295.
- 359 Schultz, O. E. 1969. The nature and control of potato storage rots. *Cornell Exp. Sta. Ext. Bull.* 1218.
- 360 Sieczka, J. B. 1972. The effect of paraquat used as potato vine killer on sprouting and tuber susceptibility to decay. *Amer. Potato J.* 49:362. (Abstr.)
- 361 Slingsby, K., and C. D. McKeen. 1970. Potato seed piece decay control by chemical treatment and controlled environment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 259-260.
- Fusarium*, Control**
- 362 Altman, J. 1958. Studies on the control of plant diseases with antibiotics, with particular reference to streptomycin. *Diss. Abstr.* 19(2):201.
- 363 Baribeau, B. 1952. Verticillium wilt and seed potato certification. *Amer. Potato J.* 29:157-159.
- 364 Benza, J. C. 1944. Results of experimentation on potato cultivation [in Spanish]. *Circ. Estac. Exp. Agr., Lima.* 62, 89 p.
- 365 Ciferri, R. 1954. The use of zinc ethylenedisithiocarbamate for dry treatments [in Italian]. *Notiz. Malatt. Piante* 1953(27):7-14.
- 366 Krasil'nikov, N. A. ed. 1961. Applications of antibiotics in plant culture. *Trans. 1st All-Union Conference on the study and application of antibiotics in plant culture* [in Russian]. *Acad. Sci. Armenian S.S.R.* 274 p.
- 367 Nielsen, L. W., and J. T. Johnson. 1971. Infectious fusarial propagules on certified seed potatoes received in North Carolina. *Amer. Potato J.* 48:307. (Abstr.)
- 368 New York. Agricultural Experiment Station. Divisions of Plant Pathology and Seed Investigations. 1944. Report, 1942 - 1943. pp. 34-43, 53-58.
- 369 Potato Association of America. Annual meeting, 55th. 1971. *Amer. Potato J.* 48:295-308. (Abstr.)
- See also 376, 517, 520
- Fusarium*, Effect of crop rotation**
- 370 Emmond, G. S., and R. J. Ledingham. 1972. Effects of crop rotation on some soil-borne pathogens of potato. *Can. J. Plant Sci.* 52:605-611.
- Fusarium*, Effect of fungicides on**
- 371 Newton, W. 1952. Effects of the application of fungicides to wounded plant tissues. *Sci. Agr.* 32:659-662.
- Fusarium*, Effect of gamma radiation on**
- 372 Rubin, B. A. L. V. Metlitskii, E. G. Sal'kova, E. N. Mukhin, N. P. Korableva, and N. P. Morozova. 1959. The use of ionizing irradiations for the regulation of the dormancy of potato tubers in storage [in Russian]. *Biokhim. Plodov. Ovoshchei* 5:5-101.

Fusarium, Effect of gibberellic acid and gibberellin derivatives on

- 373 Petroczi, I., and A. Szabo. 1966. The role of gibberellic acid and gibberellin derivatives in spindle sprout formation on potato tubers [in German]. *Acta Phytopathol. Acad. Sci. Hung.* 1(3-4):269-276.

Fusarium, Factors affecting

- 374 Folsom, D., H. Q. Roach, J. S. Wiant, and J. Kaufman. 1952. Effect of storage and railroad transit on potato diseases, injuries, and shrinkage. *Maine Agr. Exp. Sta. Bull.* 507, 28 p.
- 375 Hollomon, D. W. 1967. Observation on the phylloplane flora of potatoes. *Eur. Potato J.* 10:53-61.
- 376 Smith, J. H. 1945. Plant pathological investigations. *Rep. Dep. Agr. Queensl.* 1944-45, 15-16.

Fusarium, Fungicides ineffective against

- 377 Kuz'mina, G. N. 1969. Results of testing fungicides in the control of diseases of potato [in Russian]. *Tr. Kaz. Sel'skokhoz. Inst.* 2:203-209.

Fusarium, Legislation against

- 378 Argentina. Ministry of Agriculture. Plant Health. 1952. *Bulletin* 2:247-258. [in Portuguese].
- 379 Legislative and administrative measures. 1937. *Int. Bull. Plant Prot.* 11(2):30-31, 37.
- 380 Legislative and administrative measures. Sweden. 1935. *Int. Bull. Plant Prot.* 9(5):115-116.
- 381 United States Department of Agriculture. Bureau of Entomology and Plant Quarantine. 1936. Amendment of regulations governing the entry of potatoes into the United States. 2 p.

Fusarium, Pathogenicity of

- 382 Gradinarov, L. 1959. On the pathogenicity of *Phytophthora infestans*, some *Fusarium* spp., and *Rhizopus nigricans* to potato tubers [in Bulgarian, Russian, German summaries]. *Izv. Inst. Biol. Bulg. Akad. Nauk.* 8:223-242.

Fusarium, Resistance to

- 383 Seminario, B., E. R. French, and L. W. Nielsen. 1970. Tuber resistance to *Fusarium* spp. affecting potatoes in Peru [in Spanish, English summary]. *Amer. Potato J.* 47:118-123.

Fusarium, Varietal reaction to

- 384 Benlloch, M. 1965. Tests carried out on potato dry rot or fusariosis [in Spanish]. *Bol. Patol. Veg. Entomol. Agr.* 28:85-118.
- 385 Cunningham, H. S. 1953. A histological study of the influence of sprout inhibitors on *Fusarium* infection of potato tubers. *Phytopathology* 43:95-98.
- 386 Kochetova, Z. M. 1959. How to control fusarioses [in Russian]. *Potato, Moscow*, 4(2):56-57.

Fusarium coeruleum

- 387 Ayers, G. W. 1970. Potato seed and soil treatment for disease control. *Proc. Can. Phytopathol. Soc.* 36:21. (Abstr.)
- 388 Ayers, G. W. 1961. The susceptibility of potato varieties to storage rots caused by *Fusarium sambucinum* Fckl. F6 Wr. and *Fusarium caeruleum* (Lib) Sacc. *Can. Plant Dis. Surv.* 41:170-171.
- 389 Ayers, G. W. 1962. The susceptibility of potato varieties to storage rots caused by *Fusarium sambucinum* F6 and *Fusarium caeruleum* and to wilts caused by *Verticillium albo-atrum* in 1961. *Can. Plant Dis. Surv.* 42:115-117.
- 390 Ayers, G. W., and J. N. Richard. 1972. Potato seed treatment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 280-282.
- 391 Ayers, G. W., and D. B. Robinson. 1954. An inoculation technique for the study of dry rot of potatoes. *Amer. Potato J.* 31:278-281.
- 391a Black, W. 1947. Blight in relation to potato breeding. *Ann. Appl. Biol.* 34:631-633.
- 392 Blodgett, E. C. 1945. Potato diseases in Idaho in 1943. *Plant Dis. Rep.* 29:51-57.

- 393 Bonde, R. 1932. Summary report of progress, 1934. *Maine Agr. Exp. Sta. Bull.* 363:279.
- 394 Bonde, R. 1934. Summary report of progress, 1934. *Maine Agr. Exp. Sta. Bull.* 377:358-360.
- 395 Bonde, R. 1935. Summary report of progress, 1935. *Maine Agr. Exp. Sta. Bull.* 380:169-170.
- 395a Boyd, A. E. W. 1947. Some recent results of potato dry rot research. *Ann. Appl. Biol.* 34:634-636.
- 396 Boyd, A. E. W. 1966. Some factors associated with tuber susceptibility to potato dry rot caused by *Fusarium caeruleum*. *Proc. Third Trienn. Conf. Eur. Assoc. Potato Res. Abstr. in Rev. Plant Pathol.* 50:153. 1971.
- 397 Boyd, A. E. W. 1967. The effects of length of the growth period and of nutrition upon potato-tuber susceptibility to dry rot (*Fusarium caeruleum*). *Ann. Appl. Biol.* 60:231-240.
- 398 Brichet, J. 1944. Preservation of main crop potatoes. "Fusariosis" or "dry rot". The application of refrigeration [in French]. *Fruits & Primeurs* 14(150):156-159. (*Rev. Appl. Mycol.* 24:245)
- 399 Brook, M. 1957. Tetrachloronitrobenzene as a fungicide. *Trans. Brit. Mycol. Soc.* 40:164-165. (Abstr.)
- 400 Bustamante, R. E., and H. D. Thurston. 1964. "Hard rot" of the potato tuber [in Spanish]. *Agr. Trop.* 21:113-121.
- 401 Chona, B. L. 1932. Studies in the physiology of parasitism. XIII. An analysis of the factors underlying specialization of parasitism, with special reference to certain fungi parasitic on apple and potato. *Ann. Bot.* 46(184):1033-1050.
- 402 Delaney, D., and P. Keenan. 1944. The building up and maintenance of healthy stocks of seed potatoes. A review of the work. *J. Dep. Agr., Eire* 41(1):95-105.
- 403 Detilleux, E. 1958. Potato growing in the Elisabethville area [in French]. *Bull. Inform. Inst. Etud. Agron., Congo Belge*, 7(5):323-338.
- 404 Dillon Weston, W. A. R., and R. E. Taylor. 1944. Blight. *Agriculture, J. Min. Agr. (G.B.)* 51:111-116.
- 405 Fehmi, S. 1933. Contributions to the knowledge of the interrelations between cultivated plants, their parasites, and the environment. Note 5. Investigations on the influence of nutrition on the susceptibility of the potato tuber to storage parasites and the changes in the course of enzymatic metabolism during storage [in German]. *Phytopathol. Z.* 6:543-588.
- 406 Finland. Department of Plant Pathology of Agricultural Research Centre. 1951. The most important diseases of crop plants in Finland. *Maatalousk. Kasvitaustiasat. Tiedon.* 5, 10 p. (Mimeo).
- 407 Foister, C. E. 1940. Dry rot diseases of potatoes. *Scott. J. Agr.* 23(1):7.
- 408 Foister, C. E., A. R. Wilson, and A. E. W. Boyd. 1945. Control of dry rot of seed potatoes by dusting. *Nature* 156:394.
- 409 Foister, C. E., A. R. Wilson, and A. E. W. Boyd. 1952. Dry-rot diseases of the potato. I. Effect of commercial handling methods on the incidence of disease. *Ann. Appl. Biol.* 39:29-37.
- 410 Foister, C. E., A. R. Wilson, and A. E. W. Boyd. 1945. Potato dry rot and gangrene as soil-borne diseases. *Nature* 155:793-794.
- 411 Gorodetskii, V. S. 1971. The effect of temperature on the resistance of different potato varieties to dry rot [in Russian]. *Tr. Nauchno. Issled. Inst. Kartof.* 8:228-232.
- 412 Goss, R. W. 1934. A survey of potato scab and *Fusarium* wilt in western Nebraska. *Phytopathology* 24:517-527.
- 413 Haritonova, Z. M. 1954. An experiment on the control of transpiration and diseases of potatoes in storage by means of covering them with table beet [in Russian]. *Tr. Vses. Inst. Zashch. Rast.* 5:139-143.
- 414 Harrison, D. E., and W. A. Downie. 1960. Phoma and dry rot of potatoes. Progress report on investigations in Victoria. *J. Agr. Victoria, Aust.*, 58:372-373, 375-377, 379-381, 385.
- 414a Hawkes, J. G. 1947. Some observations on South American potatoes. *Ann. Appl. Biol.* 34:622-631.

Fusarium coeruleum (contd.)

- 415 Hellinga, J. J. A. 1940. On the effect of substances, produced by fungi, on the respiration of the tissue of potato tubers. I and II. Verh. Akad. Wet., Amst., viii;30 p.
- 416 Hooker, W. J., C. J. Kim, and H. S. Potter. 1972. Fungicide redistribution on potato leaves. Amer. Potato J. 49:369. (Abstr.)
- 417 Hopkins, J. C. F. 1936. Annual report of the Senior Plant Pathologist for the year ending 31st December, 1936. Rhodesian Agr. J. 33:413-421.
- 418 Jamalainen, E. A. 1944. On the fusaria of Finland [in German]. Valtion Maatalous, Julk. 122, 26 p.
- 419 Kraus, J. E., and G. W. Woodbury. 1943. Prevention of potato seedpiece decay in southern Idaho. Amer. Soc. Hort. Sci. 43:262-264.
- 420 Lhoste, L. 1946. On fusariosis of potato [in French]. Rev. Hort., Paris, 30(6):103-104.
- 421 Limasset, P. 1949. Progress made in the study of plant diseases since 1937 [in French]. Viticult. Arboricult., Paris, 95(10):288-291.
- 422 Lutz, J. M., R. C. Wright, and A. D. Edgar. 1948. Research on harvesting, transportation and storage of potatoes - a review of recent literature. Amer. Potato J. 25:437-445.
- 423 Lutz, L. 1940. On the gummy degeneration of potato tubers [in French]. C.R. Acad. Agr. Fr. 26(19):664-668.
- 424 Mattingley, G. H. 1951. The seed potato certification scheme. J. Dep. Agr. Victoria, Aust., 49(4):177-180.
- 425 McIntosh, T. P. 1944. Potato troubles. Card. Chron., Ser. 3. 116(3010):87-88.
- 426 McKee, R. K. 1951. Mutations appearing in *Fusarium caeruleum* treated with tetrachloronitrobenzene. Nature 167(4250):611.
- 427 McKee, R. K. 1952. Dry-rot disease of the potato. II. Fungi causing dry rot of seed potatoes in Britain. Ann. Appl. Biol. 39:38-43.
- 428 McKee, R. K. 1954. Dry-rot disease of potato. VIII. A study of the pathogenicity of *Fusarium caeruleum* (Lib.) Sacc. and *Fusarium avenaceum* (Fr.) Sacc. Ann. Appl. Biol. 41:417-434.
- 429 McKee, R. K. 1955. Host - parasite relationships in the dry-rot disease of potatoes. Ann. Appl. Biol. 43:147-148.
- 430 McKee, R. K., and A. E. W. Boyd. 1952. Dry-rot disease of the potato. III. A biological method of assessing soil infectivity. Ann. Appl. Biol. 39:44-53.
- 431 McKee, R. K., and A. E. W. Boyd. 1962. Dry-rot disease of the potato. IX. The effect of diphenyl vapour on dry-rot infection of potato tubers. Ann. Appl. Biol. 50:89-94.
- 432 Melhus, I. E., D. R. Shepherd, and M. A. Corkle. 1941. Diseases of potatoes in Iowa. Proc. Iowa Acad. Sci. 48:133-146.
- 433 Mooi, J. C. 1950. The *Fusarium* rot or dry rot of potatoes [in Dutch]. Landbouwk. Tijdschr. 62:712-725.
- 434 Müller, K. O., and H. Börger. 1949. Experimental studies on the *Phytophthora* resistance of the potato, together with a contribution to the problem of "acquired resistance" in the plant kingdom [in German]. Arb. Biol. Aust. (Reichsanst.), Berlin, 23(2):189-231.
- 435 New York. Agricultural Experiment Station. 1947. Plant pathology: report:1945 - 1946. pp. 37-48.
- 436 New York. Agricultural Experiment Station. Divisions of Plant Pathology and Seed Investigations. 1945. Report. 1944 - 1945. pp. 40-52, 59-63.
- 437 New York. Agricultural Experiment Station. Divisions of Plant Pathology and Seed Investigations. 1947. Report 1945 - 1946. pp. 37-48.
- 438 Ollila, L. 1947. On the significance of fungous diseases in stored potatoes in Finland [in Finnish, English summary]. Maataloust. Aikak. 19:89-98.
- 439 Pethybridge, G. H. 1934. Potato diseases. J. Min. Agr. Brit. 41:125-136.
- 440 Pitt, D., and C. Coombes. 1969. Release of hydrolytic enzymes from cytoplasmic particles of *Solanum* tuber tissues during infection by tuber-rotting fungi. J. Gen. Microbiol. 56:321-329.
- 442 Anon. 1949. The protection of stored seed potatoes. World Crops 1:63-65.
- 443 Radtke, W. 1969. Defence reaction of potato tissue against *Fusarium coeruleum* with special consideration of the osmotic pressure [in German]. Phytopathol. Z. 64:143-174.
- 444 Røll-Hansen, F. 1950. Investigation of *Fusarium* on cultivated plants in Norway. Meld. Norg. Landbruksheisk. 29:257-264.
- 445 Ruehle, G. D. 1944. Outstanding potato late blight control in Florida with a new organic fungicide combined with zinc sulfate. Plant Dis. Rep. 28:242-245.
- 446 Samson, R. W., and N. K. Ellis. 1943. Influence of time of planting of potatoes in Indiana muck soil on yield and scab development. Amer. Potato J. 20(12):301-308.
- 447 Sardina, J. R. 1945. Potato diseases [in Spanish]. Publ. Estac. Fitopatol. Agr. Coruna 5. viii, 111 p.
- 448 Schultz, E. S., R. Bonde, and W. P. Raleigh. 1934. Components of potato mild mosaic. Phytopathology 24:17. (Abstr.)
- 449 Seed testing and plant registration. Scott. J. Agr. 21:54-70. 1938.
- 450 Seed testing and plant registration. Scott. J. Agr. 22:38-54. 1939.
- 451 Singh, R. K. 1958. *Fusarium moniliforme* Sheld. causing rot of vegetables. Indian Phytopathol. 11:189-191.
- 452 Small, T. 1945. Dry rot of potato (*Fusarium caeruleum* (Lib.) Sacc.). Investigation on the sources and time of infection. Ann. Appl. Biol. 31:290-295.
- 453 Snyder, W. C., and H. N. Hansen. 1941. The species concept in *Fusarium* with reference to section *martiella*. Amer. J. Bot. 28:738-742.
- 454 Wallace, M. M. 1943. Diseases of potatoes. Mycol. Leaflet. Dep. Agr. Tanganyika, 17, 10 p.
See also 270, 293, 301, 371, 462, 501, A14, A43, A51, A83, A98

Fusarium coeruleum, Breeding against

- 455 Cambridge. Plant Breeding Institute. 1963. Annual report for 1961-62. 82 p.

Fusarium coeruleum, Chemical control

- 456 Chemical prevention of potato dry rot disease. Nature 162:843. 1948.
- 457 Votoupal, B. 1966. Some experiences with storing potato tubers infested by *Phytophthora infestans*, using tetrachloronitrobenzene [in Russian, English summary]. Ochr. Rost., N.S. 2:227-234.

Fusarium coeruleum, Control

- 457a Ayers, G. W., and J. N. Richard. 1971. Potato seed treatment. Pestic. Res. Rep., Res. Br., Agr. Can. pp. 297-299.
- 458 Brief reports on the plant pathological work carried out in the stations and laboratories in 1943 and 1944 [in French]. Ann. Epiphyt. 11(3-4):245-247, 249-251. 1945.
- 459 Brook, M., and C. G. C. Chesters. 1957. The use of tetrachloronitrobenzene isomers on potatoes. Ann. Appl. Biol. 45:623-634.
- 460 Choudhuri, H. C. 1956. Storage tests for the control of diseases and insect pests. Amer. Potato J. 33:6-14.
- 461 Cunningham, H. S., and O. A. Reinking. 1945. *Fusarium* seed-piece decay of potatoes on Long Island. Farm Res., N.Y., 11(3):8-9.
- 462 Foister, C. E., and A. R. Wilson. 1943. Dry rot in seed potatoes. A summary of some recent experiments. Agriculture, J. Min. Agr. (G.B.) 50:300-303.
- 463 Foister, C. E., and A. R. Wilson. 1950. Dry rot of potatoes. Agriculture, J. Min. Agr. (G.B.) 57:229-233.
- 464 Graham, D. C. 1964. The use of organo-tin compounds as potato tuber disinfectants, particularly against *Rhizoctonia solani*. Eur. Potato J. 7:33-44.

***Fusarium coeruleum*, Control (contd.)**

- 465 Guillemat, J., D. Lelièvre, and J. Montgut. 1952. Trials of fungicides against potato fusariosis [in French]. *Pomme d. Terre franc.* 15(152):3-9.
- 466 Martinovic, M. 1961. Dry rot of potato [in Croatian, English summary]. *Zast. Bilja (Plant Prot., Belgrad)*, 1961, 63-64:21-26.
- 467 Samuel, G. G. 1948. The control of potato diseases. *J. Roy. Agr. Soc. Engl.* 109:118-127.
- 468 Schippers, P. A. 1962. Dry rot of the potato; preliminary publication. *Eur. Potato J.* 5:132-146.
- 469 Servazzi, O. 1954. Dry rot of the potato [in Italian]. *Ital. Agr.* 4, 6 p.
- 470 Small, T. 1945. The effect of disinfecting and bruising seed potatoes on the incidence of dry rot (*Fusarium coeruleum* (Lib.) Sacc.). *Ann. Appl. Biol.* 32:310-318.
- 471 Tandon, R. N., and G. P. Agarwal. 1956. Dry rot of *Colocasia antiquorum* and potato caused by *Fusarium coeruleum* (Lib.) Sacc. *Allahabad Univ. Stud., Bot.* 26 p.
- 472 Todd, J. M. 1963. The development and control of potato dry rot, gangrene and skin spot. *Seed Potato* 3(1):14-18.
- 473 Wilkins, V. E. 1959. Third report of the working party on phytosanitary regulations (Bad Godesberg, 18-20 March, 1959). *Eur. Medit. Plant Prot. Organ., Paris.* 23 p.
- 474 Wilson, A. R. 1963. Dry rot of potatoes. *Adv. Leaflet.* 218, Min. Agr. Fish. Lond., 5 p.
- 475 Wilson, A. R. 1967. Dry rot of potatoes. *Adv. Leaflet.* 218, Min. Agr. Fish. Lond., 6 p.

***Fusarium coeruleum*, Effect of, on yield**

- 476 Nattrass, R. M. 1944. Potato blight. *East. Afr. Agr. J.* 10(1):18-21.

***Fusarium coeruleum*, Effect of disinfecting and bruising on**

- 477 Small, T. 1946. Further studies on the effect of disinfecting and bruising seed potatoes on the incidence of dry rot (*Fusarium coeruleum* (Lib.) Sacc.). *Ann. Appl. Biol.* 33:211-219.

***Fusarium coeruleum*, Factors affecting**

- 478 European Association for Potato Research. 1966. *Proc. 3rd Triennial Conf., Zürich, 4-10 Sept., 1966. Wageningen.* 326 p.
- 479 Kranz, J. 1959. Influence of the pre-temperature on the pathogenicity of some fungi and their growth in vitro [in German, English summary]. *Phytopathol. Z.* 37:159-163.
- 480 Kranz, J. 1959. On the varietal susceptibility of the potato tuber *Fusarium coeruleum* and *Phoma foveata* and the influence of the place of cultivation [in German, English summary]. *Phytopathol. Z.* 35:135-147.
- 481 Schippers, P. A. 1955. Some factors influencing the keeping quality of potatoes. *Neth. J. Agr. Sci.* 3:305-310.
- 482 Schneider, R. 1953. Studies on the humidity requirements of parasitic fungi [in German]. *Phytopathol. Z.* 21:63-78.
- 483 Vitukevich, E. R. 1958. A study of the conditions for the development of dry rot in potato storage (in Ukrainian). *Nauchn. Tr. Ukr. Inst. Ovoshch. Kartof.* 4:257-263.

See also 469

***Fusarium coeruleum*, Nutritional study of**

- 484 Agarwal, G. P. 1957. Sulphur and phosphorus nutrition of two strains of *Fusarium coeruleum* (Lib.) Sacc. *Phyton* 8:43-51.

***Fusarium coeruleum*, Pathogenicity of**

- 485 Gorodetskii, V. S. 1970. Investigations on the specific composition and pathogenicity of causal agents of dry rot potato tubers under conditions of the Moscow region [in Russian]. *Tr. Nauchn. Issled. Inst. Kartof. Kh.-va.* 7:147-150.
- 486 Langerfeld, E. 1971. Differential characteristics in the pathogenicity of two potato dry rot agents of the genus *Fusarium* [in German, English summary]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst., Stuttgart*, 23(11):168-169.

***Fusarium coeruleum*, Phenol metabolism of**

- 487 Baruah, P. 1964. Investigations on the phenol metabolism of certain potato rotting fungi. I. *Fusarium coeruleum*. *J. Univ. Gauhati*, 15:165-189.

***Fusarium coeruleum* on, Physiology of**

- 488 Indian Science Congress. 37th. Poona. 1950. Mycology and plant pathology, *Agric. Sci. Proc. Pt. III*, pp. 56-59, 75-92

***Fusarium coeruleum*, Susceptibility to**

- 489 Schoene, K. 1967. Studies on the susceptibility of potato tubers to *Fusarium coeruleum*, the causal agent of white rot [in German, English summary]. *Phytopathol. Z.* 60:201-236.

***Fusarium coeruleum*, Testing resistance**

- 490 Ireland. Department of Crop Husbandry, Plant Breeding and Plant Pathology. 1970. Research report. *Plant Sci. Crop Husband., Dublin* 1969, pp. 6-17, 18-36, 37-46.

***Fusarium coeruleum*, Transmission of, by soil**

- 491 Boyd, A. E. W. 1970. Transmission of dry rot (*Fusarium coeruleum*) by seed tubers and by soil. Pages 192-193 in *Proc. Fourth Trienn. Conf. Eur. Assoc. Potato Res. Brest*, 8-13 Sept. 1969. 298 p.
- 492 Edinburgh. School of Agriculture. 1966. Experimental work. 114 p.
- 493 Foister, C. E., A. R. Wilson, and A. E. W. Boyd. 1945. Potato dry rot and gangrene as soil-borne diseases. *Nature* 155(3948):793-794.

***Fusarium coeruleum*, Varietal reaction to**

- 494 Ayers, G. W. 1955. The resistance of potato varieties to storage decay caused by *Fusarium sambucinum* f.6 and *Fusarium coeruleum*. *Amer. Potato J.* 33:249-254.
- 495 Ayers, G. W., and D. B. Robinson. 1956. Control of *Fusarium* dry rot of potatoes by seed treatment. *Amer. Potato J.* 33:1-5.
- 496 Boyd, A. E. W. 1952. Dry-rot diseases of the potato. IV. Laboratory methods used in assessing variations in tuber susceptibility. V. Seasonal and local variations in tuber susceptibility. VI. Varietal differences in tuber susceptibility obtained by infection and riddle-abrasion methods. VII. The effect of storage temperature upon subsequent susceptibility of tubers. *Ann. Appl. Biol.* 39:322-357.
- 497 Cambridge. National Institute of Agricultural Botany. Report, 33rd, 1952.
- 498 Cambridge. Plant Breeding Institute. 1971. Annual report. 160 p.
- 499 Foister, C. E. 1964. The development of new varieties of the potato. *Trans. Roy. Highland Agr. Soc. Scot.* 1963. 19 p.
- 500 Knorr, L. C. 1943. Ring rot of potatoes. *Ext. Cornell Agr. Exp. Sta. Bull.* 620 (War Emergency Bull. 113). 4 p.
- 501 Lansade, M. 1949. Researches on fusariosis or dry rot of the potato, *Fusarium coeruleum* (Lib.) Sacc [in French]. *Bull. Tech. Inform. Ingln. Serv. Agr.* 41p.
- 502 Maurer, A. R., M. Van Andrichem, D. A. Young, and H. T. Davies. 1968. Cariboo, a new late potato variety of distinctive appearance. *Amer. Potato J.* 45:247-249.
- 503 Moore, F. J. 1945. A comparison of *Fusarium avenaceum* and *Fusarium coeruleum* as causes of wastage in stored potato tubers. *Ann. Appl. Biol.* 32:304-309.
- 504 Scotland. Department of Agriculture. 1948. New varieties of the potato introduced since 1939. *Leaflet.* 2, 8 p.
- 505 Small, T. 1946. Dry rot of potato (*Fusarium coeruleum* (Lib.) Sacc.) Effect of planting infected and contaminated sets on plant establishment. *Ann. Appl. Biol.* 33:219-221.

See also 88, 146, 466, 469, 474, 486

Fusarium culmorum

- 506 Skinner, F. A. 1953. Inhibition of *Fusarium culmorum* by *Streptomyces albidoflavus*. *Nature* 172(4391):1191.

Fusarium oxysporum

- 507 Dubey, H. D. 1959. Relation between nitrogen, phosphorus, and potassium fertilization and incidence of stem rot disease of sweet potato. *Sci. Cult.* 25:139-140.
- 508 Eddins, A. H. 1940. Potato seedpiece rot caused by *Fusarium oxysporum*. *Phytopathology* 30:181-182.
- 509 Peters, E. J. 1943. Stem-end vascular discoloration of potatoes due to *Fusarium oxysporum* F. *tuberosi*. *Amer. Potato J.* 20:10-12.
- 510 Qureshi, A. A., and O. T. Page. 1969. Observations on the perfect stages of *Fusarium oxysporum* and *Fusarium solani*. *Proc. Can. Phytopathol. Soc.* 36:18. (Abstr.)
- 511 Upstone, M. E. 1970. A corky rot of Jersey Royal potato tubers caused by *Fusarium oxysporum*. *Plant Pathol.* 19:165-167.

Fusarium solani

- 513 Cochrane, J. C., V. W. Cochrane, F. G. Simon, and J. Spaith. 1963. Spore germination and carbon metabolism in *Fusarium solani*. I. Requirements for spore germination. *Phytopathology* 53:1155-1160.
- 514 Goss, R. W., and J. E. Livingston. 1941. The influence of crop rotations on the occurrence of scab, Rhizoctonia, and Fusarium wilt in potatoes under dry-land conditions in western Nebraska. *Rep. Nebr. Potato Improv. Assoc.* 22:22-27.
- 515 Krantz, F. A., A. G. Tolaas, H. O. Werner, H. W. Goss, and J. H. Jensen. 1943. The Kasota potato. *Amer. Potato J.* 20:25-27.
- 516 Livingston, J. E. 1942. The present status of bacterial ring rot. *Rep. Nebr. Potato Improv. Assoc.* 23:9-12.
- 517 Matuo, T., and W. C. Snyder. 1973. Use of morphology and mating populations in identification of formae speciales in *Fusarium solani*. *Phytopathology* 63:562-565.
- 518 Michail, S. H., H. Elarosi, and E. A. Khairy. 1969. Dry rot of *Colocasia antiquorum* in U.A.R. (Egypt). *J. Phytopathol. UAR* 1:23-26.
- 519 Mitra, A. 1934. A study of certain fusaria. *J. Indian Bot. Soc.* 13(4):255-268.
- 520 Murdoch, A. W., and R. K. S. Wood. 1972. Control of *Fusarium solani* rot of potato tubers with fungicides. *Ann. Appl. Biol.* 72:53-62.
- 521 Pawuk, W. H., and F. A. Wood. 1972. Influence of sugar maple stem tissue extracts on spore germination by *Fusarium solani*. *Plant Dis. Rep.* 56:944.
- 522 Pett, B., and M. Effmert. 1972. Symptoms of artificially induced mixed infections on potato tubers by *Phoma solanicola* Prill & Del. and *Fusarium coeruleum* (Lib.) Sacc [in German]. *Zentralbl. Bakteriol. Parasitenk. Infektionskr. Hyg. Abt. B* 2(1972), 127(3):227-231.
- 523 Snyder, W. C. 1934. Notes on fusaria of the section Martiella. *Zentralbl. Bakteriol.* 91(8-10):163-184.
- See also 188, 482, 483, 485, 510, A90

Fusarium sporotrichioides

- 524 Upstone, M. E. 1970. A potato tuber rot caused by *Fusarium sporotrichioides* Sherb. *Plant Pathol.* 19:148-150.

Fusarium sulphureum

- 525 Boyd, A. E. W., and J. H. Tickle. 1972. Dry rot of potato tubers caused by *Fusarium sulphureum* Schlecht. *Plant Pathol.* 21:195.

Storage diseases

- 526 Alvarado, E. L. F., and N. J. Guzman. 1969. Potato decay in storage. *Amer. Potato J.* 46:27.
- 527 Boyd, A. E. W. 1972. Potato storage diseases. *Rev. Plant Pathol.* 51:297-321.
- 528 Blodgett, E. C. 1947. Comments on black rot, a storage disease of potatoes in Idaho. *Plant Dis. Rep.* 31:10-13.
- 529 Cunningham, H. H., M. V. Zaehring, and W. C. Sparks. 1971. Storage temperature for maintainance of internal quality in Idaho Russet Burbank potatoes. *Amer. Potato J.* 48:320-328.

- 530 Eide, C. J. 1965. The effect of storage temperature and other factors on decay of potato seed pieces. *Plant Dis. Rep.* 49:638-640.
- 531 Mikula, J. 1953. Storage losses of potatoes and their control [in Finnish, English summary]. *Valtion Maatalousk. Julk.* 137:39.
- See also 14, 216, 413, 442, 483, 496, A10, A53, A84, A92, A95

Addendum

- A1 Abdel-Rehim, M.A., and M.A. El-Goorani. 1973. Properties of *Erwinia atroseptica* and *Erwinia carotovora*. *Zentralbl. Bakteriol. Parasitenk. Infektionskr. Hyg.* 128:660-667.
- A2 Adams, M.J. 1975. Potato tuber lenticels: development and structure. *Ann. Appl. Biol.* 79:265-273.
- A3 Adams, M.J. 1975. Potato tuber lenticels: susceptibility to infection by *Erwinia carotovora* var. *atroseptica* and *Phytophthora infestans*. *Ann. Appl. Biol.* 79:275-282.
- A4 Ali, S.A., D.C. Nelsen, and J.P. Freeman. 1975. Suberization and periderm development in Norchief and Red Pontiac potatoes. *Amer. Potato J.* 52:201-209.
- A5 Ayers, G.W. 1974. Potato seed treatment for the control of verticillium wilt and fusarium seed decay. *Can. Plant Dis. Surv.* 54:74-76.
- A6 Ayers, G.W., and J.N. Richard. 1973. Potato seed treatment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 318-319.
- A7 Beczner, J., B.M. Lund, and C.E. Bayliss. 1975. The occurrence of rishitin, phytuberin, lubimin, and spirovetia-1 (10), 11-dien-2-one in potato tubers inoculated with *Erwinia carotovora* var. *atroseptica* or with *Phytophthora infestans*. *Proc. Amer. Phytopathol. Soc.* 2:50. (Abstr.)
- A8 Berlinski, K. 1972. Studies on the infectivity and pathogenicity of *Erwinia carotovora* var. *atroseptica* [In Polish, English summary]. *Biul. Inst. Ochr. Rost.* 52:179-185.
- A9 Bhagwat, V.Y. 1973. Production of pectinolytic and cellulolytic enzymes by *Fusarium* spp. associated with storage rots. *Res. J. Mahatma Phule Agr. Univ.* 4:113-117.
- A10 Bhagwat, V.Y. 1973. Storage rot of potato in Maharashtra. *Res. J. Mahatma Phule Agr. Univ.* 4:113-117.
- A11 Bhattacharyya, S.K., and V.K. Bahal. Potato seed piece decay - its control. *Ind. Phytopathol.* 25:555-557.
- A12 Biehne, W.L., D.C. Sands, and L. Hankin. 1972. Relationship between per cent dry matter content of potato tubers and susceptibility of bacterial soft rot. *Phytopathology* 62:747. (Abstr.)
- A13 Biehne, W.L., D.C. Sands, and L. Hankin. 1972. Repression of pectic enzymes and pathogenesis in *Erwinia carotovora*. *Phytopathology* 62:747. (Abstr.)
- A14 Bommer, D., and C. Patzold. 1972. Experiments under control of potato tuber rotting by *Fusarium coeruleum* [In German]. *Landbauforsch. Volkenrode* 22:123-128.
- A15 Borchert, R., J.D. McChesney, and D. Watson. 1974. Would healing in potato tuber tissue. Phosphon inhibition of developmental processes requiring protein synthesis. *Plant Physiol.* 53:187-191.
- A16 Boyd, A.E.W. 1947. Some recent results of potato dry rot research. *Ann. Appl. Biol.* 34:634-636.
- A17 Brady, R.J., E.C.S. Chan, and M.J. Pelczar, Jr. 1961. Sporulation of *Bacillus sphaericus* grown in association with *Erwinia atroseptica*. *J. Bacteriol.* 81:725-729.
- A18 Brenner, D.J., A.G. Steigerwalt, G.V. Miklos, and G.R. Fanning. 1973. Deoxyribonucleic acid relatedness among *Erwiniae* and other Enterobacteriaceae: the soft rot organisms (genus *Pectobacterium* Waldee). *Int. J. System. Bacteriol.* 23:205-216.

Addendum (contd.)

- A19 Burth, U., G. Motte, H. Stachewicz, G. Brazda, B. Pett, E. Becker, and R. Kloss. 1974. On the development of seed potato dressing - a new technique for stabilizing industrial potato production [In German, English summary]. *Nachrichtbl. Dtsch. Pflanzenschutzdienst* 28:153-158.
- A20 Chamber, S.C., and J.R. Millington. 1974. Studies on *Fusarium* species associated with a field planting of pathogen-tested potatoes. *Austral. J. Agr. Res.* 25:293-297.
- A21 Ciampi, P.L., and M.S. Gonzalez. 1973. Taxonomic study of causal agents of wet rots in potato plants [In Spanish]. *Agr. Tec. (Chile)* 33:16-20.
- A22 Colhoun, J. 1973. Effects of environmental factors on plant disease. *Annu. Rev. Phytopathol.* 11:343-364.
- A23 Coleno, A., and F. Rapilly. 1967. Study of the behaviour of two phytopathogenic bacteria in steam-sterilized soil. [In French]. *Phytiat.-Phytopharm.* 16:157-164.
- A24 Cromarty, R.W., and G.D. Easton. 1973. The incidence of decay and factors affecting bacterial soft rot of potatoes. *Amer. Potato J.* 50:398-407.
- A25 Darozhkin, M.A., and V.T. Mikhail'chik. 1975. Biological properties of the causal agent of dry rot of potato tubers, *Fusarium sambucinum* Fuck. var. *minus* Wr. [In Belorussian]. *Vestsi Akad. Navuk BSSR Biyalagich. Navuk No.* 3:59-63.
- A26 Davis, J.R. 1973. Seed and soil treatments of control of Rhizoctonia and blackleg of potato. *Plant Dis. Rep.* 57:803-806.
- A27 DeBoer, S.H., and A. Kelman. 1975. Evaluation of procedures for detection of pectolytic *Erwinia* spp. on potato tubers. *Amer. Potato J.* 52:117-123.
- A28 Dobias, K. 1973. Laboratory methods of testing the resistance in potato to *Erwinia carotovora* (Jones) Holland. [In Czech., English summary]. *Ochr. Ros.* 9:119-124.
- A29 Dobias, K. 1973. The serological relationship of strains of *Erwinia carotovora* (Jones) Holland isolated from potato. [In Czech., English summary]. *Res. Vyr.* 19:277-284.
- A30 El-Goorani, M.A., and M.K. El-Kazzaz. 1975. Occurrence of blackleg and dry rot of potato in Egypt through imported tubers. *Plant Dis. Rep.* 59:171-174.
- A31 Erinle, I.D. 1975. Blackleg of potatoes: induction through tuber inoculation. *Plant Pathol.* 24:172-175.
- A32 Ficke, W., K. Naumann, K. Skadow, H.J. Müller, and R. Zielke. 1973. Longevity of *Pectobacterium carotovora* var. *atrosepticum* (Van Hall) Dowson on seed material and in soil [In German, English summary]. *Arch. Pflanzenschutz.* 9:281-293.
- A33 Ficke, W., K. Skadow, H.J. Müller, K. Naumann, and R. Zielke. 1973. Survival of *Pectobacterium carotovorum* var. *atrosepticum* (Van Hall) Dowson on machinery and materials [In German, English summary]. *Arch. Pflanzenschutz.* 9:371-381.
- A34 Geesteranus, H.P.M. 1972. Wet rot and blackleg of potatoes [In Dutch]. *Bedrijfsontwikkeling* 3:941-945.
- A35 Geesteranus, H.P.M. 1975. Host - plant parasite relationships of *Erwinia carotovora* group causing soft rot of potato plants and tubers [In Dutch]. *Acta Bot. Neerl.* 24:249.
- A36 Graham, D.C., and M.D. Harrison. 1975. Potential spread of *Erwinia* spp. in aerosols. *Phytopathology* 65:739-741.
- A37 Guzman, V.L. 1974. Potato seed piece decay control. *Res. Rep. Belle Glade Agr. Res. Educ. Center, Fla. Univ.* 5 p.
- A38 Hall, J.A., and R.K.S. Wood. 1974. Permeability changes in tissues and other effects of cell-separation solutions from soft rots caused by *Corticium praticola* and *Erwinia atroseptica*. *Ann. Bot.* 38:129-140.
- A39 Huguelet, J.E. 1975. Synergism between *Erwinia atroseptica* and *Fusarium roseum* in blackleg and seedpiece decay development. *Amer. Potato J.* 52:245. (Abstr.)
- A40 Iriarte, M.T., and P. Fernandez. 1968. Behaviour of certain *Erwinia* species with respect to antibiotics [In Spanish, English summary]. *Microbiol. Esp.* 21:175-192.
- A41 Janke, C. 1974. The effect of intensified methods in potato growing on tuber susceptibility to the pathogen of tuber soft rot *Pectobacterium carotovorum* var. *atrosepticum* (Van Hall) Dowson after several months of storage [In German, English summary]. *Arch. Pflanzenschutz* 10:317-325.
- A42 Janke, C., and A. Heide. 1974. The effect of sprinkler irrigation and nitrogen fertilization on predisposition of potato tubers to the pathogen of tuber soft rot *Pectobacterium carotovorum* var. *atrosepticum* (Van Hall) Dowson one to eight weeks after harvest [In German, English summary]. *Archiv Pflanzenschutz* 10:263-274.
- A43 Jellis, J.G. 1975. Screening potato clones for resistance to dry rot (*Fusarium solani* var. *coeruleum*). *Ann. Appl. Biol.* 81:417-418.
- A44 Jones, E.D., and J.M. Mullen. 1973. The effect of potato virus X on susceptibility of potato tubers to *Fusarium roseum avenaceum*. *Amer. Potato J.* 50:384. (Abstr.)
- A45 Jones, E.D., and J.M. Mullen. 1974. The effect of potato virus X on susceptibility of potato tubers to *Fusarium roseum avenaceum*. *Amer. Potato J.* 51:209-215.
- A46 Jones, S.M., and A.M. Paton. 1973. L-phase of *Erwinia carotovora* var. *atrosepticum* and its possible association with plant tissue. *J. Appl. Bacteriol.* 36:279. (Abstr.)
- A47 Jones, S.M., and A.M. Paton. 1973. The L-phase of *Erwinia carotovora* var. *atrosepticum* and its possible association with plant tissue. *J. Appl. Bacteriol.* 36:729-737.
- A48 Kapustin, M.N. 1968. Biological features of the causal agents of wet rots [In Russian]. *Nauchn. Issled. Inst. Kartof. Khva.* 5:196-199.
- A49 Kelman, A., and S.H. DeBoer. 1974. Improved methods for detection of *Erwinia atroseptica* in potato tubers. *Proc. Amer. Phytopathol. Soc.* 1:124. (Abstr.)
- A50 Kelman, A., S.H. DeBoer, and D. Cuppels. 1974. Populations of pectolytic *Erwinia* spp. in the rhizosphere of potato plants. *Proc. Amer. Phytopath. Soc.* 1:90. (Abstr.)
- A51 Langerfeld, E. 1973. Effect of soil nutrient supply on the susceptibility of potato tubers to dry rot caused by *Fusarium coeruleum* (Lib.) Sacc. [In German]. *Potato Res.* 16:290-292.
- A52 Langerfeld, E. 1973. The effect of temperature on infection of potato tubers by fungi of the genus *Fusarium* Lk. [In German, English summary]. *Potato Res.* 16:224-233.
- A53 Langerfeld, E. 1973. Storage decay of potatoes [In German]. *Kartoffelbau* 24:216.
- A54 Leach, J.G. 1931. Blackleg disease of potatoes in Minnesota. *Univ. Minn. Agr. Exp. Sta. Tech. Bull.* 76:36 p.
- A55 Leach, J.G. 1931. Further studies on the seed-corn maggot and bacteria with special reference to potato blackleg. *Phytopathology* 21:387-406.
- A56 Leach, J.G. 1933. The method of survival of bacteria in the puparia of the seed-corn maggot (*Hyalemyia cilicrura* Rond.). *Zeit. Angew. Entomol.* 20:150-161.
- A57 Leach, S.S., and L.W. Nielsen. 1974. Reducing fusarial contamination of seed potatoes. *Amer. Potato J.* 51:305. (Abstr.)
- A58 Leach, S.S., and L.W. Nielsen. 1975. Elimination of fusarial contamination of seed potatoes. *Amer. Potato J.* 52:211-218.
- A59 Leach, S.S., and R.E. Webb. 1975. Screening for resistance to *Fusarium* tuber rot. *Amer. Potato J.* 52:246-247. (Abstr.)
- A60 Logan, C. 1966. Simple method of differentiating *Erwinia carotovora* var. *atroseptica* from *E. carotovora* and *E. carotovora* var. *aroideae*. *Nature* 212:1584.
- A61 Lund, B.M., and G.M. Wyatt. 1973. Nature of reducing compounds formed from sucrose by *Erwinia carotovora* var. *atroseptica*. *J. Gen. Microbiol.* 78:331-336.
- A62 Lyon, G.D., and C.E. Bayliss. 1975. Effect of rishitin on *Erwinia carotovora* var. *atroseptica* and other bacteria. *Physiol. Plant. Pathol.* 6:177-186.

Addendum (contd.)

- A63 Lyon, G.D., B.M. Lund, C.E. Bayliss, and G.M. Wyatt. 1975. Resistance of potato tubers to *Erwinia carotovora* and formation of rishitin and phytuberin in infected tissue. *Physiol. Plant Pathol.* 6:43-50.
- A64 McKeen, C.D., and K. Slingsby. 1972. Evaluation of chemical and cultural treatments to control blackleg in potatoes. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 275-277.
- A65 McKeen, C.D., and K. Slingsby. 1973. Comparative evaluation of seed piece diseases of four varieties of early potatoes. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 314-315.
- A66 McKeen, C.D., and K. Slingsby. 1974. Evaluation of seed piece decay on three varieties of early potatoes. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 303-304.
- A67 Mao, J.C., and J.E. Huguélet. 1971. Effect of temperature and relative humidity on the enzyme activity of *Fusarium roseum sambucinum* and concomitant symptom development in potato tubers. *Amer. Potato J.* 48:307. (Abstr.)
- A68 Mesterhazy, A., H. Henniger, and W. Bartel. 1973. Studies on pathological physiology of tuber rot on potatoes. Relations between rotting process and carbon dioxide production [In German, English summary]. *Arch. Pflanzenschutz* 9:245-249.
- A69 Molina, J., M.D. Harrison, and J.W. Brewer. 1974. Transmission of *Erwinia carotovora* var. *atroseptica* by *Drosophila melanogaster*. I. Acquisition and transmission of the bacteria. *Amer. Potato J.* 51:245-250.
- A70 Moreau, C. 1973. A new technique using formaldehyde gas to control fungi harmful to the storage of seed potatoes [In French]. *Pomme Terre Fr.* No. 358:13-15.
- A71 Motte, G., U. Burth, G. Brazda, R. Kloss, and S. Luck. 1974. Bercemaantispor 6459, a preparation for the treatment of seed potatoes [In German]. *Nachrichtenbl. (Dtsch. Pflanzenschutzdienst* 28:85-86.
- A72 Mullen, J.M., and D.F. Bateman. 1975. Enzymatic degradation of potato cell walls in potato virus X-free and potato virus - X infected potato tubers by *Fusarium roseum avenacearum*. *Phytopathology* 65:797-802.
- A73 Naumann, K., and W. Ficke. 1972. *Salmonella* - *Shigella* agar, a simple selective medium in comparison with other special substrates for the isolation of *Erwinia atrosepica* [In German, English summary]. *Zentralbl. Bakteriell. Parasitenk. Infektionskr. Hyg.* 11. 127:180-189.
- A74 Naumann, K., W. Ficke, H.J. Müller, K. Skadow, and R. Zielke. 1974. Transmission of potato blackleg and tuber soft rot [*Pectobacterium carotovorum* var. *atrosepticum* (Van Hall) Dowson] by soil seed material, and soil cultivation [In German, English summary]. *Arch. Pflanzenschutz* 10:301-316.
- A75 Nelson, G.A., and W.E. Torfason. 1971. Effect of inoculation and chemical treatment of potato seed pieces on potato emergence and yield. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 296-297.
- A76 Nelson, G.A., W.E. Torfason, and H.T. Allen. 1973. Control of decay of fresh-cut and pre-cut potato seed pieces by chemical treatment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 315-317.
- A77 Nielsen, L.W. 1963. Blackleg of Irish potato plants following European corn borer damage. *Plant Dis. Rep.* 47:272-275.
- A78 Nielsen, L.W. 1964. Pathogenesis of three *Erwinia* species to potato tuber tissue in a CO₂-N₂ atmosphere. *Phytopathology* 54:902. (Abstr.)
- A79 Nielsen, L.W. 1974. Isolation of *Erwinia* species from infected lenticels of potato tubers in saturated soils and immersed in water at different temperatures. *Amer. Potato J.* 51:307. (Abstr.)
- A80 Nielsen, L.W., F.L. Haynes, Jr., and J.T. Johnson. 1971. Heat preconditioning of tubers and fungicidal dusts for controlling *Fusarium* decay of seedpieces cut from seedstocks with high and low levels of contamination. *Amer. Potato J.* 48:307-308.
- A81 Ormrod, D.J. 1967. DMSO as a seed treatment additive for control of potato blackleg. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 191-192.
- A82 Ormrod, D.J. 1968. Seed treatments for control of potato blackleg. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 226-227.
- A83 Patzold, C., and H. Gehre. 1972. Effects of artificial infection of potato seeds with *Fusarium coeruleum* and influence of disinfection measures on development and yield of potato varieties [In German, English summary]. *Landbauforsch. Volkenrode* 22:129-132.
- A84 Pavék, J.J. 1975. Screening for resistance to *Fusarium* storage rot. *Amer. Potato J.* 52:247-248. (Abstr.)
- A85 Pérombelon, M.C.M. 1971. A quantal method for determining numbers of *Erwinia carotovora* var. *carotovora* and *E. carotovora atrosepica* in soils and plant material. *J. Appl. Bacteriol.* 34:793-798.
- A86 Pérombelon, M.C.M. 1973. Sites of contamination and numbers of *Erwinia carotovora* present in stored seed potato stocks in Scotland. *Ann. Appl. Biol.* 74:59-65.
- A87 Pérombelon, M.C.M. 1974. The role of the seed tuber in the contamination by *Erwinia carotovora* of potato crops in Scotland. *Potato Res.* 17:187-199.
- A88 Pérombelon, M.C.M., and R. Lowe. 1975. Studies on the initiation of bacterial soft rot in potato tubers. *Potato Res.* 18:64-82.
- A89 Philip, A., and S.C.Y. Liu. 1973. Immunodiffusional and immunoelectrophoretic comparison of *Erwinia carotovora* and *E. atrosepica*. *Phytopathology* 63:1110-1111. (Abstr.)
- A90 Radke, W., and A. Escande. 1975. Comparative studies of different methods of inoculating potato seedlings with *Fusarium solani* (Mart.) Sacc. f. sp. *eumartii* (Carp.) Snyder et Hansen. *Potato Res.* 18:243-255.
- A91 Richard, J.N. 1971. Potato seed treatment. *Pestic. Res. Rep., Res. Br., Agr. Can.* p. 305.
- A92 Samotus, B. 1971. Storage of potato tubers under water. Preliminary investigation. *Potato Res.* 14:145-149.
- A93 Slingsby, K., and C.D. McKeen. 1967. Evaluation of four seed treatments to control seed decay of potatoes. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 192-193.
- A94 Slingsby, K., and C.D. McKeen. 1973. Evaluation of potato seed piece decay control by chemical treatment and controlled environment. *Pestic. Res. Rep., Res. Br., Agr. Can.* pp. 228-229.
- A95 Sparks, W.C. 1973. Influence of ventilation and humidity during storage on weight and quality changes of Russet Burbank potatoes. *Potato Res.* 16:213-223.
- A96 Stachewicz, H. 1971. Investigations of *Fusarium* dry rot on potato tubers [In German English summary]. *Nachrichtenbl. Dtsch. Pflanzenschutzdienst.* 25:113-117.
- A97 Stanghellini, M.E., and J.C. Meneley. 1975. Identification of soft rot *Erwinia* associated with blackleg of potato in Arizona. *Phytopathology* 65:86-87.
- A98 Sturdy, M.L., and A.L.J. Cole. 1975. Cell wall degrading alpha-1, 3-arabinofuranosidase produced by potato dry rot pathogen, *Fusarium coeruleum* (Lib.) Sacc. *Ann. Botany* 39:331-335.
- A99 Tripathi, R.K., and M.N. Verma. 1975. Phenolic compounds and polyphenol oxidase activity in relation to resistance in potatoes against bacterial soft rot. *Ind. J. Exp. Biol.* 13:414-416.
- A100 Tsilosani, G.A., B.N. Khurtsiya, E.L. Dzhigauri, and N. Sh. Giorgobiani. 1973. Control measure against blackleg of potato in [Georgian, English summary]. *Tr. Nauchn. Inst. Zashch. Rust. Gruz. S.S.R.* 24:307-311.
- A101 Vrugink, H. 1975. Serological recognition of *Erwinia carotovora* var. *atrosepica* [In Dutch]. *Acta. Bot. Neerl.* 24:250.
- A102 Webb, L.E., and R.K. Wood. 1974. Infection of potato tubers with soft rot bacteria. *Ann. Appl. Biol.* 76:91-98.

Addendum (contd.)

- A103 Weingartner, D.P. and J.R. Shumaker. 1974. Results of treating freshly cut seed potatoes with fungicides and antibiotics. *Rhizoctonia solani*, *Fusarium*, *Sclerotium rolfsii*. II. *Erwinia* Proc. Fla. Hort.Soc. 87:201-205.
- A104 Wells, J.M. 1974. Growth of *Erwinia carotovora*, *E. atroseptica*, and *Pseudomonas fluorescens* in low oxygen and high carbon dioxide atmospheres. *Phytopathology* 64:1012-1015.
- A105 Yagudin, M.V. 1973. Application of zineb for pre-planting treatment of potato seed tubers [In Russian]. Tr. Khar'k. S-Kh. Inst. 172:120-121.
- A106 Zacharius, R.M., E.B. Kalan, S.F. Osman, and S.F. Herb. 1975. Solanidine in potato (*Solanum tuberosum*) tuber tissue disrupted by *Erwinia atroseptica* and by *Phytophthora infestans*. *Physiol. Plant Pathol.* 6:301-305.
- A107 Zielke, R., W. Ficke, K. Bagan, F. Linke, H.J. Müller, K. Naumann, and K. Skadow. 1975. Transmission of the potato blackleg and tuber soft rot pathogen, *Pectobacterium carotovorum* var. *atrosepticum* (Van Hall) Dowson by harvesting and screening machinery [In German, English summary]. *Arch. Pflanzenschutz* 11:31-41.
- A108 Zielke, R., H.J. Müller, W. Ficke, K. Naumann, and K. Skadow. 1974. The effect of soil and climate on the occurrence of blackleg and tuber soft rot of potato [In German, English summary]. *Arch. Pflanzenschutz* 10:245-253.
- A109 Zielke, R., H.J. Müller, W. Ficke, K. Naumann, and K. Skadow. 1974. Relation between blackleg infestation in potato stands and tuber soft rot in harvested crops [In German, English summary]. *Arch. Pflanzenschutz* 10:255-262.

Author index

Abdel' - Rékchim, M.A. 15, A1
 Adams, M.J. A2, A3
 Addy, S.K. 114, 172
 Agarwal, G.P. 471, 484
 Ali, S.A. A4
 Allen, H.T. 358, A76
 Allison, C.C. 325
 Altman, J. 16, 362
 Alvarado, E.L.F. 149, 526
 Amani. 150
 Anon. 14, 17, 442
 Arjunarao, V. 339
 Ark, P.A. 151
 Ayers, G.W. 67, 78, 263, 264, 387, 388, 389, 390, 391, 457a, 494, 495, A5, A6

Babaev, S.A. 107
 Bagan, K. A107
 Bahal, V.K. A11
 Bailey, D.L. 280
 Bailey, H.L. 246
 Bald, J.G. 265
 Barger, W.R. 163
 Baribeau, B. 363
 Bartel, W. 39, A68
 Baruah, P. 487
 Basham, H.G. 231
 Bateman, D.F. 231, A72
 Bates, G.R. 19
 Bayliss, C.E. A7, A62, A63
 Bazan de Segura, C. 266, 266a
 Beaumont, A. 268

Becker, E. A19
 Benlloch, M. 384
 Bennett, F.T. 152
 Beczner, J. A7
 Benza, J.C. 364
 Beraha, L. 215
 Berlinski, K. A8
 Bernaux, P. 334
 Bltencourt, A. 20
 Bhagwat, V.Y. A9, A10
 Bhargava, K.S. 269
 Bhattacharyya, S.K. A11
 Biehn, W.L. A12
 Binilauskait!, I. 86
 Black, W. 391a
 Blair, I.D. 21
 Blodgett, E.C. 270, 392, 528
 Blodgett, F.M. 199
 Blotskaya, Zh. 22
 Bobes, I. 23
 Bochow, H. 108
 Boerema, H. 24
 Bommer, D. A14
 Bonde, R. 25, 26, 81, 82, 83, 84, 100, 153, 154, 340, 393, 394, 395, 448
 Borg, A. 247
 Borchert, R. A15
 Börger, H. 434
 Bortels, H. 155, 248
 Bourke, P.M. 27
 Boyd, A.E.W. 112, 395a, 396, 397, 408, 409, 410, 430, 431, 491, 493, 496, 525, 526, A16
 Brandenburger, W. 271
 Brady, R.J. A17
 Brazda, G. 142, A19, A71
 Brenner, D.J. A18
 Brewer, J.W. A69
 Brewer, P.J. 272
 Brichet, J. 398
 Brook, M. 459
 Bucur, E. 98, 115
 Burkholder, W.H. 204
 Burth, U. A19, A71
 Busch, L.V. 273
 Bustamante, R.E. 400
 Butsevich, L.A. 123

Cadena-Hinojosa, M. 30
 Calderoni, A.V. 167
 Callbeck, L.C. 249
 Campbell, J.E. 67, 78
 Chamber, S.C. A20
 Chamberlain, E.E. 275
 Chan, E.C.S. A17
 Chesters, C.G.C. 459
 Chona, B.L. 401
 Choudhuri, H.C. 460
 Ciampi, L.R. 158
 Ciampi, P. 159, A21
 Ciferri, R. 365
 Clayton, E.E. 28
 Cochrane, J.C. 513
 Cochrane, V.W. 513
 Cole, A.L.J. A98
 Coleno, A. A23
 Colhoun, J. A22
 Conroy, R.J. 213
 Coombes, C. 440
 Corkle, M.A. 310, 432
 Correll, D.S. 276
 Costa, A.S. 277
 Cromarty, R.W. A24

- Crossan, D.F. 206
Cunningham, H.H. 529
Cunningham, H.S. 341, 385, 461
- Dainello, F.J. 160
Darling, H. 304
Darozhkin, M.A. A25
Das, C.R. 278
Davidson, R.S. 161, 234
Davidsson, I. 29
Davies, H.T. 146
Davis, J.R. A26
DeBoer, S.H. A27, A49, A50
de Souza, P. 84
De Wert, E. 24
Delaney, D. 402
Delgado-Sanchez, S. 30
DeLong, G.E. 260
Dembskaya, L. 31
Dennis, R.W.G. 162
Detilleux, E. 403
Deveza, M.C. 32
Dewey, D.H. 163
Dickey, R. 218
Dillon Weston, W.A.R. 404
Dippenaar, B.J. 279
Dobias, K. 238, A28, A29
Dobretsov, A.N. 33
Dorenbosch, M.M.J. 24
Downie, W.A. 414
Dowson, W.J. 79, 164, 192
Dubey, H.D. 507
Duncan, D.T. 216
Duncan, H.E. 85, 342
Dykstra, T.P. 165
Dzhigauri, E.L. A100
- Easton, G.D. 280, A24
Eddins, A.H. 166, 508
Edgar, A.D. 422
Edgerton, C.W. 281
Edmundson, W.C. 259
Effmert, M. 522
Eide, C.J. 46, 148, 349, 530
Elarosi, H. 518
El-Goorani, M.A. A1, A30
Ellis, N.K. 446
El-Kazzaz, M.K. A30
Elpidina, O.K. 282
Emilsson, B. 250
Emmond, G.S. 370
Epps, W.M. 34
Erinle, I.D. A31
Escandi, A. A90
Fanning, G.R. A18
- Feddersen, H.D. 283
Fehmi, S. 405
Felton, M.W. 284
Fernandez, P. A40
Fernandez Valiela, M.V. 167
Fernando, M. 168
Fernow, K.M. 251
Feuerbach, P. 285
Ficke, W. 35, A32, A73, A74, A107, A108, A109
Fink, H.C. 91
Fischer, R. 285a
Fisher, K.D. 343
Florea, N. 23
- Foiles, L.L. 346
Foister, C.E. 407, 408, 409, 410, 462, 463, 493, 499
Folsom, D. 288, 290, 374
Fox, L. 346
Fox, R.T.V. 128, 144, 237
Fredricks, A.L. 36
Freeman, J.P. A4
French, E.R. 40, 47, 329, 383
Friedman, B.A. 289, 290
Fucikovsky, L. 30
- Gallegly, M.E. 85, 342
Galloway, L.D. 291
Garci's, O.C. 251
Geesteranus, H.P.M. A34, A34
Gehre, H. A83
Gehring, F. 13
Gerasimova, T.P. 130, 208
Giorgobiani, N.S. A100
Glöckner, G. 292
Gonzalez, M.S. A21
Gorlenko, M.V. 210
Gorodetskii, V.S. 411, 485
Goss, H.W. 515
Goss, R.W. 240, 262, 412, 514
Goto, M. 235
Gradinarov, L. 382
Graham, D.C. 37, 79, 93, 94, 111, 113, 127, 135, 169, 293, 464, A36
- Granovsky, A.A. 294
Gratz, L.O. 295
Gray, E.G. 162
Gregg, M. 170
Gregor, J.W. 147
Griffith, R.L. 297
Guillemat, J. 465
Guimaraes, F.F. 296
Gustafsson, N. 250
Guthrie, J.W. 344
Guzman, N.J. 149
Guzman, V.L. A37
- Hall, J.A. A38
Hamilton, G.A. 293
Hankin, L. 214, A12, A13
Hansen, F. 171
Hansen, H.N. 330, 453
Hardie, J.L. 93
Haritonova, Z.M. 413
Harper, P.C. 94, 111, 112
Harrison, D.E. 414
Harrison, M.D. A36, A69
Hawkes, J.G. 414a
Hawn, W. 212
Haynes, F.L. Jr. A80
Hayward, A.C. 193
Heide, A. 108, 109, A42
Hellinga, J.J.A. 415
Hellmers, E. 38
Henniger, H. 39, 61, 140, A68
Herb, S.F. A106
Herold, M. 113
Hey, A. 239
Hidalgo, O.A. 40
Hide, G.A. 297
Hildebrandt, A.C. 106
Hingorani, M.K. 114, 172
Hirst, J.M. 297
Hollis, J.P. 240
Hollomon, D.W. 375

- Hooker, W.J. 216, 416
Hopkins, J.C.F. 417
Hoyman, W.G. 345
Hughes, I.K. 12
Huguelet, J.E. 41, A39, A67
Hurst, R.R. 99
Hyland, F. 340
- Il'icheva, A.A. 208
Iriarte, M.T. A40
Isenberg, F.M.R. 218
- Jaffe, M.J. 217, 218
Jamalainen, E.A. 301, 418
Janke, C. A41, A42
Jellis, S.G. A43
Jensen, J.H. 262, 515
Jetne, M. 42
Johnson, J.T. 319, 320, 367, A80
Jones, E.D. A44, A45
Jones, S.N. A46, A47
Joshi, M.M. 11
- Kalan, E.B. A106
Kasputin, M.N. 96, A48
Kaufman, J. 374
Keenan, P. 402
Kelman, A. A27, A49, A50
Kendrick, J.B. 173
Kerr, A. 236
Khairy, E.A. 518
Kharchenko, S.M. 174
Khurtsiya, B.N. A100
Khilkova, O. 104
Kiel, W. 126
Kirulis, A. 252
Kishore, H. 269
Klapp, E. 43
Klarnar, S. 105
Klemm, M. 44
Kloss, R. A19, A71
Knorr, L.C. 500
Knutson, K. 148
Koblet, R. 302
Kochetova, Z.M. 386
Korableva, N.P. 372
Koula, V. 226
Kovacikova, E. 97
Krantz, F.A. 515
Kranz, J. 479, 480
Krasil'nikov, N.A. 366
Kraus, J.E. 303, 419
Krug, H.P. 277
Kunkel, R. 259
Kuz'mina, G.N. 377
Kuznetso, V. 339
- Landis, B.J. 346
Langerfeld, E. 486, A51, A52, A53
Lansade, M. 501
Lapwood, D.H. 229
Lazar, I. 98, 115
Leach, J.G. 175, 176, 304, A54, A55, A56
Leach, S.S. 347, 348, A57, A58, A59
Leben, C. 325
Ledingham, R.J. 370
Lefebvre, C.L. 253
Lehmann, H. 254, 305
Lelièvre, D. 465
- Lepik, E. 45
Lhoste, L. 420
Libby, W.C. 288
Liu, S.C. 4, A89
Limasset, P. 421
Line, R.F. 46, 148, 349
Linke, F. A107
Lipsits, D.V. 64, 132
Livingston, J.E. 514, 516
Logan, C. 116, 131, 133, A60
Lovrekovich, H. 233
Lovrekovich, L. 233
Lowe, R. A88
Luck, S. A71
Lund, B.M. 117, 136, 177, A7, A61, A63
Lunden, A.P. 306
Lutman, B.F. 350
Lutz, J.M. 307, 351, 422
Lutz, L. 423
L'vova, N.M. 221
Lynch, P.B. 222, 223
Lyon, G.D. 178, A62, A63
- Maas - Geesteranus, H.P. 47, 129, 179
Macek, J. 255
Malcolmson, J.F. 49
Malyugin, P.A. 118
Mammen. 309
Manners, J.G. 128, 237
Mao, J.C. A67
Marshall, M. 56
Martinovic, M. 466
Masurat, G. 44, 50, 51, 52, 223
Mateev, A. 180
Matingley, G.H. 424
Matuo, T. 517
Maurer, A.R. 502
McChesney, J.D. A15
McIntosh, T.P. 425
McKee, R.K. 426, 427, 428, 429, 430, 431
McKeen, C.D. 352, 361, A64, A65, A66, A93, A94
Melhus, I.E. 310, 432
Meneley, J.C. A97
Mesterhazy, A. A68
Metcalf, H.M. 36
Metlitskii, L.V. 372
Michail, S.H. 518
Mikhal'chuk, V.T. A25
Miklos, G.V. A18
Mikula, J. 531
Milheiro, A.V. 53
Miller, P.R. 311
Millington, J.R. A20
Mills, W.R. 241
Miska, J.P. 1
Mitra, A. 519
Mol, J. 312
Molina, J. A69
Molnar, S. 358
Montaldo, A. 1A
Montgut, J. 465
Mool, J.C. 433
Moore, F.J. 503
Moore, W.C. 353
Moran, F. 211
Moreau, C. A70
Morgenweck, G. 43
Morozova, N.P. 372
Morwood, R.B. 313
Motte, G. A19, A71
Mount, M.S. 231

- Mujica, R.F. 181
Mukhin, E.N. 372
Mulder, D. 314
Mullen, J.M. A44, A45, A72
Müller, H.J. A33, A74, A107, A108, A109
Müller, K.O. 434
Munro, J. 146
Murdoch, A.W. 520
Murzakova, K.F. 134
Musat, D. 23
Myers, A. 128, 237
- Nadvodnyuk, Yu. N. 354, 355
Nagle, M.E. 280
Nance, N. 311
Napper, M.E. 315
Natrass, R.M. 316, 476
Naumann, K. A32, A33, A73, A74, A107, A108, A109
Nelsen, D.C. 41
Nelson, G.A. 356, 357, 358, A75, A76
Neumann, H. 285a
Newton, W. 371
Nicholls, J.C. 117
Nickel, J.L. 137
Nielsen, L.W. 9, 40, 74, 119, 243, 318, 319, 329, 367, 383, A57, A58, A77, A78, A79, A80
Noble, M. 56
Noll, A. 227
Novakova, J. 139
- Okabe, N. 235
Olgay, M. 219
Ollila, L. 438
Oloffsson, B. 244
Onsager, J.A. 346
Oort, A.J.P. 321
Ormel, H.A. 312
Ormrod, D.J. A81, A82
Osman, S.F. A106
- Padwick, G.W. 323
Page, O.T. 510
Pal, A. 278
Pall, O. 23
Palm, E.T. 324
Paton, A.M. A46, A47
Patzold, C. A14, A83
Paulus, A.O. 173
Pavek, J.J. A84
Pawuk, W.H. 521
Peacock, W.M. 189
Pelczar, M.J. Jr. A17
Peralta, G.J. 257
P!rombelon, M.C.M. 8, 10, 59, 145, 230, A85, A86, A87, A88
Perseca, E. 23
Peschel, R. 50, 183, 223
Peters, E.J. 509
Pethybridge, G.H. 439
Petroczi, I. 373
Pett, B. 39, 60, 61, 522, A19
Pett, E. 113
Philip, A. A89
Phillips, D.H. 62
Phillips, D.V. 325
Pitt, D. 440
Potter, H.S. 416
Prunier, J.P. 20
- Qureshi, A.A. 510
- Radtke, W. 443, A90
Ragozina, I.I. 64, 132
Raleigh, W.P. 100, 448
Ramsey, G.B. 197, 215
Rapilly, F. A23
Read, D.C. 78
Reid, W.J. 189
Reinking, O.A. 341, 461
Rich, A. 270
Richard, J.N. 390, 457a, A6, A91
Riker, A.J. 106
Roach, H.Q. 374
Robbs, C.F. 66, 190
Robinson, D.B. 67, 78, 99, 391, 495
Roland, G. 333
Roll-Hansen, F. 444
Rose, D.H. 191
Rowberry, R.G. 273
Rubin, B.A. 372
Rudd Jones, D. 164, 192
Ruehle, G.D. 68, 166, 445
Ruschmann, G. 69
Russell, J.D. 332
- Sabet, K.A. 70
Sal'kova, E.G. 372
Salzmann, R. 71
Samotus, B. A92
Sampson, P.J. 193
Samson, R.W. 446
Samuel, G.G. 467
Sands, D.C. A12
Sardina, J.R. 447
Schaal, L.A. 259
Schipppers, P.A. 468, 481
Schneider, R. 482
Schoene, K. 489
Scholz, M. 39
Schomer, H.A. 191
Schultz, E.S. 100, 448
Schultz, O.E. 359
Seminario, B. 329, 383
Servazzi, O. 469
Shepherd, D.R. 432
Sherf, A.F. 194
Shneider, Yu. I. 64, 104, 132, 134, 141, 208
Shumaker, J.R. A103
Shuvalova, S.Z. 101, 102
Sieczka, J.B. 360
Simon, F.G. 513
Simpson, G.W. 288
Singh, R.K. 451
Singh, R.S. 11
Skadow, K. A32, A74, A107, A108, A109
Skinner, F.A. 506
Slingsby, K. 352, 361, A64, A65, A66, A93, A94
Smale, B.C. 232
Small, T. 452, 470, 477, 505
Smarda, J. 195
Smart, H.F. 121
Smith, J.H. 376
Smith, M.A. 196, 197, 215
Smith, W.L. 72, 121, 204
Snyder, W.C. 330, 453, 517, 523
Spaith, J. 513
Spalding, D.H. 232
Sparks, W.C. 529, A95
Spenneman, F. 43
Stachewicz, H. 228, A19, A96
Stahmann, M.A. 233
Stanghellini, M.E. 332, A97
Staniland, L.N. 268

- Staples, R.R. 73
Stapp, C. 258
Starr, M.P. 211
Staruigina, L.P. 198
Stedman, O.J. 297
Steigerwolt, A.G. A18
Stephan, S. 44, 50, 51, 52, 124, 223
Stevenson, F.J. 260
Stevenson, G. 168
Stone, W.J.H. 6
Störmer, I. 261
Sturdy, M.L. A98
Szabo, A. 373
- Tandon, R.N. 471
Taylor, C.F. 199
Taylor, R.E. 404
Telneset, S.O. 245
Thurston, H.D. 400
Tickle, J.H. 525
Todd, J.M. 472
Tolaas, A.G. 515
Torfason, W.E. 356, 357, 358, A75, A76
Torres, H. 74
Townsend, G.R. 166
Tranina, N.F. 75
Tripathi, R.K. A99
Tsilosani, G.A. A100
Tucker, J. 103
- Upstone, M.E. 511, 524
Urosevic, B. 97
- Van Andrichem, M. 502
Van den Boom, T. 122
Van Kesteren, H.A. 24
Vanderwalle, R. 333
Verma, M.N. A99
Vicente, R. 125
Vielwerth, V. 209
Vinot, M. 334
- Vitukevich, E.R. 438
Volcani, Z. 106, 169
Voronkevich, I.V. 123
Votoupal, B. 457
Vruggink, H. A101
- Wade, E.K. 335
Wade, G.C. 76
Wager, R.M. 336
Waggoner, P.E. 220
Wahlin, B. 200
Wallace, M.M. 454
Watson, D. A15
Watson, R.D. 201
Webb, L.E. A102
Webb, R.E. A59
Wedding, R.T. 173
Weingartner, D.P. A103
Wells, J.M. A104
Werner, H.O. 515
White, N.H. 202
Wiant, J.S. 374
Wilkins, V.E. 473
Wilson, A.R. 408, 409, 410, 462, 463, 474, 475, 493
Wood, F.A. 521
Wood, R.K.S. 520, A38, A102
Woodbury, G.W. 419
Wright, R.C. 189, 422
Wright, W.R. 215
Wyatt, G.M. 136, A61, A63
Wyman, O.L. 288
Yagudin, M.V. A105
Young, D.A. 146
Young, L.C. 146
- Young, R.A. 324
- Zacharius, R.M. A106
Zaehring, M.V. 529
Zielke, R. A33, A74, A107, A108, A109
Zimmermann-Griess, S. 338
Zucker, M. 214

Resistance of turfgrasses to low-temperature – basidiomycete snow mold and recovery from damage¹

J. Drew Smith

Most strains of *Poa pratensis*, *Festuca rubra*, and *Festuca ovina* were heavily damaged in tests at Saskatoon when turf plots were inoculated with cultures of the nonsclerotial low-temperature basidiomycete, LTB, grown on sterile rye grain. Suspects included several of the cultivars favoured for use in other climatic regions where the range of snow molds and other turf pathogens is different from that on the prairies. No strains completely resistant to the LTB were found, but some new introductions, selections and established cultivars showed low initial damage and/or rapid recovery. Cultivars require regional testing for disease resistance, especially snow mold resistance, before being recommended for use. Varietal descriptions should specify resistance to a particular snow mold pathogen or pathogens not to "snow mold" since the spectrum of these varies greatly from region to region.

Can. Plant Dis. Surv. 55:147-154. 1975

L'inoculation d'un basidiomycète psychrophile sans sclérotés (LTB) cultivé sur des grains de seigle stériles à des parcelles de gazon a fortement endommagé la plupart des lignées de *Poa pratensis*, *Festuca rubra* et *Festuca ovina* utilisées dans des essais réalisés à Saskatoon. Plusieurs des cultivars sensibles sont ceux qu'on utilise de préférence dans les régions climatiques où les moisissures nivéales et les autres organismes pathogènes du gazon diffèrent de ceux des Prairies. Aucune lignée n'a manifesté une résistance complète au LTB, mais quelques nouvelles sélections et quelques cultivars établis ont montré peu de dégâts initiaux et (ou) se sont rétablis rapidement. Pour pouvoir être recommandé, les cultivars doivent subir des essais multilocaux de résistance à la moisissure nivéale. Les descriptions de variété devraient spécifier les types de pathogène de la moisissure nivéale auxquels les cultivars sont résistants étant donné que les pathotypes de ces dernières varient beaucoup d'une région à l'autre.

Snow mold caused by an unidentified, non-sclerotial low-temperature basidiomycete, LTB, is widespread on domestic lawns and other amenity turf in the lower snowfall regions of western Canada and may cause considerable damage in some years (4, 5). LTB snow mold may be controlled by applications of nonmercurial fungicides made before the development of a permanent snow cover. However, severe attacks on susceptible cultivars may require the use of mercurous/mercuric chloride mixtures (6).

Few studies have been made on the resistance to the LTB of cultivars of *Poa pratensis* L., *Festuca rubra* L. and *Festuca ovina* L. The first two species are the most common components of domestic lawns and other amenity turfs in the prairies. Cormack (1) found that *P. pratensis* had moderate to high resistance and that *F. rubra* and *F. ovina* had low resistance under the conditions of southern Alberta. In order to improve control of this disease and reduce the need to rely on fungicides, many introductions and local selections of *P. pratensis* and fine-leaved *Festuca* spp. were screened for resistance and recovery following artificial inoculation with cultures of the fungus grown on sterile grain. The results of field tests in 1974 are presented here.

Materials and methods

Plant materials

Tests 1, 2, and 3 included 99 accessions of *P. pratensis* cultivars and selections from many countries and the prairies of Canada. Test 1 comprised 65 accessions replicated six times in random fashion. In test 2, 17 lines were similarly replicated four times. In Test 3, 17 lines were replicated only twice. The numbers of replicates and the seeding rate were governed mainly by availability of seed. Seed was sown with a multiple belt seeder on 3 June 1971. Row spacing was 22.5 cm, row length 3.2 m, with four rows per plot. When established, the grasses were mown at 5 cm height, at first with a rotary mower mounted under a garden tractor and then with tractor-drawn reel mowers. Clippings were returned. Irrigation was supplied when needed. Plots were top-dressed in midsummer with a soil/sand/peat mixture to encourage turf formation. Although tests indicated adequate soil phosphate levels, 3 kg of 16-20-0 and 3 kg of 33-0-0 fertilizer per 100 m² were applied per annum.

Test 4 comprised 12 commercial cultivars and 12 selections of *Poa pratensis* in 1 m² plots replicated four times in a randomized block arrangement. Seeding rate depended on amount of seed available. Seed was sown by hand broadcasting in June 1972. Fertilization was similar to Tests 1, 2, and 3; mowing was done with a domestic rotary or small, self-propelled reel mower to 5 cm height. Clippings were returned.

¹ Contribution No. 592, Research Station, Agriculture Canada, 107 Science Crescent, Saskatoon, Saskatchewan S7N 0X2.

Test 5 comprised 6 cultivars and 10 selected lines of *F. rubra* (56- and 42-chromosome types) and 3 lines of *F. ovina* in 1 m² plots replicated four times in a randomized block arrangement. Seeding, fertilization and mowing were identical with that in Test 4.

Test 6. Where seed was in shorter supply than in Test 5, it was only possible to sow duplicate plots of a further 12 lines of *F. rubra* and four of *F. ovina*. Seeding, mowing and fertilizing were the same as in Tests 4 and 5.

Tests 7 and 8. Where seed from only a few plants was available or that available was of poor germination, it was sown in soil in 30 X 42 cm greenhouse flats. This sowing was done in late winter; when coherent turf was available in spring, it was removed from the flat and laid on a levelled soil bed. With suitable topdressing and judicious mowing, a well-grown turf became available for testing in the fall of the same year. Test 7 comprised 48 lines of *P. pratensis*. Test 8 comprised 37 lines of *F. rubra* and 7 of *F. ovina*. In both tests, each line was replicated three times.

Inoculation

The LTB isolate Ju714a used to establish an epidemic was derived from diseased patches of *Agrostis* sp. from the sixth green at the Golf and Country Club, Moose Jaw, Saskatchewan, in spring 1971. It had been proved highly pathogenic on *P. pratensis* and *F. rubra* in seasons previous to 1973 (6). It was grown on sterile, moist rye grain in 1.14 liter milk bottles at 6°C for 3 months, air-dried, crushed, and stored at -10°C until required. Seventeen kilograms of the dried inoculum was applied to 2125 m² (8 g/m²) of the test areas by hand broadcasting in several directions on 5 and 7 September 1973.

Rating for disease

This was done on 23 April and 6, 15, and 24 May 1974. The percentage area affected by the disease was recorded.

Results

Susceptible strains of *P. pratensis*, and *F. rubra*, and *F. ovina* were heavily damaged by the LTB isolate in all tests (Tables 1 to 8 and Figs. 1, 2, & 3). Many of the *F. rubra* and some of the *F. ovina* strains showed more than 80% damage (Tables 5, 6, & 8 and Fig. 1) on 23 and 29 April, 2 and 3 weeks respectively after the snow cover had gone. However *P. pratensis* strains (Tables 1, 2, 3, 4, & 7 and Fig. 1) generally did not show as much

initial damage and they recovered more rapidly from damage. This is apparent when percent damage for the *P. pratensis* strains on 15 May (Tables 1, 2, 3, & 4) is compared with that for *F. rubra* and *F. ovina* on the same date (Tables 5 & 6). Similar differences in rate of recovery are apparent when data for *P. pratensis*, *F. rubra* and *F. ovina* are compared in the microplot tests (Tables 7 & 8, and Fig. 3). In some tests, a strain was entered twice with seed from different lots; for example IH 2079, Primo (Table 1), Reptans (Table 5), Olds (Table 6), Dawson, Boreal, Goldfrod, Reptans, and Olds (Table 8). Differences between ratings of the same cultivar were not significant. None of the strains were completely resistant to the LTB, but the *P. pratensis* lines S-7763 (Table 1), S-8606 (Table 2), K35584 and K35605 (Table 7) showed high resistance and, except for K35605, rapid recovery from damage. *F. rubra* line S-1765 (Tables 6 & 8) and *F. ovina* lines 2069 (Table 5), 2065 (Table 6), S-1758, S-1733, S-1792, and S-3482 (Table 8) were outstanding in resistance. In some of the moderately resistant *P. pratensis* strains, eg. in those with initial ratings of less than 40% infection (Table 1), recovery was generally rapid.

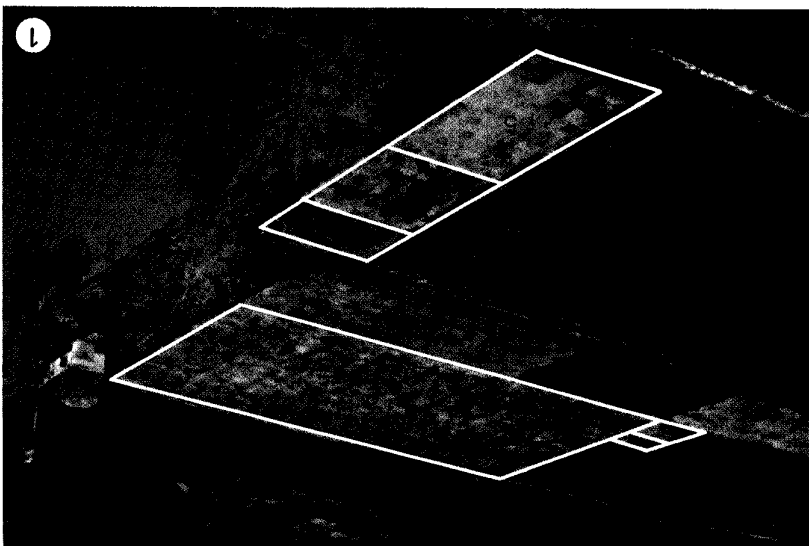
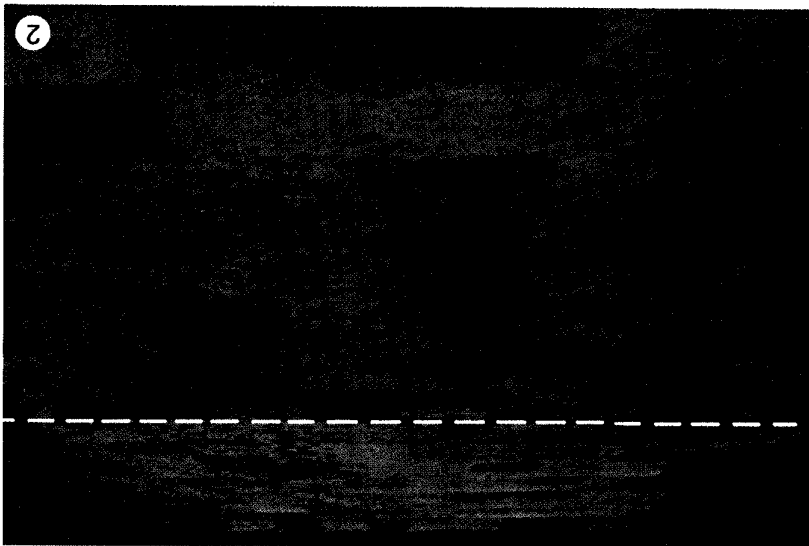
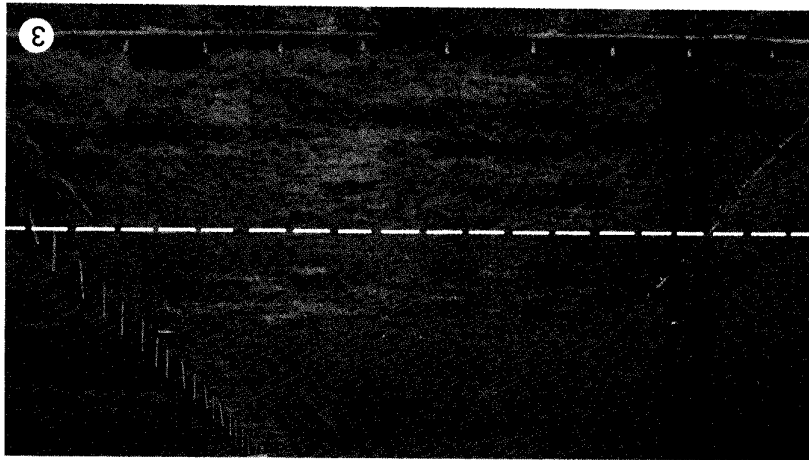
Discussion

The considerable differences in susceptibility to the LTB snow mold found among strains of *P. pratensis*, *F. rubra*, and *F. ovina* and in rate of recovery from damage suggest that field pathogenicity tests of the kind employed are suitable for the selection of disease resistant lines. The row method of seeding employed in Tests 1, 2, and 3 permitted additional data to be obtained on susceptibility of entries to powdery mildew [*Erysiphe graminis* DC. ex Méral] and rust [*Puccinia poae-nemoralis*, Otth.]; in the central prairie region these diseases are much more prevalent on *Poa pratensis* in spaced rows than in mown turf. Row seeding also allows the formation of turf from smaller amounts of seed than conventional broadcast sowing. Additional data may also be obtained on rapidity of turf formation by selections. The knitting together of turf may be encouraged by top-dressing. However, mature turf suitable for inoculation and disease studies may require several years to develop. Both the row seeding and the microplot method (Tests 7 and 8) minimized risk of test failures due to "washout", common in this region, and eliminated the need to await calm conditions necessary for successful multiple plot seedings. The microplot technique provided a very effective rapid screening

Figure 1. Aerial view of low-temperature basidiomycete (LTB) test plots on 29 April 1974, about 3 weeks after snow melt. Some recovery is apparent in *Poa pratensis* lines in particular (Tests 1-4). Key to test plots: 1-4, 7 *Poa pratensis*; 5, 6, 8 *Festuca rubra* and *F. ovina*.

Figure 2. LTB test plots, 8 May 1974. *Poa pratensis* plots (Test 4) below line; *Festuca* spp. plots (Tests 5 and 6) above line. Considerable recovery has taken place in several *P. pratensis* lines, less frequently in fescues.

Figure 3. LTB test plots, 8 May 1974. Microplots of *Festuca* spp. (Test 7) below line; microplots of *Poa pratensis* (Test 8) above line. Considerable recovery has occurred in most *P. pratensis* lines and in some *Festuca* spp. lines.



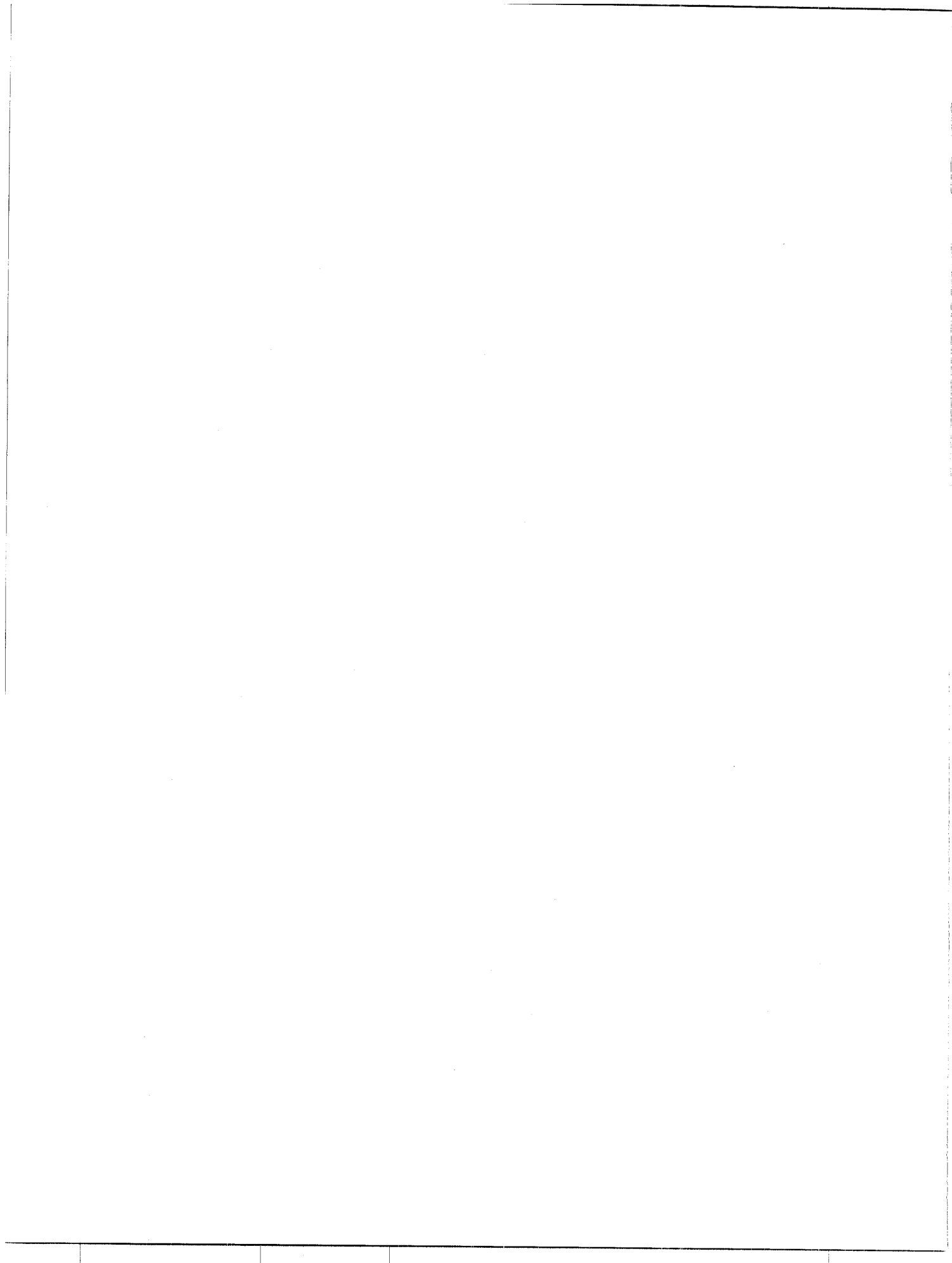


Table 1. Resistance and recovery of *Poa pratensis* lines from LTB snow mold, 1974: Test 1 (6 replicates)

Cultivar or line	Origin ¹	Percent area of turf affected			
		23 Apr	6 May	15 May	Avg
S-7763	USSR	16	7	1	8
S-7766	Canada	27	19	5	17
Park	USA	29	18	8	18
Norma Øtofte	Denmark	27	24	7	19
Mostovskij	USSR	38	18	3	19
Hunsballe	Denmark	30	25	4	20
Captan	Neth.	41	16	3	20
S-7759	Canada	38	21	2	20
S-7764	Canada	31	24	6	20
IH 2079	Canada	34	22	5	20
EFS64	Denmark	34	25	5	21
Kahnstein	Germany	33	24	8	22
Skandia II	Sweden	35	25	6	22
Delta	USA	32	25	10	22
K22011	USSR	43	23	6	23
S-7760	Canada	38	30	3	23
S-8598	Canada	30	26	19	25
S-7762	Canada	37	29	9	25
S-7767	Canada	40	29	8	26
Delft	Neth.	36	28	15	26
IV-89-61	Hungary	33	32	16	26
S-8595	Canada	33	27	19	26
S-7765	Canada	41	28	13	27
Line 59	Neth.	29	32	22	27
Adorno	Neth.	38	27	18	27
S-8596	Canada	46	30	8	28
IH 2079	Canada	36	34	7	28
K35603	Estonia	40	34	13	29
Troy	USA	51	28	9	29
S-8597	Canada	41	27	12	30
S-8599	Canada	34	32	13	30
Atlas	Sweden	39	33	18	30
Golf	Sweden	42	35	17	31
SK 46	Poland	41	30	13	31

Table 1 (continued)

Cultivar or line	Origin	Percent area of turf affected			
		23 Apr	6 May	15 May	Avg
Tygera	USSR	30	33	32	31
S-8600	Canada	40	37	18	32
Kentucky	USA	37	36	23	32
K 31808	Latvia	43	33	23	33
S-8611	Hungary	48	35	18	33
S-7758	Canada	48	33	21	34
Steinacher	Germany	49	41	13	34
RVP	Belgium	44	40	25	36
MLM 18006	Unknown	36	48	26	37
Arista	Neth.	45	43	23	37
Baron	Neth.	36	41	27	38
Windsor	USA	44	44	25	38
Primo	Sweden	41	40	33	38
S-8601	Canada	48	48	18	38
Monarch	England	42	42	32	39
S-7761	Canada	41	48	28	39
Nike Daehnfeldt	Denmark	49	47	22	39
Newport	USA	45	44	31	40
Sydsport	Sweden	43	49	35	42
Nugget	USA	48	43	38	43
S-8591	Canada	48	44	40	44
Primo	Sweden	50	52	33	45
S-8592	Canada	53	51	32	45
S-7756	Canada	54	51	40	48
Fylking	Sweden	55	47	46	49
S-5894	Canada	61	51	39	50
S-7768	USSR	65	58	32	51
MLM 18005	Unknown	63	48	41	51
S-7757	Canada	53	51	32	52
IV-89-24	Hungary	61	55	43	53
Barkenta	Neth.	65	64	56	62
LSD 1%		30	29	21	

¹Neth = The Netherlands

method for lines where seed supply was short. Turf can be produced in the winter for planting the following spring. The method probably suffers from deficiencies inherent in very small plot studies; there is an "edge effect" and great heterogeneity in resistance and susceptibility between adjacent components in a small area. This probably affects the progress of the infection. So far no satisfactory laboratory or greenhouse technique has been developed which can satisfactorily simulate the conditions of turf under a snow cover, which is necessary for this disease to develop. In culture the LTB isolate used produces hydrocyanic acid, which is probably important in pathogenesis.

Of the *P. pratensis* strains tested, S-7763, an introduction from the Murmansk region of the USSR, showed high resistance and recovery in the row-seeded (Table 1) and broadcast (Table 4) tests. This strain in these, and other tests, shows early winter dormancy and more rapid spring regrowth than any other strain so far tested

at Saskatoon. These are probably desirable characters for this region. It shows high resistance to *E. graminis* and moderate resistance to *P. poae-nemoralis* in seed rows (unpublished). It is an excellent seed producer and highly apomictic. Other lines showed considerable resistance and quick recovery from damage or, while apparently susceptible to attack, showed quick recovery from damage. For example, in the broadcast seeded plots an introduction from Italy, S-8606, rated significantly better in both characters than the cultivars Prato, Cougar, and Fylking (Table 2). In the microplot tests (Table 7), K35584 from the USSR was outstanding in resistance and recovery from damage, but six other lines from the USSR, Canada, and the USA, namely K35605, 19, C68-79, 2, K28704, and 145, had fair resistance, and, except for K35605, good recovery.

The poor resistance and slow recovery of many well known *P. pratensis* strains, some of which are commonly sold for turf formation in the prairies, are major findings

Table 2. Resistance and recovery of *Poa pratensis* lines from LTB snow mold, 1974: Test 2 (4 replicates)

Cultivar or line	Origin	Percent area of turf affected			
		23 Apr	6 May	15 May	Avg
S-8606	Italy	20	6	3	10
S-8604	Italy	26	13	5	15
S-8605	Italy	30	13	8	17
170	England	24	18	11	18
K28704	USSR	31	16	6	18
K27874	USSR	49	19	5	24
IV-89-59	Hungary	36	25	18	26
K27307	USSR	44	25	13	27
C68-79	USA	43	26	13	27
S-8602	USSR	46	33	19	33
Rogue	Canada	44	34	21	33
Dasas	Denmark	56	36	15	36
K35602	Estonia	54	45	30	43
K35604	Estonia	58	49	39	48
Prato	Neth.	65	45	41	50
Cougar	USA	64	53	48	55
Fylking	Sweden	64	49	53	55
LSD 1%		43	32	23	

Table 3. Resistance and recovery of *Poa pratensis* lines from LTB snow mold, 1974: Test 3 (2 replicates)

Cultivar or line	Origin	Percent area of turf affected			
		23 Apr	6 May	15 May	Avg
S-7831	Unknown	28	23	6	19
S-8603	England	50	23	11	28
S-8741	Unknown	40	23	23	28
S-7840	Unknown	33	23	38	31
S-7835	Unknown	35	28	35	33
S-7839	Unknown	55	33	20	36
S-7830	Unknown	70	35	6	37
Stensballe	Denmark	65	33	15	38
S-7837	Unknown	60	43	18	40
S-8609	Canada	76	35	10	40
Merion	USA	55	40	28	41
S-7834	Unknown	63	38	33	44
S-8610	Canada	65	50	16	44
S-7832	Unknown	73	50	33	52
S-7838	Unknown	68	45	43	52
S-7836	Unknown	63	58	40	53
S-7833	Unknown	83	65	30	59

of the tests. Many of these were developed for use in turf or pastures elsewhere. Barkenta, Fylking, Primo, Nugget, Sydsport, Windsor, and Baron (Table 1); Fylking and Cougar (Table 2); Merion (Table 3); and Cougar, Merion, Baron, Arista, Fylking, Barenta, Nugget, Primo, and Sydsport (Table 4) performed poorly. Some of these have received good ratings for resistance to certain diseases (2) that do not usually cause significant damage in the Saskatoon region or in adjacent areas with similar climatic conditions. On the other hand, the superior performance of Park (Tables 1 & 4), Captan (Table 1), and Delta (Table 4) is apparent.

All *F. rubra* strains tested (Tables 5, 6, and 8), except S-1765 (Tables 6 and 8), a 1947 introduction from Kazakhstan in the USSR, showed heavy damage following snow melt. The only cultivar which approached S-1765 in resistance was Arctared, but strains K34675, K25236, and K22609 (Table 5) showed a significantly faster rate of recovery than any other strains in Test 5, including all Canadian cultivars.

The *F. ovina* L. var. *saximontana* Rydb. entries 2069 (Table 5) and 2065 (Table 6), which were Saskatchewan selections from the Cypress Hills, S-1758, S-1733, and S-1792 from Kazakhstan in the USSR, and S-3482, a hard fescue, *F. ovina* var. *duriuscula* L., of unknown origin (Table 8) showed considerably greater resistance to LTB damage than the remainder. The cultivar Barenza and the strain 2107 (Tables 6 & 8) suffered severe damage which resulted in death of most of the turf on these plots.

Of the winter diseases, snow mold caused by the LTB is probably the most prevalent in the prairies on bluegrass/fescue turf commonly used in the formation of domestic lawns and other irrigated amenity turf. It is also the most difficult to control. Particularly in a cool, dry spring following heavy damage recovery is slow and turf may be severely thinned out. Observations of field cases suggest that the prolongation of vegetative growth into late fall by heavy or late summer nitrogen applications increases turf susceptibility to LTB, as with disease caused by *Fusarium nivale* (3). Winter dormancy is a mechanism related to the survival of the perennial grasses through an unfavorable period when turf is prone to attacks of psychrophilic fungal pathogens. The onset of this dormancy is signaled by a slowing of leaf production and by leaf death and attendant chlorophyll loss, which may be referred to as "browning off". *P. pratensis*, *F. ovina*, and *F. rubra*, cool season grasses, do not have a complete winter dormant period and are capable of growth during the winter in mild, bright periods, for example, in midwinter thaws in "chinook" regions. However, the earliness and completeness of the "browning off" process in early winter varies from strain to strain. Since it is thought that pathogenesis in the LTB is related to tissue damage from hydrocyanic acid production (7), it is possible that this production is dependent on the chemical composition or condition of the substrate (the turf grass plant), which may influence the amount of plant damage caused. Early and deep dormancy may also conserve soil nitrogen and husband stored plant food reserves which allows rapid "take-off"

Table 4. Resistance and recovery of *Poa pratensis* lines from LTB snow mold, 1974: Test 4 (4 replicates)

Cultivar or line	Origin	Percent area of turf affected			
		23 Apr	6 May	15 May	Avg
S-7763	USSR	28	6	0	11
IH 2079	Canada	29	26	4	20
S-7760	Canada	39	23	8	23
S-7759	Canada	39	24	13	25
Delta	USA	40	26	15	27
Park	USA	48	31	15	31
K35603	Estonia	61	25	9	32
S-8596	Canada	45	15	9	33
K35602	Estonia	60	45	28	44
Sydsport	Sweden	50	58	46	51
Primo	Sweden	55	56	43	51
Steinacher	Germany	73	55	35	54
Nugget	USA	61	53	51	55
Barkenta	Neth.	81	68	49	59
K35604	USSR	69	73	45	62
S-8591	Canada	68	62	58	62
Fylking	Sweden	65	66	58	63
Arista	Neth.	69	70	55	65
Baron	Neth.	68	74	66	69
Merion	USA	78	73	65	72
S-7756	Canada	75	71	70	73
Line 59	Neth.	84	78	70	77
S-7757	Canada	88	80	76	78
Cougar	USA	83	83	83	83
LSD 1%		50	44	44	

Table 5. Resistance and recovery of *Festuca rubra* and *Festuca ovina* lines from LTB snow mold, 1974: Test 5 (4 replicates)

Cultivar or line	Origin	Percent area of turf affected				
		23 Apr	6 May	15 May	24 May	Avg
<i>F. rubra</i>						
K34675	Latvia	90	38	9	13	27
K25236	USSR	80	30	10	4	31
K22609	USSR	83	53	19	9	41
K27390	Lithuania	79	68	39	35	55
K31197	USSR	90	79	48	40	64
Olds	Canada	88	81	58	63	69
S-7850	Canada	83	81	63	54	70
Reptans	Neth.	80	79	73	64	74
Reptans	Neth.	78	78	74	68	74
Durlawn	Canada	88	89	79	50	76
Dawson	England	94	86	76	53	77
Baron	Neth.	63	85	74	78	77
K35599	Estonia	95	89	70	61	79
Pennlawn	USA	88	91	83	76	84
Canada No. 1 (S-7374)	Canada	90	91	86	76	86
K31286	Latvia	99	94	91	63	87
MLM 15010	Unknown	91	93	92	92	92
<i>F. ovina</i>						
2069	Canada	4	33	3	8	12
1811-456	Armenia	45	40	29	25	35
562	Germany	97	92	86	68	86
LSD 1%		38	40	33	39	

Table 6. Resistance and recovery of *Festuca rubra* and *Festuca ovina* lines from LTB snow mold, 1974: Test 6 (2 replicates)

Cultivar or line	Origin	Percent area of turf affected				
		23 Apr	6 May	15 May	24 May	Avg
<i>F. rubra</i>						
S-1765	U.S.S.R.	15	15	15	15	15
Sceempter	Neth.	65	75	60	60	65
1081	Canada	70	85	55	60	68
1079	Canada	85	90	70	50	74
Duraturf	U.S.A.	100	93	70	55	79
Olds(1098)	Canada	100	90	80	50	80
Oasis	Neth.	100	97	78	55	82
1120	Canada	90	93	85	87	86
1118	France	90	93	88	75	86
1111	Canada	95	93	85	75	87
KL257	Unknown	95	95	85	80	89
Olds(1084)	Canada	100	95	88	75	89
<i>F. ovina</i>						
2065	Canada	13	10	8	10	10
2066	Canada	28	70	40	30	42
2107	Neth.	95	98	85	85	91
Barenza	Neth.	90	98	93	95	94
LSD 1%		55	35	49	51	

in growth and hence recovery from disease damage in spring when favorable climatic conditions occur.

Because of different climatic conditions and a different range of snow mold and other pathogens, including the LTB and a highly pathogenic *Typhula* sp. at present designated *Typhula* FW (6), *P. pratensis*, *F. rubra*, and *F. ovina* cultivars recommended for use elsewhere should not be employed for turf formation on the prairies on a large scale without adequate regional testing. Varietal descriptions may be faulty when the generic term "snow mold" is used to indicate disease resistance without specifying which pathogen is concerned.

Acknowledgments

I am indebted to W. W. Reiter, W. Leonard, and C. Tennant for valuable technical assistance.

Literature cited

1. Cormack, M. W. 1952. Winter crown rot or snow mold of alfalfa, clovers and grasses in Alberta. II. Field studies on host and varietal resistance and other factors related to control. Can. J. Bot. 30:537-548.
2. Ormrod, D. J. 1973. Diseases of lawns. British Columbia Dep. Agr., Victoria, B.C. 7 pp.

Table 7. Resistance and recovery of *Poa pratensis* lines from LTB snow mold 1974: Test 7 (Microplot, 3 replicates)

Cultivar or line	Origin	Percent area affected		
		24 Apr	6 May	Avg.
K35584	USSR	5	0	2
K35605	USSR	10	7	8
19	Canada	18	3	11
C68-79	U.S.A.	17	5	11
2	Canada	20	3	12
K28704	U.S.S.R.	18	7	12
14	Canada	18	10	14
21	Canada	23	10	16
K27307	U.S.S.R.	22	17	19
K27397	U.S.S.R.	30	10	20
18	Canada	25	15	20
145	Canada	37	7	22
74	England	27	20	23
S-8610	Canada	22	25	23
16	Canada	33	15	24
23	Canada	38	13	26
73	England	27	27	27
20	Canada	38	17	27
17	Canada	40	15	28
15	Canada	40	17	28
S-7844	Canada	42	18	30
K28748	U.S.S.R.	38	22	30
1	Canada	40	20	30
22	Canada	40	22	31
IV-89-59	Hungary	40	22	31
K28748	U.S.S.R.	43	22	32
80	U.S.S.R.	37	28	32
72	England	43	28	36
Dasas	Denmark	43	28	36
S-7845	Canada	48	23	36
K27874	Lithuania	47	27	37
Fylking	Sweden	43	33	38
9	Canada	47	32	39
141	Hungary	43	37	40
83	U.S.S.R.	43	43	43
81	U.S.S.R.	50	42	46
71	Hungary	60	37	48
79	U.S.S.R.	53	47	50
78	U.S.S.R.	57	52	54
Stensballe	Denmark	63	45	54
IV-89-24	Hungary	60	52	56
S-8609	Canada	67	53	60
136	Hungary	70	53	62
85	U.S.S.R.	70	60	65
Prato	Neth.	77	57	66
82	U.S.S.R.	75	63	70
131	U.S.S.R.	77	68	72
77	U.S.S.R.	82	70	76
LSD 1%		47	38	

Table 8. Resistance and recovery of *Festuca rubra* and *Festuca ovina* lines from LTB snow mold, 1974: Test 8 (microplots, 3 replicates)

Cultivar or line	Origin	Percent area of turf affected		
		24 Apr	6 May	Avg.
<i>F. rubra</i>				
S- 1765	U.S.S.R.	2	10	6
Arctared	U.S.A.	30	20	25
Oasis (1108)	Neth.	32	23	28
Boreal (1105)	Neth.	50	33	42
Duraturf	U.S.A.	47	38	43
1084	Canada	58	33	46
Dawson (1133)	England	52	40	46
1120	Canada	53	42	48
Golfrood (1079)	Neth.	53	43	48
Dawson (1104)	England	60	38	49
S59	U.K.	62	38	50
Pennlawn (1087)	U.S.A.	60	43	52
Sceempter	Neth.	67	38	53
Pennlawn (1078)	U.S.A.	68	38	53
1101	U.S.A.	60	48	54
1114	Canada	60	53	57
Polar	U.S.A.	70	43	57
Golfrood (1103)	Neth.	62	53	58
Boreal (1106)	Canada	67	48	58
1108	Neth.	67	48	58
1112	Canada	62	53	58
1102	Canada	70	50	60
1111	Canada	73	52	63
S- 7374	Canada	67	60	63
KL257	Unknown	73	55	64
Fallade	Neth.	72	57	64
Brabantia	Neth.	77	60	68
Ruby	U.S.A.	72	63	68
Reptans (1095)	Sweden	78	61	70
Reptans (1098)	Sweden	82	63	73
Agio	Neth.	80	65	73
Olds (1097)	Canada	77	70	73
Olds (1098)	Canada	70	67	73
1118	France	85	63	74
MSG Flevo-2	Neth.	82	67	74
Novorubra	Neth.	90	67	78
Highlight	Neth.	90	77	83
<i>F. ovina</i>				
S- 1758	U.S.S.R.	2	0	1
S- 1733	U.S.S.R.	2	5	3
S- 1792	U.S.S.R.	3	7	5
Durar	U.S.A.	7	13	10
Biljart	Neth.	27	23	25
Barenza	Neth.	65	65	65
2107	Neth.	70	65	68
LSD 1%		44	49	

- Smith, J. Drew. 1957. The control of certain diseases of sports turf grasses in the British Isles. M.Sc. Thesis, University of Durham. 266 pp.
- Smith, J. Drew. 1973. Overwintering diseases of turfgrasses in Western Canada. Pages 96-103 in Proc. 25th N.W. Turfgrass Assoc. Conf., Harrison Hot Springs, British Columbia. 1973.
- Smith, J. Drew. 1974. Winter diseases of turfgrasses. Pages 20-25 in 25th Annu. Nat. Turfgrass Conf., Roy. Can. Golf Assoc., Winnipeg, Manitoba 1974.

- Smith, J. Drew. 1974. Snow molds of turfgrasses in Saskatchewan. Pages 313-324 in E.C. Roberts, ed., Proc. 2nd Intern. Turfgrass Res. Conf., Blacksburg, Virginia. June 18-21, 1973. Amer. Soc. Agron.
- Ward, E. W. B., and J. B. Lebeau. 1962. Autolytic production of hydrogen cyanide by certain snow mold fungi. Can. J. Bot. 40:85-88.

Ergot tolerance in spring rye

Frank Sosulski¹ and C.C. Bernier²

In field plots in Saskatchewan, spring rye flowered later and showed higher levels of ergot infection than fall rye. Among three spring rye cultivars tested, Gazelle was consistently lower in percentage by weight of ergot sclerotia in seed samples, as well as being substantially higher in seed yield. Floret inoculation tests demonstrated that Gazelle does not possess physiological resistance to ergot. Gazelle showed less infection probably because the florets were pollinated more rapidly and more completely than those of the other cultivars.

Can. Plant Dis. Surv. 55:155-157. 1975

En parcelles d'essais installées en Saskatchewan, le seigle a fleuri plus tard et a été plus gravement touché par l'ergot [*Claviceps purpurea*] que le seigle d'automne (*Secale cereale*). Des trois cultivars testés, c'est Gazelle qui a montré le plus faible pourcentage de sclérotés d'ergot par poids dans les échantillons de grain, ainsi qu'un rendement en grain sensiblement plus élevé. Les épreuves d'inoculation des florules révèlent que Gazelle ne possède pas de résistance physiologique à l'ergot et s'il a été moins atteint, c'est que sa pollinisation a été plus rapide et plus complète que chez les autres cultivars.

Ergot, a common disease of cereals and grasses, is especially prevalent in rye. The causal fungus *Claviceps purpurea* (Fr.) Tul. infects the florets, prevents seeds from developing, and replaces some kernels with a large sclerotium or ergot body (Seaman 1971). In addition, alkaloids in ergot sclerotia may cause poisoning when ingested by animals, poultry, and humans. The concentrations of poisonous alkaloids in ergot sclerotia vary among strains and samples but as little as 0.1% ergot in the feed or diet is considered to be potentially hazardous to health. The Canada Grain Act specifies that the highest grades of rye intended for milling should be free from ergot. Rye containing more than 0.33% ergot sclerotia by weight is graded ergoty.

After the initial infection of florets by ascospores from overwintering sclerotia, the fungus produces conidia, which are spread by rain and insects to later-flowering susceptibles. Therefore, late maturing, naturally cross-pollinated cereals and grasses may show high levels of ergot infection. Spring rye, in particular, flowers later than fall-sown rye and may be severely infected by conidia from neighboring fields of fall rye or perennial grasses. Seed treatment with fungicides does not control ergot, and there are no resistant cultivars of spring or fall rye (Seaman 1971). Therefore, control methods must be designed to limit the availability of spores of the pathogen and to grow crops that flower at the same time and for as short a period as possible.

Platford and Bernier (1970) reported that a cultivar of spring wheat and another of durum wheat possessed

significant degrees of resistance to ergot. Recently a new, high yielding cultivar of spring rye named Gazelle appeared to be less susceptible to ergot infection than other cultivars of spring rye (Sosulski and Curran 1975). The present study was undertaken to determine the level of ergot contamination in seed samples of spring rye cultivars grown in field experiments during 1973 and 1974 in Saskatchewan. These results were compared with the percentages of ergot in seed samples of fall rye cultivars.

Materials and methods

Three spring rye cultivars were grown in replicated yield trials at three locations in 1973 and at four locations in 1974. The entries included Gazelle, a single plant selection from German ryes; S6204, a yellow-seeded genotype developed by Watkins and White (1964); and Prolific, the only commonly grown cultivar of spring rye in western Canada. In addition, five fall rye cultivars were grown in yield trials at Saskatoon in 1973 and 1974.

At harvest, plants from each plot were threshed in the field with a Vogel thresher which has only a minimal cleaning capacity. Loose straw was removed by hand and the seed samples were bagged and weighed. Seed from the four replications in each test was bulked before storage. Ergot sclerotia were separated by hand from the 2 to 3 kg of seed of each cultivar in each test, and the ergot content reported as the weight of sclerotia expressed as a percentage of the total sample weight.

Results and discussion

Fall rye from the 1973 and 1974 replicated field trials had a low incidence of ergot infection. The cultivars Antelope, Cougar, Frontier, Kodiak, and Puma contained between 0.01% and 0.10% ergot in the two experi-

¹ Department of Crop Science, University of Saskatchewan, Saskatoon, Saskatchewan S7N 0W0

² Department of Plant Science, University of Manitoba, Winnipeg, Manitoba R3T 2N2

Table 1. Relationship between percentage of ergot sclerotia and seed yield of spring rye cultivars in experimental plots at four locations in Saskatchewan

Spring rye cultivar	1973			1974				1973—74 Average
	Saskatoon	Indi	Aberdeen	Saskatoon	Indi	Indian Head	Arcola	
Percentage by weight of ergot sclerotia in seed samples								
Gazelle	0.04	0.00	0.01	0.65	0.41	0.57	0.01	0.24
Prolific 1	0.16	0.68	0.21	1.70	0.53	1.23	0.04	0.65
Prolific 2	0.13	0.88	0.16	1.82	0.68	1.66	0.05	0.77
S6204	0.14	0.51	0.33	1.46	0.93	2.10	0.06	0.79
Seed yield, quintals per hectare								
Gazelle	31.4	28.9	30.5	28.4	14.7	25.0	26.9	26.5
Prolific 1	22.6	23.6	24.7	21.1	12.2	25.1	21.6	21.6
Prolific 2	23.6	24.6	24.9	21.0	12.2	19.9	21.4	21.1
S6204	23.0	25.1	24.4	15.8	11.7	12.8	18.3	18.7
LSD 0,05	4.2	2.4	2.4	1.5	2.1	—*	4.0	

* Due to flooding, only one replicate was harvested at Indian Head.

ments. The fall rye cultivars headed during the first week of June and the seed was mature about 1 month later.

The cultivars of spring rye headed after June 20 and ripened 1 to 2 weeks after the fall ryes. The percentages of ergot bodies in the spring rye samples were much greater than for fall rye in six of the seven tests grown in 1973 and 1974 (Table 1). The percentages of ergot were particularly high at Saskatoon, Indi, and Indian Head in 1974 when several samples contained over 1.0% ergot. The Arcola test in 1974 showed a low level of infection in the three cultivars.

In all spring rye tests the percentage of sclerotia in seed samples of Gazelle was lower than in those of the other cultivars (Table 1). The average ergot level in the seed samples of Gazelle was only one-third that of Prolific and S6204. The consistency of the differences in each test suggested that Gazelle contained specific tolerance or resistance to ergot infection of the florets or to the formation of ergot bodies after infection.

However, in subsequent single-floret inoculation tests in the greenhouse, Gazelle was found to be as susceptible to ergot as Stewart 63, a nonresistant cultivar of durum wheat (Platford and Bernier 1970). No differences were observed between the rye and the wheat cultivars in amount of honeydew produced and in number and final size of sclerotia formed. It appears that GAzelle does not possess physiological resistance to ergot.

Gazelle yielded about 24% more seed than Prolific in the 1973-74 series of tests (Table 1) and this factor may have accounted for the lower incidence of ergot sclerotia. Presumably Gazelle florets were pollinated more rapidly and more completely than those of the other cultivars, thus limiting the opportunity for ergot infection. Gazelle headed on the same date as Prolific (avg 48 days after seeding) and the seeds matured in 98 days for both cultivars (Sosulski and Curran 1975), so that a difference in date of flowering was not a factor.

Conclusions

Spring rye cultivars showed higher levels of ergot infection than fall ryes, which was probably due to their later period of flowering. Ergot infection of some spring rye plots exceeded 1.0% because of small plot size and close proximity to roadways sown to perennial grasses. It appears that natural infections in experimental plot areas are sufficient to demonstrate differences in tolerance or resistance to ergot by spring rye cultivars.

Gazelle spring rye was consistently lower in percentage of ergot sclerotia in seed samples from plots grown at seven locations in 1973 and 1974. Single floret inoculation tests demonstrated that Gazelle lacked physiological resistance to ergot. The lower incidence of ergot observed in this cultivar in the field was apparently due to a better flowering habit which allowed Gazelle to escape infection to a greater degree than other spring cultivars.

Acknowledgments

The authors wish to thank Mr. R. N. McIver and Dr. D. B. Fowler for providing seed samples, and Miss P. G. Hoffman for technical assistance.

Literature cited

- Platform, R. G., and C. C. Bernier. 1970. Resistance to *Claviceps purpurea* in spring and durum wheat. *Nature* 226:770.
- Seaman, W. L. 1971. Ergot of grains and grasses. Agriculture Canada Pub. 1438. 12 pp.
- Sosulski, F. W., and W. A. Curran. 1975. Gazelle spring rye. *Can. J. Plant Sci.* 55: 629.
- Watkins, R., and W. J. White. 1964. The inheritance of anthocyanin in rye (*Secale cereale* L.). *Can. J. Genet. Cytol.* 6: 403-410.

Author index to volume 55

- BARR, D. J. S., and W. G. KEMP. *Olpidium brassicae*, tobacco necrosis virus, and *Pythium* spp. in relation to rusty root of carrots in Ontario and Quebec 77
- BERKENKAMP, B., and J. MEERES. Observations on silvertop of grasses in Alberta 83
- BERNIER, C. C. (see Platford, R. G. & C. C. Bernier) 75
- BERNIER, C. C. (see Sosulski, Frank & C. C. Bernier) 155
- CHYKOWSKI, L. N., and D. L. CRAIG. Reaction of strawberry cultivars to clover phyllody (green petal) agent transmitted by *Aphrodes bicincta* 66
- CLARK, R. V., C. O. GOURLEY, H. W. JOHNSTON, L. J. PIENING, G. PELLETIER, J. SANTERRE, and H. GENEUREUX. Oat yield losses from septoria leaf blotch at four locations in eastern Canada 36
- CRAIG, D. L. (see Chiykowski, L. N.) 66
- DAVIDSON, J. G. N. (see McKenzie, J. S., & J. G. N. Davidson) 121
- DYCK, P. L. (see Green, G. J., & P. L. Dyck) 85
- FUSHTEY, S. G. The nature and control of snow mold of fine turfgrass in southern Ontario 87
- FUSHTEY, S. G., and C. B. KELLY. A new record of stem and bulb nematode in Ontario 27
- GAGNON, C. (see Richard, C.) 45
- GALWAY, D. A. (see Wallen, V. R., & D. A. Galway) 73
- GAYED, S. K., and M. C. WATSON. Diseases of flue-cured tobacco in Ontario and estimates of disease losses, 1972-73 31
- GENEUREUX, H. (see Clark, R. V., et al.) 36
- GOURLEY, C. O. (see Clark, R. V., et al.) 36
- GREEN, G. J. Air-borne rust inoculum over western Canada in 1974 48
- GREEN, G. J. Stem rust of wheat, barley, and rye in Canada in 1974 51
- GREEN, G. J., and P. L. DYCK. The reaction of Thatcher wheat to Canadian races of stem rust 85
- HARDER, D. E. Crown rust of oats in Canada in 1974 63
- JARVIS, W. R., and K. SLINGSBY. Tolerance of *Botrytis cinerea* and rose powdery mildew to benomyl 44
- JARVIS, W. R., H. J. THORPE, and B. H. MACNEILL. A foot and root rot disease of tomato caused by *Fusarium oxysporum* 25
- JOHNSTON, G. R. (see Rowberry, R. G., & G. R. Johnston) 15
- JOHNSTON, H. W. (see Clark, R. V., et al.) 36
- KELLY, C. B. (see Fushthey, S. G., & C. B. Kelly) 27
- KEMP, W. G. (see Barr, D. J. S., & W. G. Kemp) 77
- LOCKHART, C. L. Effect of temperature on the development of *Godronia cassandrae* f. *vaccinii* cankers on lowbush blueberry 29
- MACNEILL, B. H. (see Jarvis, W. R., et al.) 25
- MARTENS, J. W. Stem rust of oats in Canada in 1974 61
- MCINTOSH, D. L. Proceedings of the 1974 APDW workshop on crown rot of apple trees 109
- MCKEEN, W. E. *Pythium* root rot of barley in southwestern Ontario 12
- MCKENZIE, D. L., and R. A. A. MORRALL. Fababean diseases in Saskatchewan in 1973 1
- MCKENZIE, D. L., and R. A. A. MORRALL. Diseases of specialty crops in Saskatchewan: II. Notes on field pea in 1973-74 and on lentil in 1973 97
- MCKENZIE, D. L. (see Morrall, R. A. A. & D. L. McKenzie) 69
- MCKENZIE, J. S., and J. G. N. DAVIDSON. Prevalence of alfalfa crown and root diseases in the Peace River region of Alberta and British Columbia 121
- MEERES, J. (see Berkenkamp, B. & J. Meeres) 83
- MILLS, J. T. Cooperative seed treatment trials - 1974 8
- MILLS, J. T. Cooperative seed treatment trials - 1975 117
- MISKA, J. P., and G. A. NELSON. Potato seed-piece decay: a bibliography, 1930-1975 126
- MORRALL, R. A. A., and D. L. MCKENZIE. Diseases of specialty crops in Saskatchewan: 1. Notes on buckwheat and sunflower, 1972-73 69
- MORRALL, R. A. A. (see McKenzie, D. L., & R. A. A. Morrall) 1
- MORRALL, R. A. A. (see McKenzie, D. L., & R. A. A. Morrall) 97
- MURANT, A. F. Occurrence of mottle and redleaf components of carrot motley dwarf disease in British Columbia 103
- NELSON, G. A. (see Miska, J. P., & G. A. Nelson) 126
- PELLETIER, G. (see Clark, R. V., et al.) 36
- PETRIE, G. ALLAN. Prevalence of oospores of *Albugo cruciferarum* in *Brassica* seed samples from western Canada, 1967-73 19
- PIENING, L. J. (see Clark, R. V., et al.) 36
- PLATFORD, R. G., and C. C. BERNIER. Diseases of rapeseed in Manitoba, 1973-74 75
- RICHARD, C., et C. GAGNON. Pourridié fusarien et maladies du feuillage chez la luzerne au Québec en 1974 45
- ROWBERRY, R. G., and G. R. JOHNSTON. Virus infection of potato seed stocks in Ontario under commercial insect-control practices 15
- SAMBORSKI, D. J. Leaf rust of wheat in Canada in 1974 58
- SANTERRE, J. (see Clark, R. V., et al.) 36
- SLINGSBY, K. (see Jarvis, W. R., & K. Slingsby) 44
- SLYKHUIS, J. T. Effects of drying on the transmissibility of wheat spindle streak mosaic virus in soils from wheat fields in Ontario 106
- SMITH, J. DREW. Snow molds on winter cereals in northern Saskatchewan in 1974 91
- SMITH, J. DREW. Resistance of turfgrasses to low-temperature-basidiomycete snow mold and recovery from damage 147
- SOSULSKI, FRANK, and C. C. BERNIER. Ergot tolerance in spring rye 155
- THORPE, H. J. (see Jarvis, W. R., et al.) 25
- VAARTAJA, O. *Pythium sylvaticum* in Canadian forest nurseries 101
- WALLEN, V. R., and D. A. GALWAY. Incidence of bacterial blight of field beans in southwestern Ontario in 1973 and 1974 73
- WATSON, M. C. (see Gayed, S. K.) 31

Recommandations aux auteurs

Les articles et les communiqués sont publiés en anglais ou en français. Les manuscrits (l'original et une copie) et toute la correspondance qui s'y rapporte doivent être envoyées au Rédacteur M. W. L. Seaman, à la Station de recherches d'Ottawa, ministère de l'Agriculture du Canada, Ottawa (Ontario) K1A 0C6.

Les manuscrits doivent être concis et faire preuve de suite dans le style, l'orthographe et l'emploi des abréviations. Ils doivent être dactylographiés à double interligne, de préférence sur des feuilles à lignes numérotées. Toutes les pages doivent être numérotées y compris celles portant le résumé, les tableaux et les légendes. Pour plus de renseignements sur le format des feuilles et le style, prière de consulter nos dernières publications et le *CBE Style Manual* (3e ed. 1972) de l'American Institute of Biological Sciences, Washington (DC). Dans la mesure du possible, les données numériques doivent être exprimées en unités métrique, (SI) ou être suivies de leur équivalent métrique. L'emploi de crochets est autorisé pour l'identification du nom scientifique d'un micro-organisme pathogène après le nom commun de la maladie dont il est l'agent causal.

Les titres doivent être courts et révélateurs en contenant, avec le résumé, les mots clés les plus utiles pour le classement et l'extraction de l'information.

Chaque article doit être accompagné d'un *résumé* d'au plus 200 mots en anglais et en français, si possible.

Les figures doivent pouvoir, après réduction, remplir une colonne (maximum 84 x 241 mm) ou deux colonnes (maximum 175 x 241 mm) et devraient être taillées ou montrer les parties essentielles à garder. Les figures groupées sur une même planche doivent être montées côte à côte, sans intervalle. L'article doit être accompagné d'un double des photographies non montées et des graphiques. Les figures doivent être numérotées, porter le nom de l'auteur et une légende abrégée.

Les tableaux doivent être numérotés en chiffres arabes et avoir un titre concis. Ils ne devraient pas avoir de lignes verticales. Les renvois doivent être identifiés par un signe typographique particulier (* † § # ¶ ** ††) surtout lorsqu'il s'agit de nombres.

Les références bibliographiques devraient être citées par ordre alphabétique comme dans les livraisons courantes. On peut utiliser le système de numération ou le système nom-et-année. Pour l'abrégé du titre des périodiques, on suivra l'édition la plus récente de *Biosis List of Serials* publiée par les Biosciences Information Services de Biological Abstracts ou la *NCPTWA Word Abbreviation List* et l'American National Standards Institute, Standards Committee Z39.

Instructions to authors

Articles and brief notes are published in English or French. Manuscripts (original and one copy) and all correspondence should be addressed to the Editor, Dr. W. L. Seaman, Ottawa Research Station, Agriculture Canada, Ottawa, Ontario K1A 0C6.

Manuscripts should be concise and consistent in style, spelling, and use of abbreviations. They should be typed, double spaced throughout, on line-numbered paper. All pages should be numbered, including those containing abstract, tables, and legends. For general format and style, refer to recent issues of the *Survey* and to *CBE Style Manual*, 3rd ed. 1972. American Institute of Biological Sciences, Washington, D.C. Whenever possible, numerical data should be in metric units (SI) or metric equivalents should be included. Square brackets may be used to enclose the scientific name of a pathogen, following the common name of a disease, to denote cause.

Titles should be concise and informative providing, with the Abstract, the key words most useful for indexing and information retrieval.

Abstracts of no more than 200 words, in both English and French, if possible, should accompany each article.

Figures should be planned to fit, after reduction, one column (maximum 84 X 241 mm) or two columns (maximum 175 X 241 mm), and should be trimmed or marked with crop marks to show only essential features. Figures grouped in a plate should be butt-mounted with no space between them. A duplicate set of unmounted photographs and line drawings is required. Figures should be identified by number, author's name, and abbreviated legend.

Tables should be numbered using arabic numerals and have a concise title; they should not contain vertical rules; footnotes should be identified by reference marks (* † § # ¶ ** ††), particularly when referring to numbers.

Literature cited should be listed alphabetically in the form appearing in current issues; either the number system or the name-and-year system may be used. For the abbreviated form of titles of periodicals, refer to the most recent issue of *Biosis List of Serials* published by Biosciences Information Service of Biological Abstracts or to the *NCPTWA Word Abbreviation List*, American National Standards Institute.